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#### **PCT**

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#### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6: C12N 15/12, C07K 14/47, C12N 5/10, A61K 48/00, G01N 33/68, C12Q 1/68, C07K 16/18

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(71) Applicants (for all designated States except US): MEDICAL RESEARCH COUNCIL [GB/GB]; 20 Park Crescent, London WIN 4AL (GB). LEIDEN UNIVERSITY [NL/NL]; P.O. Box 9500, NL-2300 RA Leiden (NL). UNIVERSITY OF WALES COLLEGE OF MEDICINE [GB/GB]; Heath Park, Cardiff CF4 4XN (GB). ERASMUS UNIVERSITY ROTTERDAM [NL/NL]; Burg Ondlaan 50, P.O. Box 1738, NL-3000 DR Rotterdam (NL).

(72) Inventors; and

(75) Inventors/Applicants (for US only): HARRIS, Peter, Charles [GB/GB]; 65 Freelands Road, Oxford OX4 4BS (GB). PERAL, Belen [ES/GB]; 77 Lock Crescent, Kidlington, Oxford OX5 1HF (GB). WARD, Christopher, James [GB/GB]; 30 Benson Road, Oxford OX3 7EH (GB). HUGHES, James [GB/GB]; 225 Cowley Road, Oxford OX4 1XD (GB). BREUNING, Martin, Hendrik [NL/NL]; Brigantijnstraat 57, NL-1503 BR Zaandam (NL). PETERS, Dorothea, Johanna, Maria [NL/NL]; Zuster Meijboomstraat 267, NL-2331 PH Leiden (NL). ROELFSEMA, Jeroen, Hendrik [NL/NL];

Vijf Meilaan 2006, NL-2321 RR Leiden (NL). SAMP-SON, Julian [GB/GB]; 34 Bridge Street, Cardiff CF5 2EL (GB). HALLEY, Dirkje, Jorijntje, Johanna [NL/NL]; Van Aerssenlaan 35 d, NL-3039 KD Rotterdam (NL). NEL-LIST, Mark, David [GB/NL]; Noordmolenstraat 57b, NL-3053 RG Rotterdam (NL). JANSSEN, Lambertus, Antonius, Jacobus [NL/NL]; Schokker 37, NL-2991 DJ Barendrecht (NL). HESSELING, Arjenne, Ligue, Wilhelma [NL/NL]; Haya van Someren Downerpad 7, NL-3207 DK Spijkenisse

- (74) Agents: NEWELL, William, Joseph et al.; Wynne-Jones, Laine & James, 22 Rodney Road, Cheltenham, Gloucestershire GL50 1JJ (GB).
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#### Published

With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

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#### (54) Title: POLYCYSTIC KIDNEY DISEASE 1 GENE AND USES THEREOF

#### (57) Abstract

The present invention relates to the polycystic kidney disease 1 (PKD1) gene and its nucleic acid sequence, mutations thereof in patients having PKD1-associated disorders, the protein encoded by the PKD1 gene or its mutants, and their uses in disease diagnosis and therapy.

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A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/12 C07K14 A61K48/00 G01N33/68 CO7K14/47 C12N5/10 C07K16/18 C12Q1/68 According to International Patent Classification (IPC) or to both national classification and IPC Minimum documentation searched (classification system followed by classification symbols) C12N A61K C12Q C07K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages 1-4,6-30 X J. AM. SOC. NEPHROL., vol. 4, no. 3, page 814 G. GERMINO ET AL 'A novel approach to the identification of the PKD1 gene' 31-40 Y see abstract 91p 1-40 Υ KIDNEY INTERNATIONAL, vol. 43, no. supp 3, 19 May 1993 pages s20-s25, G. GERMINO ET AL 'Positional cloning approach to the dominant polycystic kidney disease gene, PKD1' see the whole document X Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents: "T" later document published after the international fiting date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled "O" document referring to an oral disclosure, use, exhibition or other means in the art. document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 05.12.1995 28 November 1995 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016 Van der Schaal, C

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Inter nal Application No
PCT/GB 95/01386

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mational application No.

PCT/GB95/01386

#### INTERNATIONAL SEARCH REPORT

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This int	ernational search report has not been established in respect of certain claims under Article 17(2)(2) for the following reasons:
ı. 🛛	Claims Nos.: 31-33 because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claims 31-33 are directed to a method of treatment of the human body, the search has been carried out and based on the alleged effect of the compound.
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet).
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:
•	
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ,	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remari	on Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.

Information on patent family members

Int ional Application No PCT/GR 95/01386

Information on patent family members		PCT/GB	95/01386		
Patent document cited in search report	Publication date	Patent far member	nily (s)	Publication date	
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## PATENT & TRADEMARK OFFICE DEPOSIT ACCOUNT CHARGE(S)

Patent No.: <u>09/548,882</u> Date of Charge: December 18, 2002

**Client Matter No. 23893-7083** 

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Issue Fee	<u>\$</u>
Notice of Appeal (Small Entity)	<u>\$</u>
multiple dependent	<u>\$</u>
total claims	<u>\$</u>
Additional Claims independent	<u>\$</u>
Petition for Extension of Time: (Small Entity)  1 2 3 4 5 Months	<u>\$</u>
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Surcharge-Late Filing of Declaration	<u>\$</u>
Filing Fee for filing Provisional Application	<u>\$</u>

amount indicated.

Approved By	:	January 6, 2002
	(Sign Name)	Date
	Antoinette Konski	7491
	(Print Name)	Attorney Numbe

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CMS EVENT SERVER

Sent:

Wednesday, January 01, 2003 10:01 AM

To:

Nichols, Peggy .

Subject:

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Client:

Sidney Kimmel Cancer Center

Matter: '

US-Methods for Classifying Tumors Accord

Invoice #:

508957

Invoice Date:

9/30/2002

Invoice Amount:

\$26,130.33\_\_\_

Payment Received From: Bingham McCutchen trustee

Payment Date: 12/31/2002 Payment Amount: \$15,000.00

Payment Amount:

\$15,000.00

Invoice Balance, all Payors: \$11,130.33

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#### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(71) Applicants (for all designated States except US): MEDICAL RESEARCH COUNCIL [GB/GB]; 20 Park Crescent, London W1N 4AL (GB). LEIDEN UNIVERSITY [NL/NL]; P.O. Box 9500, NL-2300 RA Leiden (NL). UNIVERSITY OF WALES COLLEGE OF MEDICINE [GB/GB]; Heath Park, Cardiff CF4 4XN (GB). ERASMUS UNIVERSITY ROTTERDAM [NL/NL]; Burg Ondlaan 50, P.O. Box 1738, NL-3000 DR Rotterdam (NL).

(72) Inventors; and

(75) Inventors/Applicants (for US only): HARRIS, Peter, Charles [GB/GB]; 65 Freelands Road, Oxford OX4 4BS (GB). PERAL, Belen [ES/GB]; 77 Lock Crescent, Kidlington, Oxford OX5 1HF (GB). WARD, Christopher, James [GB/GB]; 30 Benson Road, Oxford OX3 7EH (GB). HUGHES, James

[GB/GB]; 225 Cowley Road, Oxford OX4 1XD (GB). BREUNING, Martin, Hendrik [NL/NL]; Brigantijnstraat 57, NL-1503 BR Zaandam (NL). PETERS, Dorothea, Johanna, Maria [NL/NL]; Zuster Meijboomstraat 267, NL-2331 PH Leiden (NL). ROELFSEMA, Jeroen, Hendrik [NL/NL]; Vijf Meilaan 2006, NL-2321 RR Leiden (NL). SAMP-SON, Julian [GB/GB]; 34 Bridge Street, Cardiff CF5 2EL (GB). HALLEY, Dirkje, Jorijntje, Johanna [NL/NL]; Van Aerssenlaan 35 d, NL-3039 KD Rotterdam (NL). NEL-LIST, Mark, David [GB/NL]; Noordmolenstraat 57b, NL-3053 RG Rotterdam (NL). JANSSEN, Lambertus, Antonius, Jacobus [NL/NL]; Schokker 37, NL-2991 DJ Barendrecht (NL). HESSELING, Arjenne, Ligue, Wilhelma [NL/NL]; Haya van Someren Downerpad 7, NL-3207 DK Spijkenisse (NL).

- (74) Agents: NEWELL, William, Joseph et al.; Wynne-Jones, Laine & James, 22 Rodney Road, Cheltenham, Gloucestershire GL50 1JJ (GB).
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#### (57) Abstract

The present invention relates to the polycystic kidney disease 1 (PKD1) gene and its nucleic acid sequence, mutations thereof in patients having PKD1-associated disorders, the protein encoded by the PKD1 gene or its mutants, and their uses in disease diagnosis and therapy.

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# POLYCYSTIC KIDNEY DISEASE 1 GENE AND USES THEREOF BACKGROUND TO THE INVENTION

In humans, one of the commonest of all genetic disorders is autosomal dominant polycystic kidney disease (ADPKD) also termed adult polycystic kidney disease (APKD), affecting approximately 1/1000 individuals (Dalgaard, 1957). ADPKD is a progressive disease of cyst formation and enlargement typically leading to end stage renal disease (ESRD) in late middle age. The major cause of morbidity in ADPKD is progressive renal disease characterized by the formation and enlargement of fluid filled cysts, resulting in grossly enlarged kidneys. Renal function deteriorates as normal tissue is compromised by cystic growth, resulting in end stage renal disease (ESRD) in more than 50% of patients by the age of 60 years (Gabow, et al., 1992). ADPKD accounts for 8-10% of all renal transplantation and dialysis patients in Europe and the USA (Gabow, 1993).

ADPKD also causes cystic growth in other organs (reviewed in Gabow, 1990) and occasionally presents in childhood (Fink, et al., 1993; Zerres, et al., 1993). Extrarenal manifestations include liver cysts (Milutinovic, et al., 1980), and more rarely cysts of the pancreas (Gabow, 1993) and other organs. Intracranial aneurysms occur in approximately 5% of patients and are a significant cause of morbidity and mortality due to subarachnoid haemorrhage (Chapman, et al., 1992). ADPKD is associated with a higher prevalence of various connective tissue disorders. An increased prevalence of heart valve defects (Hossack, et

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al., 1988), hernia (Gabow, 1990) and colonic diverticulae (Scheff, et al., 1980) have been reported.

Considerable progress has been made in the last few years in understanding the pathophysiology of ADPKD (and other animal models of cystic disease). Cysts in ADPKD are develop from outpouchings of descending or ascending kidney tubules and the early stages characterized by a thickening and disorganization of the basement membrane; accompanied by a de-differentiation of tubular epithelial cells. Several of the characteristics of  ${\mathbb R}^2$ epithelia: altered growth responses, abnormal expression of various proteins and reversal of polarity, may be a sign of this de-differentiation and important in cyst expansion. The nature of the primary defect which triggers 15 these changes is, however, unknown and consequently much 🍮 effort has been devoted to identifying the causative agent by genetic means. . . . . .....

The first step towards positional cloning of an ADPKD gene was the demonstration of linkage of one locus now designated the polycystic kidney disease 1 (PKD1) locus to the  $\alpha$  globin cluster on the short arm of chromosome 16 (Reeders, et al., 1985). Subsequently, families with ADPKD - unlinked to markers of 16p were described (Kimberling, et al., 1988; Romeo, et al., 1988) and a second ADPKD locus (PKD2) has recently been assigned to chromosome region 4g13q23 (Kimberling, et al., 1993; Peter, et al., 1993). It is estimated that approximately 85% of ADPKD is due to PKD1 (Peters and Sankuijl, 1992) with PKD2 accounting for most of

the remainder. PKD2 appears to be milder condition with a later age of onset and ESRD (Parfrey et al., 1990; Gabow, et al., 1992; Ravine, et al., 1992).

The position of the PKD1 locus was refined to chromosome band 16p13.3 and many markers were isolated from that region (Breuning, et al., 1987; Reeders, et al., 1988; Breuning, et al., 1990; Germino, et al., 1990; Hyland, et al., 1990: Himmelbauer, et al., 1991). Their order, and the position of the PKD1 locus, has been determined by extensive 10 linkage analysis in normal and PKD1 families and by the use of a panel of somatic cell hybrids (Reeders et al., 1988; Breuning, et al., 1990; Germino, et al., 1990). genetically heterogenous with loci mapped not only to 16pl3.3 (PKD1), but also to chromosome 4 (PKD2). Although the phenotype of PKD1 and PKD2 are clearly similar, it is now well documented that PKD1 (which accounts for about 85% of ADPKD; (Peters, 1992) is a more severe disease with an average age at ESRD of about 56 years compared to about 71.5 years for PKD2 (Ravine, 1992). An accurate long range 20 restriction map of the 16p13.3 region (Harris, et al., 1990; Germino, et al., 1992) has located the PKD1 locus in an interval of approximately 600 kb between the markers GGG1 and SM7 (Harris, et al., 1991; Somlo, et al., 1992) (see Figure la). The density of CpG islands and identification 25 of many mRNA transcripts indicated that this area is rich in gene sequences. Germino et al. (1992) estimated that the candidate region contains approximately 20 genes.

Identification of the PKDl gene from within this area

has thus proved difficult and other means to pinpoint the disease gene have been sought. Linkage disequilibrium has been demonstrated between PKD1 and the proximal marker VK5, in a Scottish population (Pound, et al., 1992) and between PKD1 and BLu24 (see Figure 1a), in a Spanish population (Peral, et al., 1994). Studies with additional markers have shown evidence of a common ancestor in a proportion of each population (Peral, et al., 1994; Snarey, et al., 1994), but the association has not precisely positioned the PKD1 locus.

Disease associated genomic rearrangements, detected by -10 cytogenetics or pulsed field gel electrophoresis (PFGE) have been instrumental in the identification of various genes associated with various genetic disorders. Hitherto, no 🛶 such abnormalities related to PKD1 have been described. This situation contrasts with that for the tuberous sclerosis locus, which lies within 16p13.3 (TSC2). In that case, TSC associated deletions were detected by PFGE within the interval thought to contain the PKD1 gene and their characterisation was a significant step toward the rapid identification of the TSC2 gene (European Chromosome 16 20 Tuberous Sclerosis Consortium, 1993). The TSC2 gene therefore maps within the candidate region for the hitherto unidentified PKD1 gene; as polycystic kidneys are a feature common to TSC and ADPKD1 (Bernstein and Robbins, 1991) the possibility of an etiological link, as proposed by Kandt et 25 al. (1992), was considered. A contiguous gene syndrome resulting from the disruption of PKD1 and the adjacent tuberous sclerosis 2 (TSC2) gene, which is associated with

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TSC and severe childhood onset polycystic kidney disease, has also been defined (Brook-Carter et al, 1994).

We have now identified a pedigree in which the two distinct phenotypes, typical ADPKD or TSC, are seen in different members. In this family, the two individuals with ADPKD are carriers of a balanced chromosome translocation with a breakpoint within 16p13.3. We have located the chromosome 16 translocation breakpoint and a gene disrupted by this rearrangement has been defined; the discovery of additional mutations of that gene in other PKD1 patients shows that we have identified the PKD1 gene. characterisation of the PKD1 transcript significantly complicated because of the unusual genomic region containing most of the gene. All but 3.5 kb at the 3' end of the transcript (which is about 14 kb in total) is encoded by a region which is reiterated several times elsewhere on the same chromosome (in 16p13.1 and termed the HG area). The structure of the duplication is complex, with some regions copied more times than others, and the HG region encoding three large transcripts. The transcripts. from the HG area are: HG-A (21 kb), HG-B (17 kb) and HG-C (8.5 kb) and although these have 3' ends which differ from PKD1, over most of their length they share substantial homology to the PKD1 transcript. Consequently, cloning and characterizing a bona fide PKD1 cDNA has proven difficult. To overcome the problem caused by duplication we have cloned cDNAs covering the entire transcript from a cell line which contains the PKD1 but not the HG loci. Characterisation of

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these cDNAs has enabled the PKD1 protein sequence to be predicted and led to the identification of several homologies with described motifs.

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#### SUMMARY OF THE INVENTION

Accordingly, in one aspect, this invention provides an isolated, purified or recombinant nucleic acid sequence comprising:

- 5 (a) a PKD1-encoding nucleic acid or its complementary strand,
  - (b) a sequence substantially homologous to, or capable of hybridizing to, a substantial portion of a molecule defined in (a) above, or ;
- 10 (c) a fragment of a molecule defined in (a) or (b) above.

In particular, there is provided a sequence wherein the PKD1 gene has the nucleic acid sequence according to Fig. 15, or the partial sequence of Figs. 7 or 10. The invention therefore includes a DNA molecule coding for a polypeptide having the amino acid sequence of Figure 15, or a polypeptide fragment thereof; and genomic DNA corresponding to a molecule as in (a) - (c) above.

As used herein, "substantially homologous" refers to a nucleic acid strand that is sufficiently duplicative of the PKD1 sequence presented in Fig. 15 such that it is capable of hybridizing to that sequence under moderately stringent, and preferably stringent conditions, as defined herein below. Preferably, "substantially homologous" refers to a homology of between 97 and 100%. Further, such a strand will encode or be complementary to a strand that encodes PKD1 protein having the biological activity described below. As used herein, a "substantial portion of a molecule" refers

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to at least 60%, preferably 80% and most preferably 90% of the molecule in terms of its linear residue length or its molecular weight. "Nucleic acid" refers to both DNA and RNA.

The PKD1 gene described herein is a gene found on human chromosome 16, and the results of studies described herein form the basis for concluding that this PKD1 gene encodes a protein called PKD1 protein which has a role in the prevention or suppression of ADPKD. The PKD1 gene therefore includes the DNA sequences shown in Figure 15, and all functional equivalents. By "functional equivalents"; we mean nucleic acid sequences that are substantially homologous to the PKD1 nucleic acid sequence, as presented to in Fig. 15; and encoding a protein that possesses one or 15 more of the biological functions or activities of PKD1: , i.e., that is involved in cell/cell adhesion, cell/cell recognition or cell/cell communication; for example to effect adhesion of cells to other cells or components of the extracellular - matrix; effect communication interaction between epithelial cells and the basal membrane (whether in kidneys or otherwise); assist in development of connective tissue such as assembly and/or maintenance of the basal membrane; in signal transduction between cells or cells and components of the extracellular matrix; and/or to promote binding of cells carrying proteins such as integrins or carbohydrates to target cells. The biological function PKD1 of course includes maintaining a physiological state; that is, the native protein's

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aberrations or absence results in ADPKD or an associated disorder.

The PKD1 gene may furthermore include regulatory regions which control the expression of the PKD1 coding sequence, including promoter, enhancer and terminator regions. Other DNA sequences such as introns spliced from the end-product PKD1 RNA transcript are also encompassed. Although work has been carried out in relation to the human gene, the corresponding genetic and functional sequences present in lower animals are also encompassed.

The present invention therefore further provides a PKD1 gene or its complementary strand having the sequence according to Figure 15 which gene or strand is mutated in some ADPKD patients (more specifically, PKD1 patients). Therefore, the invention further provides a nucleic acid sequence comprising a mutant PKD1 gene as described herein, including wherein Intron 43 as defined hereinbelow has a deletion of 18 or 20bp resulting in an intron of 55 or 57bp.

As used herein, "PKD1 mutant" or "mutation" encompasses alterations of the native PKD1 nucleotide or amino acid sequence, as defined by Fig. 15, i.e., substitutions, deletions or additions, and also encompasses deletion of DNA containing the entire PKD1 gene.

The invention further provides a nucleic acid sequence

25 comprising a mutant PKDl gene, especially one selected from
a sequence comprising a partial sequence according to
Figures 7 and/or 10, or the corresponding sequences
disclosed in Fig. 15, when:

	(a) [OX114] base pairs 1746-2192 as defined in Figure
	7 are deleted (446bp);
	(b) [OX32] base pairs 3696-3831 as defined in Figure
-	7 are deleted by a splicing defect;
5	(c) [OX875] about 5.5kb flanked by the two Xbal sites
	shown in Figure 3a are deleted and the EcoRl site separating
	the CW10 (41kb) and JH1 (18kb) sites is thereby absent
	(d) [WS53] about 100kb extending between the JH1 and
	CW21 and the SM6 and JH17 sites shown in Figure 6 and the
10	PKD1 gene is thereby absent, the deletion lying proximally
	between SM6 and JH13;
	(e)[461] 18bp are deleted in the 75bp intron
1.	amplified by the primer pair 3A3C insert at position 3696 of
	the 3' sequence as shown in Figure 11;
15	(f) [OX1054] 20bp. are deleted in the 75bp intron
1	amplified by the primer pair 3A3C insert at position 3696 of
•	the 3' sequence as shown in Figure 11:
	(g) [WS212] about 75kb are deleted between SM9-CW9
	distally and the PKD1 3'UTR proximally as shown in Figure
20	12;
	(h) [WS-215] about 160kb are deleted between CW20 and
	SM6-JH17 as shown in Figure 12;
	(i) [WS-227] about 50kb are deleted between CW20 and
	JH11 as shown in Figure 12;
25	(j) [WS-219] about 27kb are deleted between JH1 and
	JH6 as shown in Figure 12;
	(k) [WS-250] about 160kb are deleted between CW20 and
	Blu24 as shown in Figure 12;

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(1) [WS-194] about 65kb is deleted between CW20 and CW10.

The invention therefore extends to RNA molecules comprising an RNA sequence corresponding to any of the DNA sequences set out above. Such molecule may be the transcript reference PBP and identifiable with respect to the restriction map of Figure 3a and having a length of about 14 KB.

In another aspect, the invention provides a nucleic acid probe having a sequence as set out above; in particular, this invention extends to a purified nucleic acid probe which hybridizes to at least a portion of the DNA or RNA molecule of any of the preceding sequences. Preferably, the probe includes a label such as a radiolable, for example, a <sup>32</sup>P label.

In another aspect, this invention provides a purified DNA or RNA coding for a protein comprising the amino acid sequence of Figure 15, or a protein polypeptide having homologous properties with said protein, or having at least one functional domain or active site in common with said protein.

The DNA molecule defined above may be incorporated in a recombinant cloning vector for expressing a protein having the amino acid sequence of Figure 15, or a protein or a polypeptide having at least one functional domain or active site in common with said protein. Such a vector may include any vector for expression in bacteria, e.g., E. coli; yeast, insect, or mammalian cells.

The invention also features a nucleic acid probe for detecting PKD1 nucleic acid comprising 10 consecutive nucleotides as presented in Fig. 15. Preferably, the probe may comprise 15, 20, 50, 100, 200, or 300, etc., consecutive nucleotides (nt) presented in Fig. 13, and may fall within the size range 15nt-13kb, 100nt-5kb, 150nt-4kb, 300nt-2kb, and 500nt-1kb.

Probes are used according to the invention in hybridization reactions to identify PKD1 sequences, whether they be native or mutated PKD1 DNA or RNA, as disclosed herein. Such probes are useful for identifying the PKD1 gene or a mutation thereof, as defined herein.

The invention also features a synthetic polypeptide corresponding in amino acid residue sequence to at least a portion of the sequence of naturally occurring PKD1, and having a molecular weight equal to less than that of the native protein. A synthetic polypeptide of the invention is useful for inducing the production of antibodies specific for the synthetic polypeptide and that bind to naturally occurring PKD1.

Preferred embodiments of this aspect of the invention include a group of synthetic polypeptides whose members correspond to a fragment of the PKD1 protein comprising a stretch of amino acids of at least 8, and preferably 15, 30, 50, or 100 residues in length from the sequence disclosed in Fig. 15.

In another aspect, the invention provides a polypeptide encoded by a sequence as set out above; or having the amino

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acid sequence according to the amino acid sequence of Figure 15, or a protein or polypeptide having homologous properties with said protein, or having at least one functional domain or active site in common with said protein. In particular, there is provided an isolated, purified or recombinant polypeptide comprising a PKD1 protein or a mutant or variant thereof or encoded by a sequence set out above or a variant thereof having substantially the same activity as the PKD1 The present invention may further comprise a protein. polypeptide having 9 or 13 transmembrane pairs instead of 11 transmembrane domains as described hereinbelow. Further comprising this invention is a molecule which interacts with a polypeptide as herein described which molecule synergises, causes, enhances or is necessary for the functioning of the PKD1 protein as herein described.

The invention also encompasses recombinant expression vectors comprising a nucleic acid or isolated DNA encoding PKD1 and a process for preparing PKD1 polypeptide, comprising culturing a suitable host cell comprising the vector under conditions suitable for promoting expression of PKD1, and recovering said PKD1.

This invention also provides an <u>in vitro</u> method of determining whether an individual is at risk of a PKD1-associated disorder, comprising assaying a biological sample from the individual to determine the presence and/or amount of PKD1 protein or polypeptide having the amino acid sequence of Figure 15.

As used herein, "biological sample" includes any fluid

or tissue sample from a mammal, preferably a human, including but not limited to blood, urine, saliva, any body organ tissue, cells from any body tissue, including blood cells.

- 5 Additionally or alternatively, a sample may be assayed to determine the presence and/or amount of mRNA coding for the protein or polypeptide having the amino acid sequence of "Figure 15, or to determine the fragment lengths of fragments nucleotide sequences coding for the protein or polypeptide of Figure 15, or to detect inactivating mutations in DNA coding for a protein having the amino acid sequence of Figure 15 or a protein having homologous The screening preferably includes applying a nucleic acid amplification process, as described herein in 15 detail, to said sample to amplify a fragment of the DNA sequence. The nucleic acid amplification advantageously utilizes at least one of the following sets of primers as identified herein: AH3 F9 : AH3 B7; 3A3 C1 : 3A3 C2; and AH4 F2 : JH14 B3.
- Alternatively, the screening method may comprise digesting the sample DNA to provide ECORI fragments and hybridizing with a DNA probe which hybridizes to the EcoRI fragment identified (A) in Figure 3(a), and the DNA probe may comprise the DNA probe CW10 identified herein.
- Another screening method may comprise digesting the sample to provide BamHI fragments and hybridizing with a DNA probe which hybridizes to the BamHI fragment identified (B) in Figure 3(a), and the DNA probe may comprise the DNA probe

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1AlH.6 identified herein.

A method according to the present invention may comprise detecting a PKD1-associated disorder in a patient suspected of having or having predisposition to the disorder (i.e., a carrier), the method comprising detecting the presence of and/or evaluating the characteristics of PKD1 DNA, PKDl mRNA and.or PKDl protein in a sample taken from the patient. Such method may comprise detecting and/or evaluating whether the PKD1 DNA is deleted, missing, mutated, aberrant or not expressing normal PKDl protein. One way of carrying out such a method comprises: A. taking a biological, tissue or biopsy sample from the patient; B. detecting the presence of pand/or evaluating the characteristics of PKD1 DNA, PKD1 mRNA and/or PKD1 protein in the sample to obtain a first set of results; C.comparing the first set of results with a second set of results obtained using the same or similar methodology for an individual that is not suspected of having the disorder; and if the first and second sets of results differ in that the PKD1 DNA is deleted, missing, aberrant, mutated or not expressing PKD1 protein then that is indicative of the presence, predisposition or tendency of the patient to develop the disorder. As used herein, a "PKD1-associated disorder" refers to adult polycystic kidney disease, as described herein, and also refers to tuberous sclerosis, as well as other disorders having symptoms such as cyst formation in common with these diseases.

A specific method according to the invention comprises

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extracting from a patient a sample of PKDl DNA or DNA from the PKDl locus purporting to be PKDl DNA, cultivating the sample <u>in vitro</u> and analyzing the resulting protein, and comparing the resulting protein with normal PKDl protein according to the well-established Protein Truncation Test. Less sensitive tests include analysis of RNA using RT PCR (reverse transcriptase polymerase chain reaction), and examination of genomic DNA.

Step C of the above method may be replaced by: comparing the first set of results with a second set of results obtained using the same or similar methodology in an individual that is known to have the or at least one of the disorder(s); and if the first and second sets of results are substantially identical, this indicates that the PKD1 DNA in the patient is deleted, mutated or not expressing normal PKD1 protein.

The invention further provides a method characterizing a mutation in a subject suspected of having a mutation in the PKD1 gene, which method comprises: A amplifying each of the exons in the PKDl gene of the subject; B. denaturing the complementary strands of the amplified exons; C.diluting the denatured complementary strands to allow each single-stranded DNA molecule to assume a secondary structural confirmation; D. subjecting the DNA molecule to electrophoresis under nondenaturing conditions; E. comparing the electrophoresis pattern of the single-stranded molecule with electrophoresis pattern of a single-stranded molecule

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containing the same amplified exon from a control individual which has either a normal or PKDl heterozygous genotype; and, F. sequencing any amplification product which has an electrophoretic pattern different from the pattern obtained from the DNA of the control individual.

The invention also extends to a diagnostic kit for carrying out a method as set out above, comprising nucleic acid primers for amplifying a fragment of the DNA or RNA sequences defined above, and packaging means therefore. The kit may optionally include written instructions stating that the primers are to be used for detection of disorders associated with the PKD1 gene. The nucleic acid primers may comprise at least one of the following sets: AH3 F9: AH3 B7; 3A3 C1: 3A3 C2; and AH4 F2: JH14 B3.

Another embodiment of kit may combine one or more substances for digesting a sample to provide EcoRI fragments and a DNA probe as previously defined. A further embodiment of kit may comprise one or more substances for digesting a sample to provide BamHI fragments and a DNA probe as previously defined.

A vector (such as Bluescript (available from Stratagene)) comprising a nucleic acid sequence set out above; and a host cell (such as E. coli strain SL-1 Blue (available from Stratagene)) transfected or transformed with the vector are also provided, together with the use of such a vector or a nucleic acid sequence set out above in gene therapy and/or in the preparation of an agent for treating or preventing a PKD1-associated disorder.

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Therefore, there is further provided a method of treating or preventing a PKD1-associated disorder which method comprises administering to a patient in need thereof a functional PKD1 gene to affected cells in a manner that permits expression of PKD1 protein therein and/or a transcript produced from a mutated chromosome (such as the deleted WS-212 chromosome) which is capable of expressing functional-PKD1 protein therein.

As used herein, the term "hybridization" refers to conventional DNA/DNA or DNA/RNA hybridization conditions. For example, for a DNA or RNA probe of about 10 - 50 nucleotides, moderately stringent hybridization conditions are preferred and include 10% SSC, 5% Denhardts, 0.1% SDS, at 35 - 50 degrees for 15 hours; for a probe of about 50 -15 300 nucleotides, "stringent" hybridization conditions are preferred and refer to hybridization in 6X SSC, 5X Denhardts, 0.1% SDS at 65 degrees for 15 hours.

The present invention further provides the use of PKD1 protein or polycystin or a mutant or variant thereof having substantially the same biological activity there as in therapy. In particular, to effect cell recognition or communication for example to effect adhesion of cells to other cells or components of the extracellular matrix; effect communication and/or interaction between epithelial cells and the basal membrane (whether in kidneys or otherwise); assisting in development of connective tissue such as assembly and/or maintenance of the basal membrane; in signal transduction between cells or cells and components

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of the extracellular matrix; and/or to promote binding of cells carrying proteins such as integrins or carbohydrates to target cells.

Accordingly, where it is preferred to administer the polypeptide directly to a patient in need thereof, the invention further provides the use of a PKDl protein or polycystin in the preparation of a medicament. Therefore, there is also provided a pharmaceutical formulation comprising a PKDl protein, functional PKDl gene and/or a transcript produced from a mutated chromosome which is capable of expressing functional PKDl protein, in association with a pharmaceutically acceptable carrier therefor.

The invention also features an immunoglobin, i.e., a polyclonal or monoclonal antibody specific for an epitope of PKD1, which epitope is found in the amino acid sequence presented in Fig. 15.

The invention also features a method of assaying for the presence of PKDl in a sample of mammalian, preferably human cells, comprising the steps of: (a) providing an antibody specific for said PKDl; and (b) assaying for the presence of PKDl by admixing an aliquot from a sample of mammalian cells with antibody under conditions sufficient to allow for formation and detection of an immune complex of PKDl and the antibody. Such method is useful for detecting disorders involving aberrant expression of the PKDl gene or processing of the protein, as described herein.

Preferably, this method includes providing a monoclonal

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antibody specific for an epitope that is antigenically the same, as determined by Western blot assay, ELISA or immunocytochemical staining, and substantially corresponds in amino acid sequence to the amino acid sequence of a portion of PKD1 and having a molecular weight equal to less than that of PKD1.

The invention thus also features a kit for detecting PKD1, the kit including at least one package containing an antibody or idiotype-containing polyamide portion of an antibody raised to a synthetic polypeptide of this invention or to a conjugate of that polypeptide bound to a carrier. An indicating group or label is utilized to indicate the formation of an immune reaction between the antibody and PKD1 when the antibody is admixed with tissue or cells.

Further features will become more fully apparent in the following description of the embodiments of this invention and from the appended claims.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Before describing preferred embodiments of the invention in detail, the drawings will briefly be described.

Figure la (top): A long range map of the terminal region of the short arm of chromosome 16 showing the PKD1 candidate region defined by genetic linkage analysis. The positions of selected DNA probes and microsatellites used for haplotype, linkage or heterozygosity analyses are indicated. Markers previously described in linkage

- disequilibrium studies are shown in bold (from: Harris, et al., 1990; Harris, et al., 1991; Germino, et al., 1992; Somlo, et al., 1992; Peral, et al., 1994; Snarey, et al., 1994).
- (bottom): A detailed map of the distal part of the

  PKD1 candidate region showing: the area of 16p13.3;

  duplicated in 16p13.1 (hatched); C, Cla I restriction sites;

  the breakpoints in the somatic cell hybrids, N-OH1 and P
  MWH2A; DNA probes and the TSC2 gene. The limits of the

  position of the translocation breakpoint found in family 77
- 20 (see b), determined by evidence of heterozygosity (in 77-4)
  and PFGE (see c and text) is also indicated. The contig
  covering the 77 breakpoint region consists of the cosmids:
  1, CW9D; 2, ZDS5; 3, JH2A; 4, REP59; 5, JC10.2B; 6, CW10III;
  7, SM25A; 8, SMII; 9, NM17.
- Figure 1b: Pedigree of family 77 which segregates a 16;22 translocation; showing the chromosomal composition of each subject. Individuals 77-2 and 77-3 have the balanced products of the exchange and have PKD1; 77-4 is monosomic

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for 16p13.3-->16pter and 22q11.21-->22pter - and has TSC.

Figure 1c: PFGE of DNA from members of the 77 family: 77-1 (1); 77-2 (2); 77-3 (3); 77-4 (4); digested with Cla I and hybridised with SM6: In addition to the normal 5 fragments of 340 and partially digested fragment of 480 kb. a proximal breakpoint fragment of approximately 100 kb (arrowed) is seen in individuals, 77-2, 77-3 and 77-4; concordant with segregation of the der(16) chromosome.

Figure 2: FISH of the cosmid CW10III (cosmid 6; Figure la) to a normal male metaphase. Duplication of this locus is illustrated with two sites of hybridisation on 16p; the distal site (the PKD1 region) is arrowed. The signal from the proximal site (16pl3.1) is stronger than that from the distal, indicating that sequences homologous to CW10III are 15 reiterated in 16pl3.1. The state of the s

Figure 3a: " A detailed map of the 77, translocation region showing the precise localisation of the 77 breakpoint and the region that is duplicated in 16p13.1 (hatched). DNA probes (open boxes); the transcripts, PKD1 and TSC2 (filled 20 boxes; with direction of transcription indicated by an arrow) and cDNAs (grey boxes) are shown below the genomic map... The known genomic extent of each gene is indicated at the bottom of the diagram and the approximate genomic locations of each cDNA is indicated under the genomic map. The positions of genomic deletions found in PKDl patients, OX875 and OX114, are also indicated. Restriction sites for EcoR I (E) and incomplete maps for BamH I (B); Sac I (S) and Xba I (X) are shown: SM3 is a 2kb BamHl fragment shown at

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the 5' end of the gene.

Figure 3b: Southern blots of BamH I digested DNA from individuals: 77-1 (1); 77-2 (2); and 77-4 (4) hybridised with: left panel, 8S3 and right panel, 8S1 (see a). detects a novel fragment on the telomeric side of the breakpoint (12 kb: arrowed) associated with the der(22) chromosome in 77-2, but not 77-4; 8S1 identifies a novel fragment on the centromeric side of the breakpoint (9 kb: arrowed) - associated with the der(16) chromosome - in 77-2 and 77-4. The telomeric breakpoint fragment is also seen 10 weakly with 8S1 (arrowed) indicating that the breakpoint lies in the distal part of 8S1. The 8S3 and 8S1 loci are both duplicated: the normal BamH I fragment detected at the 16pl3.3 site by these probes is 11 kb (see a), but a similar sized fragment is also detected at the 16pl3.1 site. Consequently, the breakpoint fragments are much fainter than the normal (16p13.1 plus 16p13.3) band.

Figure 4a: PBP cDNA, 3A3, hybridised to a Northern blot containing about 1 µg polyA selected mRNA per lane of the tissue specific cell lines: lane 1, MJ, EBV-transformed lymphocytes; lane 2, K562, erythroleukemia; lane 3, FS1, normal fibroblasts; lane 4, HeLa, cervical carcinoma; lane 5, G401, renal Wilm's tumour; lane 6, Hep3B, hepatoma; lane 7, HT29, colonic adenocarcinoma; lane 8, SW13, adrenal carcinoma; lane 9, G-CCM, astrocytoma. A single transcript of approximately 14 kb is seen; the highest level of expression is in fibroblasts and in the astrocytoma cell line; G-CCM. Although in this comparative experiment little

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expression is seen in lanes 1, 4 and 7, we have demonstrated at least a low level of expression in these cell lines on other Northern blots and by RT-PCR (see later).

Figure 4b: A Northern blot containing about 20 µg of
testal RNA from the cell line G-CCM hybridised with cDNAs or
a genomic probe which identify various parts of the PBP
gene. Left panel, a single about 14 kb transcript is seen
with a cDNA from the single copy area, 3A3. Right panel, a
cDNA, 21P.9, that is homologous to parts of the region that
is duplicated (JH12, JH8 and JH10; see Figure 3a) hybridises
to the PBP transcript and three novel transcripts; HG-A
(about 21 kb), HG-B (about 17 kb) and HG-C (8.5 kb). A
similar pattern of transcripts is seen with cDNAs and
genomic fragments that hybridise to the area between JH5 and
JH13, with the exception of the JH8 area. Middle panel, JH8
hybridises to the transcripts PBP, HG-A and HG-B but not to
HG-C.

Figure 4c: A Northern blot of 20 µg total fibroblast RNA from: normal control (N); 77-2 (2); 77-4 (4) hybridised with 8S1, which contains the 16;22 translocation breakpoint (see Figure 3). A transcript of about 9 kb (PBP-77) is identified in the two patients with this translocation but not in the normal control. PBP-77 is a chimeric PBP transcript formed due to the translocation and is not seen in 77-2 or 77-4 RNA with probes which map distal to the breakpoint.

Figure 5a: FIGE of DNA from: normal (N) and ADPKD patient OX875 (875), digested with EcoR I and hybridised

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with, left panel, CW10; middle panel, JH1. Normal fragments of 41 kb (plus a 31 kb fragment from the 16p13.1 site), CW10, and 18 kb, JHI, are identified with these probes; OX875 has an additional 53 kb band (arrowed). The ECOR I site separating these two fragments is removed by the deletion (see Figure 3a). The right panel shows a Southern blot of BamH I digested DNA (as above) hybridised with 1A1H.6. A novel fragment of 9.5 kb is seen in OX875 DNA, as well as the normal 15 kb fragment. These results indicate that OX875 has a 5.5 kb deletion; its position was determined more precisely by mapping relative to two Xba I sites which flank the deletion (see figure 3a).

Figure 5b: Northern blot of total fibroblast RNA, as (a), hybridised with the cDNAs, AH4, 3A3 and AH3. A novel transcript (PBP-875) of about 11 kb is seen with AH4 (the band is reduced in intensity because the probe is partly deleted) and AH3 (arrowed), which flank the deletion, but not 3A3 which is entirely deleted (see figure 3a). The transcripts HG-A, HG-B and HG-C, from the duplicated area, are seen with AH3 (see figure 4b).

Figure 5c: Left panel; FIGE of DNA from: normal (N) and ADPKD patient OX114 (114), digested with EcoR I and hybridised with CW10; a novel fragment of 39 kb (arrowed) is seen in OX114. Middle panel; DNA, as above, plus the normal mother (M) and brother (B) of OX114 digested with BamH I and hybridised with CW21. A larger than normal fragment of 19 kb (arrowed) was detected in OX114 but not other family members due to deletion of a BamH I site; together these

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results are consistent with a 2 kb deletion (see Figure 3a).

Right panel; RT-PCR of RNA, as above, with primers flanking the OX114 deletion (see Experimental Procedures). A novel fragment of 810 bp (arrowed) is seen in OX114, indicating a deletion of 446 bp in the PBP transcript.

Figure 5d: RT-PCR of RNA from: ADPKD patient 0X32 (32) plus the probands, normal mother (M) and affected father (F) and sibs (1) and (2) using the C primer pair from 3A3 (see Experimental Procedures). A novel fragment of 125 bp is detected in each of the affected individuals.

Figure 6: Map of the region containing the TSC2 and PBP genes showing the area deleted in patient WS-53 and the position of the 77 translocation breakpoint. Localisation of the distal end of the WS-53 deletion was described (European Chromosome 16 Tuberous Sclerosis Consortium, 1993) and we have now localised the proximal end between SM6 and JH17. . The size of the aberrant Mlu A fragment in WS-53, detected by JH1 and JH17, is 90kb and these probes lie on adjacent Mlu I fragments of 120kb and 70kb, respectively. Therefore the WS-53 deletion is about 100kb. Restriction sites for: Mlu I (M); Nru I (R); Not I (N); and partial maps for Sac II (S) and BssH II (H) are shown. DNA probes (open boxes) and the TSC2 and PBP transcripts (filled boxes) are indicated below the line with their known genomic extents (brackets). The locations of the microsatellites KG8 and SM6 are also indicated.

Figure 7: The partial nucleotide sequence (cDNA) of the PKD1 transcript extending 5631bp to the 3' end of the

gene. The corresponding predicted protein (also shown in SEQ ID NO: 4:) is shown below the sequence and extends from the start of the nucleotide sequence. The GT-repeat, KG8, is in the 3' untranslated region between 5430-5448 bp.

This sequence corresponds to GenBank Accession No. L33243 and is shown in SEQ ID NO: 3:

Figure 8: The sequence of the probe 1A1H0.6 (also shown in SEQ ID NO: 5:).

Figure 9: The sequence (SEQ ID NO: 6:) of the probe CW10 which is about 0.5kb.

Figure 10: The larger partial nucleotide sequence (SEQ ID NO: 1:) of the PKD1 transcript (cDNA) extending from bp 2 to 13807bp to the 3' end of the gene together with the corresponding predicted protein (also shown in SEQ ID NO: 2:). This larger partial sequence encompasses the (smaller) partial sequence of Figure 7 from amino acid no. 2726 in SEQ ID NO: 3: and relates to the entire PKD1 gene sequence apart from its extreme 5' end.

Figure 11: A map of the 75bp intron amplified by the primer set 3A3C insert at position 3696 of the 3' sequence showing the positions of genomic deletions found in PKD1 patients 461 and OX1054.

Figure 12: A map of the region of chromosome 16 containing the TSC2 and PKD1 genes showing the areas affected in patients WS-215, WS-250, WS-212, WS-194, WS-227 and WS-219; also WS-53 (but cf. Figure 6). Genomic sites for the enzymes Mlul (M), Clal (C), Pvul (P) and Nrul (R) are shown. Positions of single copy probes and cosmids used

to screen for deletions are shown below the line which represents about 400kb of genomic DNA. The genomic distribution of the approximately 45kb TSC2 gene and known extent of the PKD1 gene are indicated above. The hatched area represents an about 50kb region which is duplicated more proximally on chromosome 16p.

Figure 13 is a genomic map of the PKDl gene. (Top) A restriction map of the genomic area containing the PKD1 gene showing sites for Bam H1(B), EcoRI(E) and partial maps for Xbal (X) and Hind III(H), and the duplicated area (hatched). 10 The position of genomic clones and the cosmid JH2A are shown 🗈 above the map (open boxes). The positions of the 46 exons of the PKD1 gene are shown below the map (solid boxes, 45 translated areas; open boxes, untranslated regions; UTRs). 15 Each 5th exon is numbered and the direction of transcription 🐲 arrowed. The area sequenced in Figs. 7 and 10 is bracketed and the approximate location of the 3' end of the TSC2 gene is shown on the left (dashed line and hatched box). (Bottom) The cDNA contig covering the PKD1 transcript. 20 cDNAs are: 1, rev1; 2, S13;3, S3/4; 4, S1/3;5, GAP e; 6, GAP d; 7, GAP g; 8, GAP a (see table 2 for details); 9, A1C; 10, AH3; 11, 3A3; 12, AH4.

Figure 14 (a) (Top): Map of the genomic BamH I fragment, SM3 which contains the CpG island at the 5' end of the PKD1 gene, showing the probe CW45 (open box). Genomic restriction sites for the methylation sensitive enzymes: SacII (S), Notl (N), Mlul (M) and BssHII (H) are illustrated. The approximate position of the DNasel

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hypersensitive site is also shown (large arrow), plus the location of the first exon including the proposed transcription start site (small arrow), the 5'UTR (open box) and the translated region (solid bar). (Bottom) The GC content across the area is plotted with a window size of 50 nt. A peak of GC content of over 80% is seen in the area of the transcriptional start site and the first exon. A corresponding lack of CpG suppression was also found with an average CpG/GC ratio of 0.84 between 800-1,800 bp.

Figure 14(b). Analysis of DNase I hypersensitivity at the PKD1 CpG island. DNA isolated from HeLa cells treated with an increasing amount of DNase I (left to right; first lane contains no DNase 1), digested with BamH I and hybridised with CW45. A fragment about 400 bp smaller than the restriction fragment is seen with increasing DNase 1, indicating a hypersensitive site as shown in (a). SM3 is within the duplicated area and so both the PKD1 and HG loci are assayed together. The degree of DNasel digestion seen at the end of the assay indicates that cleavage occurs at the PKD1 and HG loci.

Figure 15 provides the sequence of the PKD1 transcript and predicted protein. The full sequence of 14,148 bp from the transcription start site to the poly A tail is shown. The probable signal sequence of 23 amino acids is shown after the first methionine (underlined) plus the cleavage site (arrow). The predicted transmembrane (TM) domains (double underlined and numbered) and N-linked glycosylation sites (asterisk) are indicated. The position of a possible

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hinge sequence is underlined and tyrosine kinase and protein kinase C phosphorylation sites marked with a box and circle, respectively.

Figure 16(a). The leucine rich repeats (LRRs) found in the PKD1 protein (72-125aa) are compared with each other and to the LRR consensus (Rothberg, 1990; Kobe, 1994); a, aliphatic. A total of just over 2 full repeats are present in PKD1 but they have been arranged into 3 incomplete repeats to show their similarity to those found in slit (Rothberg, 1990). The black boxes show identity to the LRR consensus and shaded boxes other regions of similarity between the repeats which have also been noted in other LRRs (Kobe, 1994).

Figure 16(b). The amino flanking region to the LRR in the PKD1 protein (33-71aa) is compared similar regions from & a variety of other proteins. Black boxes shown identity with the consensus (adapted from [Rothberg, 1990 #1126]) and shaded boxes conserved amino acids. The different types of residue indicated in the consensus are: a, as above; p, polar or turn-like; h, hydrophobic. The listed proteins, with the species and Protein Identification Resource no. (PIR) shown in brackets, are: OMgp, oligodendrocyte myelin glycoprotein (Human, A34210); Slit (Drosophila; A36665); Chaoptin (Drosophila; A29943); GP-IB Beta, glycoprotein 1bB chain (Human; A31929); Pgl; proteoglycan-1 (mouse; 520811); Biglycan (Human; A40757); Trk (Human; A25184) and LH-CF, lutropinchoriogonadotrophin receptor (Rat: A41343).

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Figure 16(c). The carboxy flanking region of the LRR repeat from the PKD1 protein (126-180 aa) compared to similar regions in other proteins and a consensus accepted from [Rothberg, 1990 #1126]. The shading and amino acid types are as above. The proteins not described above are: Toll (Drosophila; A29943) and GP IX, platelet glycoprotein IX (Human; A46606).

Figure 17 is a sequence comparison of the C-type lectin domain. The PKD1 lectin domain (403-532aa) is compared to those of: BRA3, acorn barnacle lectin (JC1503); Kupffer cell carbohydrate-binding receptor (Rat; A28166), CSP, cartilage specific protoglycan (Bovine; A27752); Agp; asialoglycoprotein receptor (Human; 55283), E-Selectin (Mouse; B42755) and glycoprotein gp120 (Human; A46274). Black squares show identify with the consensus and shaded boxes conserved residues. Amino acid types are: Very highly conserved residues are shown in bold in the consensus which is adapted from Drickamer 1987, Drickamer 1988.

The 16 copies of the PKD1 Ig-like repeat (PKDI 273-356 aa;
PKDII-XVI, 851-2145aa) are compared to each other and to:
V.a. colAi, and C.p. colA collagenases of Vibrio alginolyticus (S19658) and Clostridium perfringens (D13791),
respectively; Pmel17, melanocyte specific glycoprotein
(Human; A41234), FLT4; Ig repeat IV of fms-like tyrosine kinase 4 (Human; X68203), CaVPT, Ig repeat I of target protein of the calcium vector protein (CAVP) (amphioxius; P05548). black boxes shown amino acids identical in more

than 5 repeats and shaded boxes related residues. An Ig consensus determined from Harpaz et al. 1994 and Takagi et al. 1990 is shown in the symbols: a, aliphatic; h, hydrophobic; s, small and b, base with the predicted positions of the B-strands indicated below. The PKD repeat IV has an extra repetition of 20 aa in the centre of the repeat while all of the others are between 84-87 aa.

Figure 19 reveals type III-related fibronectin domains. The four fibronectin-related domains from the PKD1 protein (2169-2573aa) are compared to similar domains in: Neuroglian 10 (Drosophila; A32579); L1, neural recognition molecule L1 (X59847); F11, neural cell recognition molecule F11 (X14877); TAG 1, transiently expressed axonal surface glycoprotein-1 (Human; S28830); F3, Neuro-1 antigen (mouse; 15 SO5944); NCAM, neural cell adhesion molecule (Rat; X06564); -DCC, deleted in colorectal cancer (Human; X76132); LAR, Leukocyte-common antigen related molecule (Human; Y00815); HPTP, B protein tyrosine phosphate beta (Human; X54131) and FN, fibronectin (Human; X02761). The consensus sequence is compiled from Borh and Doolittle (1993), Kuma et al. (1993), 20 Baron et al. (1992) and Borh and Doolittle (1992). boxes show identity to highly conserved residues and shaded boxes conserved changes or similarity in less highly The approximate positions of the  $\boldsymbol{\beta}$ conserved positions. 25 strands are illustrated. The fibronectin repeats in the PKD1 protein are linked by sequences of 27aa (A-B), 22aa (B-C) and 7aa (C-D) which are not shown.

Figure 20 presents a proposed model of the PKD1

protein, polycystin. The predicted structure of the PKD1 protein is shown.

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#### DETAILED DESCRIPTION

All references mentioned herein are listed in full at the end of the description which are herein incorporated by reference in their entirety. Except where the context clearly indicates otherwise, references to the PBP gene, transcript, sequence, protein or the like can be read as referring to the PKDl gene, transcript, sequence, protein or the like, respectively.

#### A translocation associated with ADPKD

provided by a Portuguese pedigree (family 77) with both ADPKD and TSC (Figure 1b). Cytogenetic analysis showed that the mother, 77-2, has a balanced translocation, 46XX t(16;22) (p13.3;q11.21) which was inherited by her daughter, 77-3. The son, 77-4, has the unbalanced karyotype, 45XY-16-22+der(16) (16qter-->16p13.3: :22q11.21-->2qter) and consequently is monosomic for 16p13.3-->16pter as well as for 22q11.21-->22pter. This individual has the clinical phenotype of TSC (see Experimental Procedures); the most likely explanation is that the TSC2 locus located within 16p13.3 is deleted in the unbalanced karyotype.

Further analysis revealed that the mother (77-2), and the daughter (77-3) with the balanced translocation, have the clinical features of ADPKD (see Experimental Procedures), while the parents of 77-2 were cytogenetically normal, with no clinical features of TSC and no renal cysts on ultrasound examination (aged 67 and 82 years). Although kidney cysts can be a feature of TSC, no other clinical

signs of TSC were identified in 77-2 or 77-3, making it unlikely that the polycystic kidneys were due to TSC. We therefore investigated the possibility that the translocation disrupted the PKDl locus in 16pl3.3 and proceeded to identify and clone the region containing the breakpoint.

The 77 family was analyzed with polymorphic markers from 16p13.3. Individual 77-4 was hemizygous for MS205.2 and GGG1, but heterozygous for SM6 and more proximal markers, locating the translocation breakpoint between GGG1 and SM6 (see Figure 1a). Fluorescence in situ hybridization (FISH) of a cosmid from the TSC2 region, CW9D (cosmid 1 in Figure 1a), to metaphase spreads showed that it hybridized to the der(22) chromosome of 77-2; placing the breakpoint proximal to CW9D and indicating that 77-4 was hemizygous for this region consistent with his TSC phenotype. DNA..from members of the 77 family was digested with Cla I, separated by PFGE and hybridized with SM6; revealing a breakpoint fragment of about 100 kb in individuals with the der(16) chromosome (Figure 1c). The small size of this novel fragment enabled the breakpoint to be localized distal to SM6 in a region of just 60 kb (Figure la). A cosmid contig covering this region was therefore constructed Experimental Procedures for details).

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The translocation breakpoint lies within a region duplicated elsewhere on chromosome 16p (16p13.1)

It is noted hereabove that the region between CW21 and .N54 (Figure 1a) was duplicated at a more proximal site on

the short arm of chromosome 16 (Germino, et al., 1992; European Chromosome 16 Tuberous Sclerosis Consortium, 1993). Figure 2 shows that a cosmid, CW10III, from the duplicated region hybridized to two points on 16p; the distal, PKD1 region and a proximal site positioned in 16p13.1. The structure of the duplicated area is complex with each fragment present once in 16p13.3 re-iterated two-four times in 16p13.1 (see Figure 2). Cosmids spanning the duplicated area in 16p13.3 were subcloned (see Figure 3a and Experimental Procedures for details) and a restriction map was generated. A genomic map of the PKD1 region was constructed using a radiation hybrid, Hy145.19 which contains the distal portion of 16p but not the duplicate site in 16p13.1.

from the target region were hybridized to 77-2 DNA; digested with Cla I and separated by PFGE. Once probes mapping across the breakpoint were identified they were hybridized to conventional Southern blots of 77 family DNA. Figure 3b shows that novel BamH I fragments were detected from the centromeric and telomeric side of the breakpoint, which was localized to the distal part of the probe 8S1 (Figure 3a). Hence, the balanced translocation was not associated with a substantial deletion, and the breakpoint was located more than 20 kb proximal to the TSC2 locus (Figure 3a). These results supported the hypothesis that polycystic kidney disease in individuals with the balanced translocation (77-2 and 77-3) was not due to disruption of the TSC2 gene, but

indicated that a separate gene mapping just proximal to TSC2, was likely to be the PKD1 gene.

The polycystic breakpoint (PBP) gene is disrupted by the translocation

Localization of the 77 breakpoint identified a precise region in which to look for a candidate or the PKD1 gene. During the search for the TSC2 gene we identified other transcripts not associated with TSC including a large transcript (about 14 kb) partially represented in the cDNAs 10 3A3 and AH4 which mapped to the genomic fragments CW23 and CW21 (Figure 3a). The orientation of the gene encoding this transcript had been determined by the identification of a polyA tract in the cDNA, AH4: the 3' end of this gene lies very close to the TSC gene, in a tail to tail orientation (European Chromosome 16 Tuberous Sclerosis Consortium, To determine whether this gene crossed the translocation breakpoint genomic probes from within the duplicated area and flanking the breakpoint were hybridized to Northern blots. Probes from both sides of the breakpoint, between JH5 and JH13 identified the 14 kb 3a and see below for details). (Figure Therefore, this gene, called 3A3, but not designated the PBP gene extended over the 77 breakpoint and consequently was a candidate for the PKD1 gene. A walk was initiated to 25 increase the extent of the PBP cDNA contig and several new cDNAs were identified using probes from the single copy (non-duplicated) region (see Experimental Procedures for details). A cDNA contig was constructed which extended

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about 5.7 kb, including about 2 kb into the area that is duplicated (Figure 3a).

### Expression of the PBP gene

Initial studies of the expression pattern of the PBP gene were undertaken with cDNAs that map entirely within the single copy region (e.g. AH4 and 3A3). Figure 4a shows that the about 14 kb transcript was identified by 3A3 in various tissue-specific cell lines. From this and other Northern blots we concluded that the PBP gene was expressed in all of the cell lines tested, although often at a low level. The two cell lines which showed the highest level of expression were fibroblasts and a cell line derived from an astrocytoma, G-CCM. Significant levels of expression were also obtained in cell lines derived from kidney (G401) and liver (Hep3B). Measuring the expression of the PBP gene in tissue samples by Northern blotting proved difficult because such a large transcript is susceptible to minor RNA degradation. However, initial results with an RNAse protection assay, using a region of the gene located in the single copy area (see Experimental Procedures), showed a moderate level of expression of the PBP gene in tissue obtained from normal and polycystic kidney (data not shown). The widespread expression of the PBP is consistent with the systemic nature of ADPKD.

Identification of transcripts that are partially homologous to the PBP transcript

New cDNAs were identified with the genomic fragments,

JH4 and JH8, that map to the duplicated region (Figure 3a)

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and see Experimental Procedures). However, when these cDNAs were hybridized to Northern blots a more complex pattern than that seen with 3A3 was observed. As well as the  $^{-}14~\mathrm{kb}$ transcript, three other, partially homologous PBP 5 transcripts were identified designated homologous gene-A (HG-A; ~21 kb), HG-B (~17 kb) and HG-C (8.5 kb) Figure 4b). There were two possible explanations for these results, either the HG transcripts were alternatively spliced forms of the PBP gene, or the HG transcripts were encoded by gene located in 16pl3.1. To determine the genomic location of "the HG loci a fragment from the 3' end of one HG cDNA (HG-4/1.1) was isolated. HG-4/1.1 hybridized to all three HG transcripts, but not to the PBP transcript and on a hybrid panel it mapped to 16p13.1 (not the PKD1 area). These 15 results show that all the HG transcripts are related to each other outside the region of homology with the PBP transcript and that the HG loci map to the proximal site (16p13.1). An abnormal transcript associated with the 77 translocation

As the PBP gene was transcribed across the region disrupted by the 77 translocation breakpoint, in a proximal to distal direction on the chromosome (see Figure 3a) it was possible that a novel transcript originating from the PBP promotor would be found in this family. Figure 4c shows that using a probe to the PBP transcript that mapped mainly proximal to the breakpoint, a novel transcript of approximately 9 kb (PBP-77) derived from the der(16) product of the translocation was detected. Interestingly, the PBP-77 transcript appears to be expressed at a higher level than

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the normal PBP product. These results confirmed that the 77 translocation disrupts the PBP gene and supports the hypothesis that this is the PKD1 gene:

Mutations of the PBP gene in other ADPKD patients

To prove that the PBP gene is the defective gene at the PKD1 locus, we analyzed this region for mutations in patients with typical ADPKD. The 3' end of the PBP gene was most accessible to study as it maps outside the duplicated area. To screen this region BamH'I digests of DNA from 282 apparently unrelated ADPKD patients were hybridized with the probe 1AlH.6, (see Figure 3a). In addition, a large EcoR I fragment (41 kb) which contains a significant proportion of the PBP gene was assayed by field inversion gel electrophoresis (FIGE) in 167 ADPKD patients, using the 15 probe CW10. Two genomic rearrangements were identified in ADPKD patients by these procedures; each identified by both And a control of the first of the first methods.

The first rearrangement was identified in patient 0X875 (see Experimental Procedures for clinical details) who was 20 shown to have a 5.5 kb genomic deletion without the 3' end of the PBP gene, producing a smaller transcript (PBP-875) (see Figures 5a, b and 3a for details). This genomic deletion results in a ~3 kb internal deletion of the transcript with the 500 bp adjacent to the polyA tail In this family linkage of ADPKD to chromosome 16 could not be proven because although OX875 has a positive family history of ADPKD there were no living, affected relatives. However, paraffin-embedded tissue from her

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affected father (now deceased) was available. We demonstrated that this individual has the same rearrangement as OX875 by PCR amplification of a 220bp fragment spanning the deletion (data not shown). This result and analysis of two unaffected sibs of OX875, that did not have the deletion, showed that this mutation was transmitted with ADPKD.

The second rearrangement detected by hybridization was a 2 kb genomic deletion within the PBP gene, in ADPKD patient OX114 (see Experimental Procedures for clinical details and Figures 5c and 3a). No abnormal PBP transcript was identified by Northern blot analysis, but using primers flanking the deletion (see Experimental Procedures), a shortened product was detected by RT-PCR (Figure 5c). This was cloned and sequenced and shown to have a frame-shift deletion of 446 bp (between base pair 1746 and 2192 of the sequence shown in Figure 7). OX114 is the only member of the family with ADPKD (she has no children) and ultrasound analysis of her parents at age 78 (father) and 73 years old (mother) showed no evidence of renal cysts. Somatic cell hybrids were produced from OX114 and the deleted chromosome was found to be of paternal origin by haplotype analysis. The father of OX114 is now deceased but analysis of DNA from the brother of OX114 (OX984) with seven microsatellite markers from the PKD1 region (see Experimental Procedures) showed that he shares the same paternal chromosome, in the PKD1 region, as OX114. Renal ultrasound revealed no cysts in 0X984 at age 53 and no deletion was detected by DNA

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analysis (Figure 5c). Hence, the deletion in OX114 is a de novo event associated with the development of ADPKD. Although it is not possible to show that the ADPKD is chromosome 16-linked, the location of the PBP gene indicated that this is a de novo PKDI mutation.

regions of the PBP gene were analyzed by RT-PCR using RNA isolated from lymphoblastoid cell lines established from ADPKD patients. cDNA from 48 unrelated patients was amplified with the primer pair 3A3 C (see Experimental Procedures) and the product of 260 bp was analyzed on an agarose gel. In one patient, OX32, an additional smaller product (125bp) was identified, consistent with a deletion or splicing mutation. OX32 comes from a large family in which the disease can be traced through three generations. Analysis of RNA from two affected sibs of OX32 and his parents showed that the abnormal transcript segregates with PKD1 (Figure 5d).

Amplification of normal genomic DNA with the 3A3 C primers generates a product of 418 bp; sequencing showed that this region contains two small introns (5', 75 bp and 3', 83 bp) flanking a 135 bp exon. The product amplified from OX32 genomic DNA was normal in size, excluding a genomic deletion. However, heteroduplex analysis of that DNA revealed larger heteroduplex bands, consistent with a mutation within that genomic interval. The abnormal OX32, RT-PCR product was cloned and sequenced: this demonstrated that, although present in genomic DNA, the 135 bp exon was

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genomic DNA demonstrated a G-->C transition at +1 of the splice donor site following the 135 bp exon. This mutation was confirmed in all available affected family members by digesting amplified genomic DNA with the enzyme Bst NI: a site is destroyed by the base substitution. The splicing defect results in an in-frame deletion of 135 bp from the PBP transcript (3696 bp to 3831 bp of the sequence shown in Figure 7). Together, the three intragenic mutations confirm that the PBP gene is the defective gene at the PKD1 locus. Deletions that disrupt the TSC2 and the PKD1 gene

The deletion called WS-53 disrupts both the TSC2 gene and the PKD1 gene (European Chromosome 16 Tuberous Sclerosis Consortium, 1993), although the full proximal extent of the deletion was not determined. Further study has shown that the deletion extends ~100 kb (see Figure 6 for details) and deletes most if not all of the PKD1 gene. This patient has TSC but also has unusually severe polycystic disease of the kidneys. Other patients with a similar phenotype have also been under investigation. Deletions involving both TSC2 and PKD1 were identified and characterized in six patients in whom TSC was associated with infantile polycystic kidney disease. As well as the deletion in WS-53, those in WS-215 and "S-250 also extended proximally well beyond the known distribution of PKD1 and probably delete the entire gene. The deletion in WS-194 extended over the known extent of PKD1, but not much further proximally, while the proximal breakpoints in WS-219 and WS-227 lay within PKD1 itself.

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Northern analysis of case WS-219 with probe JH8, which lies outside the deletion, showed a reduced level of the PKD1 transcript but no evidence of an abnormally sized transcript (data not shown). Analysis of samples from the clinically unaffected parents of patients WS-53, WS-215, WS-219, WS-227 and WS-250 showed the deletions in these patients to be de novo. The father of WS-194 was unavailable for study.

In a further case (WS-212), renal ultrasound showed no cysts at four years of age but a deletion was identified which removed the entire TSC2 gene and deleted an XbaI site which is located 42 bp 5' to the polyadenylation signal of To determine the precise position of the proximal breakpoint in PKD1; a 587bp probe from the 3' untranslated region (3 UTR) was hybridized to XbaI digested DNA. A 15kb 15 XbaL 1 breakpoint fragment was detected with an approximately equal intensity to the normal fragment of 6kb, indicating that most of the PKD1 3'UTR was preserved on the mutant chromosome. Evidence that a PKD1 transcript is produced from the deleted chromosome in WS-212 was obtained by 3' rapid identification of cDNA ends (RACE) with a novel, product generated from WS-212 CDNA. Characterization of this product showed that polyadenylation occurs 546bp 5' to the normal position, within the 3'UTR of PKD1 (231bp 3' to the stop codon at 5073bp of the described PKDl sequence14). A transcript with an intact open reading frame is thus produced from the deleted WS-212 chromosome. It is likely that a functional PKD1 protein in produced from this transcript, explaining the lack of cystic disease in

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this patient. The sequence preceding the novel site of polyA addition is:

AGTCAGTAATTTATATGGTGTTAAAATGTG(A)n.

Although not conforming precisely to the consensus of AATAAA, it is likely that part of this AT rich region acts as an alternative polyadenylation signal if, as in this case, the normal signal is deleted (a possible sequence is underlined).

The WS-212 deletion is 75kb between SM9-CW9 distally 10 and the PKD1 3'UTR proximally. The WS-215 deletion is 160kb between CW15 and SM6-JH17. WS-194 has 65kb deleted between CW20 and CW10-CW36. WS-227 has a 50kb deletion between CW20 and JH11 and WS-219 has a 27kb deletion between JH1 and JH6. The distal end of the WS-250 deletion is in CW20 but the 15 precise location of the proximal end is not known. However, the same breakpoint fragment of 320kb is seen with Pvuldigested DNA using probes on adjacent Pvul fragments, CE18 (which normally detects a 245kb fragment) and Blu24 (235kb). Hence this deletion can be estimated 160kb. b. PFGE analysis of the deletion in WS-219. Mlul digested DNA from a normal control (N) and WS-219 probed with the clones H2, JH1, CW21 and CW10 which detect an ~130kb fragment in normal individuals. CW10 also detects a much smaller fragment from the duplicated region situated more proximally on 16p. A novel fragment of 100kb is seen in WS-219 with probes H2 and CW10 which flank the deletion in this patient. JH1 is partially deleted but detects the novel band weakly. aberrant fragment is not detected by CW-21, which is deleted

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BamHl digested DNA of normal on the mutant chromosome. control (N) and WS-219 separated by conventional gel electrophoresis and hybridized to probes JHl and JH6 which flank the deletion. The same breakpoint fragment of 3kb is seen with both probes, consistent with a deletion of ~27kb ending within the BamHl fragments seen by these probes. Two further deletions

In addition we have characterized two further mutations of this gene which were identified in typical PKD1 families. 10 In both cases the mutation is a deletion in the 75bp intron amplified by the primer pair 3A3C (European Polycystic Kidney Disease Consortium, 1994). The deletions are of 18bp and 20bp, respectively, in the patients 461 and 0X1054. Although these deletions do not disrupt the highly conserved 15 sequences flanking the exon/intron boundaries, they do result in aberrant splicing of the transcript. On both cases, two abnormal mRNAs are produced, one larger and one smaller than normal. Sequencing of these cDNAs showed that the larger transcript includes the deleted intron, and so 20 has an in-frame insertion of 57bp in 461, while OX1054 has a frameshift insertion of 55bp. The smaller transcript is due to activation of a cryptic splice site in the exon preceding the deleted intron and results in an in-frame deletion of 66bp in both patients. The demonstration of two additional mutations of this gene in PKDl patients further confirms that this is the PKD1 gene.

Partial Characterization of the PKD1 gene

To characterize the PKD1 gene further, evolutionary

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conservation was analyzed by 'zoo blotting'. Using probes from the single copy, 3' region (3A3) and from the duplicated area (JH4, JH8) the PKD1 gene was conserved in other mammalian species, including horse, dog, pig and rodents (data not shown). No evidence of related sequences were seen in chicken, frog or drosophila by hybridization at normal stringency. The degree of conservation was similar when probes from the single copy of the duplicated region were employed.

Although the full genomic extent of the PKD1 gene was not yet known, results obtained by hybridization to Northern blots showed that it extended from at least as far as JH13. Several CpG islands were localized 5' of the known extent of the PKD1 gene (Figure 6), although there was no direct evidence that any of these are associated with this gene.

The cDNA contig extending 5631 bp to the 3' end of the PKD1 transcript was sequenced; where possible more than one cDNA was analyzed and in all regions both strands were sequenced (Figure 7). We estimated that this accounts for 40% of the PKD1 transcript. An open reading frame was detected which runs from the 5' end of the region sequenced and spans 4842 bp, leaving a 3' untranslated region of 789 bp which contains the previously described microsatellite, KG8 (Peral, et al., 1994; Snarey, et al., 1994). A polyadenylation signal is present at nucleotides 5598-5603 and a polyA tail was detected in two independent cDNAs (AH4 and AH6) at position, 5620. Comparison with the cDNAs HG-4 and 11BHS21, which are encoded by genes in the duplicate,

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16pl3.1 region, show that 1866 bp at the 5' end of the partial PKD1 sequence shown in Figure 7 lies within the duplicated area. The predicted amino acid sequence from the available open reading frame extends 1614 residues, and is shown in Figure 7. A search of the swissprot and NBRF data bases with the available protein sequence, using the Blast program (Altschul, et al., 1990) identified only short regions of similarity (notably, between amino-acids 690-770 and 1390-1530) to a diverse group of proteins; no highly significant areas of homology were recognized. importance of the short regions of similarity is unclear as the search for protein motifs with the ProSite Program did not identify any recognized functional protein domains within the PKDI gene.

The test of identifying and characterizing the PKD1 gene has been more difficult than for other disorders because more than three quarters of the gene is embedded in a region of DNA that is duplicated elsewhere on chromosome 16. This segment of 40-50 kb of DNA, present as a single 20 copy in the PKD1 area (16pl3.3), is re-iterated as several divergent copies in the more proximal region, 16p13.1. This proximal site contains three gene loci (HG-A, -B and -C) that each produce polyadenylated mRNAs and share substantial homology to the PKD1 gene; it is not known whether these partially homologous transcripts are translated functional proteins.

Although gene amplification is known as a major mechanism for creating protein diversity during evolution,

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the discovery of a human disease locus embedded within an area duplicated relatively recently is a new observation. In this case because of the recent nature of the reiteration the whole duplicated genomic region retains a high level of homology, not just the exons. The sequence of events leading to the duplication and which sequence represents the original gene locus are not yet clear. However, early evidence of homology of the 3' ends of the three HG transcripts which are different from the 3' end of the PKD1 gene indicated that the loci in 16p13.1 have probably arisen by further reiteration of sequences at this site, after it separated from the distal locus.

To try to overcome the duplication problem we employed an exon linking approach using RNA isolated from a radiation hybrid, HY145.19, that contains just the PKD1 part of chromosome 16, and not the duplicate site in 16p13.1. Hence, this hybrid produces transcripts from the PKD1 gene but not from the homologous genes (HG-A, HG-B and HG-C). We have also sequenced much of the genomic region containing the PKD1 gene, from the cosmid JH2A, and have sequenced a number of cDNAs from the HG locus. To determine the likely position of PKD1 exons in the genomic DNA we compared HG CDNAs, (HG-4 and HG-7) to the genomic sequence. We then designed primers with sequences corresponding to the genomic DNA, to regions identified by the HG exons and employing DNA generated from the hybrid HY145.19, we amplified sections of The polymerase Pfu was used to the PKD1 transcript. minimise incorporation errors. These amplified fragments

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were then cloned and sequenced. The PKD1 cDNA contig whose sequence is shown in Figure 10 is made up of (3'-5') the original 5.7 kb of sequence shown in Figure 7, and the cDNAs: gap a 22 (890 bp), gap gamma (872 bp), a section of genomic DNA from the clone JH8 (2,724 bp) which corresponds to a large exon, S1-S3 (733 bp), S3-S4 (1,589 bp) and S4-S13 (1,372 bp). Together these make a cDNA of 13,807nt. When these cDNAs from the PKD1 contig were sequenced an open reading frame was found to run from the start of the contig to the stop codon, a region of 13,018 bp. The predicted protein encoded by the PKD1 transcript is also shown in Figure 10 and has 4,339 amino acid residues.

#### Cloning a full length PKD1 cDNA

cDNAs known to originate from the PKD1 or HG
transcripts show on average a sequence divergence of less
than 3%. Consequently, although many cDNAs were identified
by hybridisation of various PKD1 genomic probes to cDNA
libraries, it proved difficult to differentiate genuine PKD1
clones from those of the HG transcripts. For this reason a
novel strategy was employed to clone the PKD1 transcript.

To obtain a template of genomic sequence of the PKD1 gene, clones which contain the transcribed region, JH6 and JH8-JH13, were sequentially truncated and sequenced. These clones were isolated from the cosmid JH2A, which extends into the single copy area containing the 3' portion of the PKD1 gene (figure 13) and hence represents the PKD1 and not the HG loci. As a result of this analysis a contig of about 18 kb of genomic sequence was generated, which was

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ultimately found to encode >95% of the unsequenced portion of the PKD1 transcript.

A number of HG cDNA clones identified by the DNA probes JH8 or JH13 (including HG-4, HG-7C and 13A1) were sequenced. 5 Clones identified by JH8 were chosen because this genomic area is duplicated fewer times than the surrounding DNA, with only the HG-A and HG-B transcripts (not HG-C) homologous to this region. The comparison of these cDNA and genomic sequences showed a characteristic intron/exon pattern and we concluded that the exons highlighted in the genomic sequence were likely to be exons of the PKD1 gene. To prove this, pairs of primers matching the sequence of the putative PKD1 exons and spaced 0.7 - 2 kb apart in the proposed transcript, were synthesised. Employing RNA from 15 a radiation hybrid, HY145.19, that contains the PKD1 but not the HG loci, PKD1 specific cDNAs were amplified by RT-PCR and cloned (see Experimental Procedures for details). In this way, a number of overlapping cDNAs spanning the PKD1 transcript, for the cDNAs at the 3' end to those homologous to JH13 were cloned (Figure 13).

Analysis of a further cDNA, HG-6 showed that a short region (-100 bp) of HG-6 lay 5' to the sequenced genomic region and this was located by hybridisation to the genomic clone SM3 (figure 13); SM3 was subsequently sequenced. position of the cDNA in SM3 was identified and the possible 5' extent of this exon was determined in the genomic sequence; and in-frame stop codon was identified hear the 3' end of the exon. This exon lay at a CpG\_island (described

hereinafter) suggesting, along with the presence of the stop codon, that this may be the first exon of the PKDl gene. to determine the likely transcriptional start site the method of primer extension from three different oligos within the first exon was employed (see Experimental Procedures). In all cases, a transcriptional start was identified at the same G nucleotide and showed the first exon to be 426 bp. The structure of the PKDl transcript was confirmed by a final exon link, revl which starts 3 bp 3' to the proposed transcriptional start (see figure 13 and Experimental Procedures for details).

The intron/exon structure of the PKD1 gene

Sequencing the cDNA contig revealed a total sequence of 14, 148 bp which extends over approximately 52 bp of genomic sequence from SM3 to BFS5 (Figure 13). We were able to determine the intron/exon structure of much of the gene by direct comparison between the cDNA and genomic sequence. In the 3' region of the gene (JH5-BFS5), a partial genomic sequence was obtained at intron/exon borders by sequencing the corresponding genomic clone from exonic primer.

#### The PKD1 CpG island

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The 5' end of the gene lies at CpG island SM3. SM3 is located entirely within the duplicated region, but this clone was isolated from the cosmid SM11 which extends through the duplicated area into the proximal flanking single copy region and therefore is known to originate from this area. Figure 14 shows a map of the PKD1 CpG island including genomic sites for several methylation sensitive

enzymes, the location of the first exon and the GC content across the island. Evidence that the enzyme sites in the PKD1 region (and not just the HG area) digest, was obtained by pulsed field gel electrophoresis with the enzymes Mlu I, Not II and BssH II using probes outside the duplicated area. Digestion of the Sac II sites and confirmation of the Not I site was made with a panel of somatic cell hybrids which either contain just the HG (P-MWH2A) or just the PKD1 locus (Hy145.19). These results showed that the Sac II and Not I sites digest in both sets of hybrids (data not shown), indicating that this region is a CpG island in the HG as well as the PKD1 area. Further proof that this is the likely position of a functional promoter was obtained by analysis for DNAase 1 hypersensitivity. hypersensitive site in the region 5' to the transcription start site in SM3 was detected (figures 14a and b). Analysis of the PKD1 transcript.

Analysis of the sequence shows an open reading frame running from the start of the sequence to position 13,117 bp (Figure 15). Detailed sequencing of the genomic region containing the 3' portion of the gene revealed two extra Cs at positions 13,081-2 (Figure 15). An in-frame start codon which is consistent with the Kozak consensus was detected at position 212 bp; just 3' to the stop codon in the 5'UTR. Analysis for a signal sequence cleavage site using the von Hinge (von Hinge 1986) algorithm showed a high probability of a hydrophobic signal sequence with cleavage at amino acid 23 (see Figure 15). The total length of the predicted

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protein is 4302 as with a calculated molecular mass after excision of the signal peptide of 460 kD and an estimated isoelectric point of 6.26. However, this may be an underestimate of the total mass of the protein as many potential sites for N-linked glycosylation are present (Figure 15).

# Homologies with the PKD1 protein

The predicted PKD1 protein was analysed for homologies with know proteins in the SwissProt and NBRF databases using the BLAST Altschul et al 1990) and FASTA algorithms. This analysis revealed two clear homologies and also a number of other potential similarities which were studied on detail. Leucine rich repeat

Near the 5' end of the PKDl protein is a region of leucine rich-repeats (LRRs). LRRs are a highly conserved motif usually of 24 residues with precisely spaced leucines (or other aliphatic amino acids) and, an asparagine at position 19 (Figure 16a and reviewed in Kobe and Reisenhofer (1994)). Two complete LRRs plus a partial repeat unit are found in the PKDl protein, which have complete homology with the LRR consensus.

Surrounding the LRRs are distinctive cysteine-rich amino and carboxy flanking regions (Figures 16b and c). This flank-LRR-flank structure is exclusively found on proteins in extracellular locations and is thought to be involved in protein-protein interactions such as adhesion to other cells or to components of the extracellular matrix or as a receptor concerned with binding or signal transduction.

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The structure found in the PKD1 protein is similar to that found in the Drosophila protein, slit, which is important for normal central nervous system development (Rothberg, 1990). Although slit contains far more LRRs than the PKD1 protein, with four blocks each consisting of 4 or 5 repeat units, the structure of each block is similar as they finish on the amino and carboxy side with shortened LRRs which are immediately flanked by the cysteine rich regions. In the PKD1 protein two shortened LRRs surround one complete repeat unit and immediately abut the amino and carboxy flanking regions.

The amino flanking region consists of four invariant cysteines and a number of other highly conserved residues in an area of 30-40 amino acids; comparison of the PKD1 region to amino flanking motifs of other proteins is shown in figure 4b. The carboxy flanking region extends over an area of between 50-60 residues and consists of an invariant proline and four cysteines plus several other highly conserved amino acids. The similarity of the PKD1 region to carboxy flanking regions from other proteins is shown in figure 4c.

Some LRR proteins, such as slit (Rothberg 1990) and small proteoglycans are wholly extracellular but others including Toll (Hashimoto et al, 1990) and trkc (Lamballe 1991) have a single transmembrane sequence, while the LH-CRG receptor and related proteins have seven trans-membrane segments and are involved in signal transduction.

C type lectin domain

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Analysis of the sequence from exons 6 and 7 showed a high level of homology with a C type lectin domain. C type lectins are found in a variety of proteins in extracellular locations where they bind specific carbohydrates in the presence of Ca<sup>2</sup>+ ion (Drickamer 1987, 1988; Weiss 1992). Figure 17 illustrates the similarity of the PKD1 lectin domain to those found in a number of proteins including: proteogylcans, which interact with collagens and other components of the extracellular matrix; endocytic receptors, and selectins which are involved in cell adhesion and have been Three different selectins recognition. identified: E-selectin (endothelium), P-selectin (platelets) and L-selectin (lymphocytes) and these work with other cell adhesion molecules to promote binding of the cell 15 carrying the selectin to various other target cells: Immunoglobulin-like repeat motif

:Significant homologies were detected between a region Tof exon 5 and three regions of exon 15, with the same conserved sequence, WDFGDGS, which is also found in a melanocyte-specific secreted glycoprotein, Pmel17 (Kwon et al, 1991) and three prokaryotic collagenases or proteinases (Ohara et al, 1989, Takeuchi et al, 1992 and Matsushita et al, 1994). Further analysis of the amino acid sequence of the PKD1 protein showed that a conserved region of approximately 85 bp. could be discerned around this central sequence and that 16 copies of this repeat were present in the PKD1 protein; 1 in exon 5 and the other 15 as a tandem array in exons 11 to 15. Figure 18 shows that a highly

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conserved structure is maintained between the repeats although in some cases less similarity is noted with the WDFGDGS sequence. Further analysis of the most conserved residues found in the repeat units showed similarity to various immunoglobulin (Ig) domains; two Ig repeats which show particular homology to the PKD1 protein are shown (figure 18). The repeat unit is most similar to that found in a number of cell adhesion and surface receptors which have recently been defined as the I set of Ig domains (Harpaz 1994). Ig repeats consist of 7-9  $\beta$  strands of 5-10 residues linked by turns which are packed into two  $\beta$  sheets. The B, C, F and G  $\beta$ -strands of the I set are particularly similar to the PKD1 repeat, although the highly conserved cystine residues which stabilise the two  $\beta$  sheets through a 15 disulphide bond are absent. The D and E β strands, however, seem less similar and in some cases are significantly shortened or apparently absent. Further evidence that this PKD1 repeat has an Ig-like structure is found by analysis of the secondary structure with the predominant configuration found of  $\beta$  strands linked by turns. The WDFGDS area of the Ig molecule is one that often has a specific binding function (Jones et al., 1995) and this sequence may have a specific binding role in polycystin.

#### Type III fibronectin-related domains

Analysis of the secondary structure of the PKD1 protein beyond the carboxy end of the region of Ig-like repeats - showed a continuation of the β stand and turn structure. No evidence of further Ig-like repeats could be found in this

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area but three pairs of evenly spaced (38-40aa) tryptophan and tyrosine residues was noted which are the most highly conserved positions of the type III fibronectin repeat which has a similar secondary structure to Ig domains. Further analysis and comparison with other type III fibronectin domains showed that in total four fibronectin repeats (one with leucine replacing the conserved tyrosine) could be recognised in this area with many of the most highly conserved residues of this domain found in the PKD1 repeat (Figure 20)...

A large number of proteins with Ig-like repeats have now been described which are involved in cell-cell interactions and cell adhesion (reviewed in Brummendork and Rathjen, 1994), while type III fibronectin (FNIII) domains are found on extracellular matrix molecules and adhesion proteins. A number of cell adhesion proteins which are located mainly on neural cells, have both Ig-like and FNIIIrelated domains. In these cases the FNIII repeats are e always positioned C-terminal of the Ig-like units and close 20 to a transmembrane domain; a similar pattern is seen in the proposed structure of polycystin. These Ig/FNIII containing proteins such as neuroglican and NrCAM are thought to be involved in neuron-neuron interactions and the patterning of the axonal network. 

Many cell adhesion proteins of the Ig superfamily are also involved in communication and signal transduction mediated through their cytoplasmic tails. These cytoplasmic regions are known to bind to cytoskeletal proteins and other

intracellular components, and phosphorylation of this part of the molecule is also thought to affect adhesive properties of the protein; potential phosphorylation sites are found in the cytoplasmic tail and one intracellular loop of polycystin (Figure 20).

## Transmembrane regions

Analysis of hydrophobicity predicted that the deduced protein is an integral membrane protein with a signal peotide and multiple transmembrane (TM) domains located in the C-terminal region. From this analysis 11 regions 10 (including the signal peptide) had a mean hydrophobicity indice higher than 1.4 and therefore were considered as certain membrane spanning domains (see Experimental Procedures for details). Three others with a mean 15 hydrophobicity indice between 0.75-1.0 were considered as putative TM domains. The most likely topology of the protein was predicted using TopPed II programme (see Experimental Procedures for details) and the resulting model included one putative segment plus the transmembrane domains and the signal peptide. According to 20 this model the N-terminal end is extracellular and the (highly hydrophobic) carboxy-terminal region is anchored to the membrane by 11 membrane-spanning segments, with the highly charged carboxy end located in the cytoplasm. topology is supported by the study of N-glycosylation sites 25 with all but one site, out of a total of 61 predicted, in an extracellular location according to the model, including 11 in the two large extracellular loops between TM regions.

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However, if degree of hydrophobicity required to define a certain putative transmembrane region is altered within the model, the predicted number of such domains can change to 9 (excluding the most N-terminal pair) or 13 (with two new domains defined between TM7 and TM8). This can be ascertained by studies with specific antibodies.

Most transmembrane proteins containing the types of cell adhesion domain found on polycystin have a single transmembrane domain. The role of the multiple membrane 10 spanning domains found in polycystin is not yet clear. Proposed structure of the PKD1 protein

From the detailed analysis of the predicted PKD1 protein sequence a model of the likely structure of the protein can be formulated; (Figure 20). This model predicts 15 an extracellular N-terminal region of approximately 2550 aa containing several distinctive extracellular domains and an intracellular C-terminus of approximately 225 aa. \_\_intervening region of nearly 1500 aa is associated with the membrane with 11 transmembrane regions predicted and 10 variously sized extracellular and cytoplasmic loops (see A proline rich hinge is found between the flank-LRR-flank region and the first Ig-like repeat. Two phosphorylation sites for tyrosine kinase and protein kinase C are found in cytoplasmic locations (Figures 15 and 20).

25 Therefore, the PKD1 protein, named polycystin, has highlighted several clear domains, plus a reiterated motif that occupies over 30% of the protein.

Characterisation of the PKD1 gene has proven to be a

uniquely difficult problem because most of the gene lies in a region which is reiterated elsewhere on the chromosome. The high degree of similarity between the two areas (97%) both in exons and introns has meant that a novel approach has been required to clone the full length transcript; involving extensive genomic sequencing and generating cDNAs from a cell line with the PKDl but not the HG loci. In this way a contig containing the entire PKD1 transcript has now been cloned.

10 Preliminary analysis shows that the HG genes are very similar to PKD1 both in terms of genomic structure and sequence over most of their length (apart from the novel 3' regions). The 5' end of the PKD1 gene is at a CpG island which lies within the duplicated area. Homologous areas to this island, in the HG region, also have cleavable sites for methylation sensitive enzymes; these duplicate islands probably lie at the 5' ends of the various HG genes. Analysis for DNAase hypersensitivity also indicates that the HG, CpG islands probably contain active promoters. These results with the observation of are consistent polyadenylated mRNA from the HG genes on Northern blots and the similarity of the expression pattern of the HG and PKD1 genes in different tissue specific cell lines. The HG genes may have complete open reading frames and may encode 25 functional proteins. Antibodies to their 'unique' 3' regions will be required to determine this. Although the PKD1 transcript is large, the overall size of the gene, at 52 kb, is not (the Duchenne muscular dystrophy (DMD) gene which

encodes a slightly smaller transcript has a genomic size of over 2Mb). Indeed, if the first intron of PKD1 is excluded from the analysis, 40.3% of the remainder of the gene is found in the mature mRNA. In the compact structure of the PKD1 gene, some of the introns are close to or smaller than the minimal size of 80 bp thought to be required for efficient splicing, although they are presumably excised effectively. We have shown that deletion of 18 or 10 bp from one small intron (intron 43), resulting in an intron of 55 or 57 bp, leads to aberrant splicing (Peral, 1995). Similar mutations may be found in the other small introns of this gene. The compact nature of the PKD1 gene probably reflects the GC rich area of the genome in which it is found .... (the PKD1 transcript has a total GC content of about 65%); e# 15 a similar organisation is seen in other genes from the area of chromosome 16 (Vyas, 1992) is in an AT rich genomic 1 2 3 30 region.

adhesion or recognition molecule with multiple different extracellular domains. These various binding domains are likely to have different specificities so that it can be envisaged that it will bind to a variety of different proteins (and carbohydrates) both on other cells and possibly in the extracellular matrix. Although provisional evidence indicates a wide range of expression of polycystin in tissue specific cell lines, detailed analysis by in situ of the mRNA and with antibodies to determine the cells expressing this protein both in adult tissue and during

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development will provide further evidence.

Initial analysis has revealed little clear evidence of alternate splicing; although one cDNA (out of 6 studied) had an extra exon of 255 bp positioned in intron 16. This exon contains an in-frame stop codon and it is not known at this stage if this represents an incompletely spliced mRNA or a splice form of polycystin which terminates at this point. Truncation of the protein here would leave a secreted protein lacking all of the transmembrane and cytoplasmic regions. Interestingly, a similar secreted form of the neural adhesion protein, NCAM, which is normally attached to the cell membrane, is produced by alternate splicing by insertion of an exon containing a stop codon (Gower et al., 1988).

The initial changes that have been noted in ADPKD kidneys are abnormal thickening and splitting of the basement membrane (BM) and simultaneous de-differentiation of associated epithelial cells at the point of tubular Similar results have been noted in the dilation. heterozygote Han: SPRD rat (Schafer et al., 1994) which is a dominant model of PKD, although it is not known if it is a rat model of PKD1. Concurrent changes in cellular characteristics and the BM suggests that a disruption or alteration of communication between the cell and the BM may be the primary change in this disease. Polycystin could play an important role in interaction and communication between epithelial cells and the BM. It is known that signals are required from cells to the extracellular matrix

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(ECM) for normal BM development and also that communication from the ECM to cells is required for control of cellular differentiation. Communication between the ECM and cells occurs by several different means including through integrins and so polycystin may bind to integrins, although it may interact directly with components of the ECM. Although ADPKD is generally a disease of adulthood, there is plenty of evidence that the cystic changes in the kidney may start much earlier (Milutinovic et al., 1970), even in utero (Reeders, 1986). Expression of polycystin during renal development may be when its major role occurs, perhaps in assembly of the BM and it is then that the errors, which later lead to cyst development, occur.

The plethora of connective tissue abnormalities

associated with ADPKD indicate that the adhesion/communication roles of polycystin may be important for assembly and/or maintenance of the BM in many tissues, as well as the kidney. Hence, it is possible that disruption of normal cell adhesion and communication mediated by polycystin may explain the primary defects seen in the kidney and other organs in ADPKD. Clearly molecules that interact with polycystin or have a similar role are candidates for the other renal polycystic diseases of man.

A study of the mutations of the PKD1 gene highlight important functional regions of the protein. All of the mutations described so far in typical PKD1 families involve deletion or other disruption in the 3' end of gene. Two large deletions detected on Southern blots remove a large

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part of the protein (or make an out of frame product) including the last 6 transmembrane domains and the Cterminal end. The in-frame splicing change described in the same paper would remove most of TM10 and part of the preceding cytoplasmic loop. Two recently described splicing mutations (Peral, 1995) create three different products which either delete part of the cytoplasmic loop between TM7 and TM8 or a larger region of this loop including part of TM7 or insert an extra region into that loop. These mutated genes may make functional protein (they all produce abnormal mRNA) and it is interesting to note that, in each case, these proteins would have an intact extracellular region with disrupted cytoplasmic and transmembrane areas. proteins may bind to extracellular targets but are unable to 15 communicate in a normal way.

A group of mutations of PKD1 which completely delete the gene and hence are clearly inactivating have been described (Brook-Carter, 1994). However, in each of these cases the deletions also disrupt the adjacent TSC2 gene making interpretation of these cases difficult (TSC2 mutations alone can cause the development of renal cysts). Nevertheless, the severity of the polycystic disease in these patients indicate that inactivation of one PKD1 allele does promote cyst development. Further more, all these children are often severely affected at birth, cyst formation must occur in utero in these cases and hence polycystin has an important developmental role. 'A second somatic hit in the target tissue may also be required in

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these cases (and normal PKD1 patients) before cyst development can occur. ""...

### PKD1 GENE AND POLYCYSTIC KIDNEY DISEASE

We have therefore compelling evidence that mutations of 5 the PKD1 gene give rise to the typical phenotype of ADPKD. The location of this gene within the PKD1 candidate region and the available genetic evidence from the families with mutations show that this is the PKD1 gene. The present invention therefore includes the complete PKD1 gene itself and the six PKD1 - associated mutations which have been described: a de novo translocation, which was subsequently : transmitted with the phenotype; two intragenic deletions (one a de novo event); two further deletions; and a splicing defect.

It has been argued that PKD1 could be recessive at the cellular level, with a second somatic mutation required to give rise to cystic epithelium (Reeders, 1992). This "two hit" process is thought to be the mutational mechanism giving rise to several dominant diseases, such as 20 neurofibromatosis (Legius, et al., 1993) and tuberous sclerosis (Green, et al., 1994) which result from a defect in the control of cellular growth. If this were the case, however, we might expect that a proportion of constitutional PKD1 mutations would be inactivating deletions as seen in 25 these other disorders.

The location of the PKD1 mutations may, however, reflect some ascertainment bias as it is this single copy area which has been screened most intensively for mutations.

Nevertheless, no additional deletions were detected when a large part of the gene was screened by FIGE, and studies by PFGE showed no large deletions of this area in 75 PKD1 patients. It is possible that the mutations detected so far result in the production of an abnormal protein which causes disease through a gain of function. However, it is also , possible that these mutations eliminate the production of functional protein from this chromosome and result in the PKD1 phenotype by haploinsufficiency, or only after loss of 10 the second PKD1 homologue by somatic mutation.

: At least one mutation which seems to delete the entire PKD1 gene has been identified (WS-53) but in this case it also disrupts the adjacent TSC2 gene and the resulting phenotype is of TSC with severe cystic kidney disease. Renal cysts are common in TSC so that the phenotypic significance of deletion of the PKD1 gene in this case is difficult to assess. It is clear that not all cases of renal cystic disease in TSC are due to disruption of the PKD1 gene; chromosome 9 linked TSC (TSC1) families also 20 manifest cystic kidneys and we have analysed many TSC2 patients with kidney cysts who do not have deletion of the PKD1 gene.

Preliminary analysis of the PKD1 protein sequence has highlighted two regions which provide some clues to the 25 possible function of the PKD1 gene. At the extreme 5' end of the characterised region are two leucine-rich repeats (LRRs) (amino acids 29-74) flanked by characteristic amino flanking (amino acids 6-28) and carboxy flanking sequences

(amino acids 76-133) (Rothberg et al., 1990). LRRs are thought to be involved in protein-protein interations (Kobe and Deisenhofer, 1994) and the flanking sequences are only found in extracellular proteins. Other proteins with LRRs flanked on the amino and carboxy sides are receptors or are involved in adhesion or cellular signalling. Further 3' on the protein (amino acids 350-515) is a C-type lectin domain (Curtis et al., 1992). This indicates that this region binds carbohydrates and is also likely to be extracellular. These two regions of homology indicate that the 5' part of the PKD1 protein is extracellular and involved in proteinprotein interactions. It is possible that this protein is a constituent of, or plays a role in assembling, the extracellular matrix (ECM) and may act as an adhesive 15 protein in the ECM. It is also possible that the extracellular portion of this protein is important in signalling to other cells. The function of much of the PKD1 \* protein is still not fully known but the presence of several hydrophobic regions indicates that the protein may be threaded through the cell membrane.

Familial studies indicate that de novo mutations probably account for only a small minority of all ADPKD cases; a recent study detected 5 possible new mutations in 209 families (Davies, et al., 1991). However in our study one of three intragenic muttions detected was a new mutation and the PKDl associated translocation was also a de novo event. Furthermore, the mutations detected in the two familial cases do not account for a significant proportion

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of the local PKD1. The OX875 deletion was only detected in 1 of 282 unrelated cases, and the splicing defect was seen in only 1 of 48 unrelated cases. Nevertheless, studies of linkage disequilibrium have found evidence of common 5 haplotypes associated with PKD1 in a proportion of some populations (Peral, et al., 1994; Snarey, et al., 1994) suggesting that common mutations will be identified.

Once a larger range of mutations characterised it will be possible to evaluate whether the type and location of mutation determines disease severity, and if there is a correlation between mutation and extrarenal manifestations. Previous studies have provided some evidence that the risk of cerebral aneurysms 'runs true' in families (Huston, et al., 1993) and that some PKD1 families exhibit a consistently mild phenotype (Ryynanen, et al., 1987). A recent study has concluded that there is evidence of anticipation in ADPKD families, especially if the disease is transmitted through the mother (Fink, et al., 1994). Furthermore, analysis of families with early manifestations of ADPKD show that there is a significant intra-familial recurrence risk and that childhood cases are most often transmitted maternally (Rink, et al., 1993; Zerres, et al., 1993). This pattern of inheritence is reminiscent of that seen in diseases in which an expanded trinucleotide repeat 25 was found to be the mutational mechanism (reviewed in Mandel, 1993). However, no evidence for an expanding repeat correlating with PKDl has been found in this region although such a sequence cannot be excluded.

There is ample evidence that early presymptomatic diagnosis of PKD1 is helpful because it allows complications such as hypertension and urinary tract infections to be monitored and treated quickly (Ravine, et al., 1991). The identification of mutations within a family allow rapid screening of that and other families with the same mutation. However, genetic linkage analysis is likely to remain important for presymptomatic diagnosis. The accuracy and ease of linkage based diagnosis will be improved by the identification of the PKD1 gene as a microsatellite lies in the 3' untranslated region of this gene (KG-8) and several CA repeats are located 5' of the gene (see Figure 1a and 6; Peral, et al., 1994; Snarey, et al., 1994).

# Experimental Procedures

# 15 Clinical Details of Patients

# Family 77

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77-2 and 77-3 are 48 and 17 years old, respectively and have typical ADPKD. Both have bilateral polycystic kidneys and 77-2 has impaired renal function. Neither patient manifests any signs of TSC (apart from cystic kidneys) on clinical and ophthalmological examination or by CT scan of the brain.

77-4 is 13 years old, severely mentally retarded and has multiple signs of tSC including adenoma sebaceum, depigmented macules and periventricular calcification on CT scan. Renal ultrasound reveals a small number of bilateral renal cysts.

#### ADPKD patients

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OX875 developed ESRD from ADPKD, aged 46. Progressive decline in renal function had been observed over 17 years; ultrasound examinations documented enlarging polycystic kidneys with less extensive hepatic cystic disease. Both kidneys were removed after renal transplantation and pathological examination showed typical advanced cystic disease in kidneys weighing 1920g and 340g (normal average 120g).

OX114 developed ESRD from ADPKD aged 54: diagnosis was

10 made by radiological investigation during an episode of
abdominal pain aged 25. A progressive decline in renal
function and the development of hypertension was
subsequently observed. Ultrasonic examination demonstrated
enlarged kidneys with typical cystic disease, with less

15 severe hepatic involvement.

ADPKD in which several members have developed ESRD. The patient himself has been observed for 12 years with progressive renal failure and hypertension following ultrasonic demonstration of polycystic kidneys.

No signs of TSC were observed on clinical examination of any of the ADPKD patients.

DNA Electrophoresis and Hybridisation.

DNA extraction, restriction digests, electrophoresis,

Southern blotting, hybridisation and washing were performed
by standard methods or as previously described (Harris, et
al., 1990). FIGE was performed with the Biorad FIGE Mapper
using programme 5 to separate fragments from 25-50 kb. High

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molecular weight DNA for PFGE was isolated in agarose blocks and separated on the Biorad CHEF DRII apparatus using appropriate conditions.

## Genomic DNA probes and somatic cell hybrids

Many of the DNA probes used in this study have been described previously: MS205.2 (D16S309; Royle, et al., 1992); GGG1 (D16S259; Germino, et al., 1990); N54 (D16S139; Himmelbauer, et al., 1991); SM6 (D16S665), CW23, CW21, and JH1 (European Chromosome 16 Tuberous Sclerosis Consortium, 1993). Microsatellite probes for haplotype analysis were KG8 and W5.2 (Snarey, et al., 1994)SM6, CW3 and CW2, (Peral, et al., 1994), 16AC2.5 (Thompson, et al., 1992); SM7 (Harris, et al., 1991), VK5AC (Aksentijevich, et al., 1993).

JH6, 11 kb, 6 kb and 6 kb BamH I fragments, respectively, and JH13 and JH14, 4 kb and 2.8 kb BamH I-EcoR I fragments, respectively, all from the cosmid JH2A; JH8 and JH10 are 4.5 kb and 2 kb Sac I fragments, respectively and JH12 a 0.6 Sac I-BamH I fragment, all from JH4; 8S1 and 8S3 are 2.4 kb and 0.6 kb Sac II fragments, respectively, from JH8; CW10 is a 0.5 kb Not I-Mlu I fragment of SM25A; JH17 is a 2 kb EcoR I fragment of NM17.

The somatic cell hybrids N-OH1 (Germino, et al., 1990),
P-MWH2A (European Chromosome 16 Tuberous Sclerosis
Consortium, 1993) and Hy145.19 (Himmelbauer, et al., 1991)
have previously been described. Somatic cell hybrids
containing the paternally derived (BP2-10) and maternally
derived (BP2-9) chromosomes from OX114 were produced by the

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method of Deisseroth and Hendrick (1979).

Constructing a cosmid contig

Cosmids were isolated from chromosome 16 specific and total genomic libraries, and a contig was constructed using the methods and libraries previously described (European Chromosome 16 Tuberous Sclerosis Consortium, 1993). To ensure that cosmids were derived from the 16p13.3 region (not the duplicate 16p13.1 area) initially, probes from the single copy area were used to screen libraries (e.g. CW21 and N54). Two cosmids mapped entirely within the area duplicated, CW10III and JC10.2B. To establish that these were from the PKD1 area, they were restriction mapped and hybridised with the probe CW10. The fragment sizes detected were compared to results obtained with hybrids containing only the 16p13.3. are (Hy145.19) or only the 16p13.1 region (P-MWH2A).

#### FISH

FISH was performed essentially as previously described (Buckle and Rack, 1993). The hybridisation mixture contained 100 ng of biotin-II-dUTP labelled cosmid DNA and 2.5 mg human Cot-1 DNA (BRL), which was denatured and annealled at 37°C for 15 min prior to hybridisation at 42°C overnight. After stringent washes the site of hybridisation was detected with successive layers of fluorescein-conjugated avidin (5 mg/ml) and biotinylated ani-avidin (5 mg/ML) Vector Laboratories). Slides were mounted in Vectashield (Vector Laboratories) containing 1 mg/ml propidium iodide and 1 mg/ml 4', 6-diamidino-2-phenylindole

(DAPI), to allow concurrent G-banded analysis under UV light. Results were analysed and images captured using a Bio-Rad MRC 600 confocal laser scanning microscope.

 $\Phi^{(1)}(x,y) = (x_1 + y_1) + (y_1 + y_2) + (y_2 + y_3) + (y_1 + y_2) + (y_2 + y_3) + (y_3 + y_3) + (y_4 + y_3) + (y_4 + y_3) + (y_4 + y_4) +$ 

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cDNA screening and characterisation.

Foetal brain cDNAs libraries in A phage (Clonetech and Stratagene) were screened by standard methods with genomic fragments in the single copy area (equivalent to CW23 and CW21) or with a 0.8 kb Pvu II-Eco RI single copy fragment of AH3. Six PBP cDNAs were characterised: AH4 (1.7 kb) and 3A3 (2.0 kb) are described in European Chromosome 16 Tuberous Sclerosis Consortium, 1993, and four novel cDNAs AH3 (2.2 kb), AH6 (2.0 kb), A1C (2.2 kb) and B1E (2.9 kb). Striatum library (Stratagene) was screened with JH4 and a HG-C cDNA, 11BHS21 (3.8 KB) WAS ISOLATED, 21p.9 is a 0.9 kb Pvu II-EcoR I subclone of this cDNA. A HG-A or HG-B cDNA, . HG-4 (7 kb) was also isolated by screening the foetal brain library (Stratagene) with JH8...HG-4/1.1 is a 1.1 kb Pvu-II-15 EcoR I fragment from the 3' end of HG-4. 1A1H.6 is a 0.6 kb Hind III-EcoR I subclone of a TSC2 cDNA, 1A-1 (1.7 kb), which was isolated from the Clonetech library. Each cDNA was subcloned into Bluescript and sequenced utilising a combination of sequential truncation and liigonucleotide primers using DyeDeoxy Terminators (Applied Biosystems) and an ABI 373A DNA Sequencer (Applied Biosystems) or by hand with 'Sequenase' T7 DNA polymerase OUSB).

#### RNA Procedures

Total RNA was isolated from cell lines and tissues by the method of Chomczynskiand Sacchi (1987) and enrichment for mRNA made using the PolyAT tract mRNA Isolation System (Promega). For RNA electrophoresis 0.5% agarose denaturing formaldehyde gels were used which were Northern blotted,

hybridised and washed by standard procedures. The 0.24 - 9.5 kb RNA (Gibco BRL) size standard was used and hybridisation of the probe (1-9B3) to the 13 kb Utrophin transcript (Love, et al., 1989) in total fibroblast RNA was used as a size marker for the large transcripts.

RT-PCR was performed with 2.5 mg of total RNA by the method of Brown et al. (1990) with random hexamer primers, except that AMV-reverse transcriptase (Life Sciences) was employed. To characterise the deletion of the PBP transcript in OX114 we used the primers:

AH# F95' TTT GAC AAG CAC ATC TGG CTC TC 3'.

AH3 B75' TAC ACC AGG AGG CTC CGC AG 3'

in a DMSO containing PCR buffer (Dode, et al., 1990) with 0.5 mM MgCl<sub>2</sub> and 36 cycles of: 94°C, 1 min; 61°C, 1 min; 70°C, 2 min plus a final extension of 10 min. The 3A3 C primers used to amplify the OX32 cDNA and DNA were:

3A3 C15' CGC CGC TTC ACT AGC TTC GAC 3' S

These were employed in a PCR buffer and cycle 20 previously described (Harris, et al., 1991) with lmM  $MgCl_2$  and an annealing temperature of 61°C.

PCR products for sequencing were amplified with Pfu-1 (Stratagene) and ligated into the Srf-1 site in PCR-Script (Stratagene) in the presence of Srf-1.

## 25 RNAse protection

Tissues from normal and end-stage polycystic kidneys
were immediately homogenised in guanidinium thiocyanate.

RNA\_was purified on a cesium chloride gradient and 30 mg

total RNA was assayed by RNAse protection by the method of Melton, et al., (1984) using a genomic template generated with the 3A3; C primers.

#### Heteroduplex Analysis

Heteroduplex analysis was performed essentially as described by Keen et al. (1991). Samples were amplified from genomic DNA with the 3A3, C primers, heated at 95°C for 5 minutes and incubated at room temperature for at least 30 minutes before loading on a Hydrolink gel (AT Biochem).

Hydrolink gels were run for 12-18 hours at 250V and 10 fragments observed after staining with ethidium bromide.

## Extraction and amplification of paraffin-embedded DNA

DNA from formalin fixed, paraffin wax embedded kidney tissue was prepared by the method of Wright and Manos 15 (1990), except that after proteinase K digestion overnight at 55°C, the DNA was extracted with phenol plus chloroform before ethanol precipitation. Approximately 50 ng of DNA was used for PCR with 1.5 mM MgCl, and 40 cycles of 94°C for 1 min, 50°C for 1 min and 72°C for 40 s, plus a 10 min extension at 72°C.

The oligonucleotide primers designed to amplify across the genomic deletion of OX875 were:

5' - GGG CAA GGG AGG ATG ACA AG - 3'

JH14B3 : 5' - GGG TTT ATC AGC AGC AAG CGG - 3'

which produced a product of about 220 bp in individuals with the OX875 deletion.

- 3' RACE analysis of WS-212
  - RACE was completed essentially as described

(European Polycystic Kidney Disease Consortium (1994)).

Reverse transcription was performed with 5µg total RNA with 0.5µg of the hybrid dT. adapter primer using conditions previously described (Fronman et al. (1988)). A specific 3' RACE product was amplified with the primer F5 and adapter primer in 0.5mM MgCl, with the program: 57°C, 60s; 72°C, 15 minutes and 30 cycles of 95°C, 40s; 57°C, 60s; 72°C, 60s plus 72°C, 10 minutes. The amplified product was cloned using the TA cloning system (Invitrogen) and sequenced by conventional methods.

## Genomic and cDNA Probes and somatic cell hybrids

The genomic clones CW21, JH5, JH6, JH8, JH10. JH12,

JH13 and JH14 and the cDNAs A1C, AH3, 3A3 and AH4 are

described herein. Newly described probes are: SM3 a 2.0 kb

BamH 1 subclone of the cosmid SM11, JH9, 2.4kb Sac 1

fragment and JH11, 1.2kn Sac 1 - BamH1 fragment, both from

JH4. See Eur. Polycystic Kidney Disease Consortium, 1994

and Eur. Chromosome 16 Tuberous clerosis Consortium 1993 for

all above clones. DFS5 is a 4.2 kb Not 1-Hind 111 fragment

of CW23 (Eur. Chromosome 16 Tuberous Sclerosis Consortium,

1993). The cDNAs; BPG4, BPG6, BPG7C and 13-A were isolated

from a fetal brain cDNA library in A phage (Stratagene) and

are 7 kb, 2 kb, 4.5 kb and 1.2 kb respectively.

The somatic cell hybrids have previously been described, P-MWH2A (Eur., Chromosome 16 Tuberous Sclerosis Consortium, 1993) and Hy145.19 (Himmelbauer et al., 1991).

Total cellular RNA from the radiation hybrid Hyl45.19

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reverse transcribed using random hexamers (Eur. Polycystic Kidney Disease Consortium, 1994). This material was used as a template for PCR using the proof reading polymerase Pfu-1 with the primer pairs described in Table 2. The resultant products were cloned into the Srf-1 site of pPCRscript (SK+) plasmid. Sequencing

Full length sequence was obtained from the genomic clones, HG cDNAs and exon link clones using the progressive 10 unidirectional deletion technique of Henikoff, (1984). Both strands were then sequenced using DyeDeoxy Terminator Cycle Sequencing and an Applied Biosystems Sequencer 373A. Contig assembly was done using the programmes Assembly line (vs 1.0.7), SeqEd (vs 1.03) and MacVector (4.1.4).

#### Primer Extension

Primer extension was performed on total cellular fibroblast RNA. 25µg of RNA was annealed at 60°C in the .. presence of 400mM NaCl 0.01pM to, of HPLC. oligonucleotide which had been end labelled to a specific 20 activity of 3 x 10 cpm/pM with P. Primer extension was then performed in the presence of 50mM Tris pH8.2, 10mM DTT, 6mM MgCl<sub>2</sub>, 25mg/ml Actinomycin D. 0.5mM dNTPs, and 8 units of AMV reverse transcriptase. The extension reaction was continued for 60 min at 42°C. The extension products were compared to a sequencing ladder generated using the same . primer on the genomic clone SM3. The primers used were: N2765:5'-GGCGCGGCGGCGCATCGTTAGGGCAGCG-3' N5496:5'-GGCGGGCGCATCGTTAGGGCAGCGCGCGC-3'

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N5495:5'-ACCTGCTGCTGAGCGACGCCCGCTCGGGGC-3'.

## Analysis of sequence homology

The predicted PKD1 protein was analyzed for homologies with known proteins in the SwissProt and NBRF database using the BLAST (Altschul et al., 1990) and FASTA (Pearson et al., 1988) algorithms. Layouts were prepared by hand and using the programme Pileup.

#### Transmembrane regions

Potential transmembrane segments were identified by the method of Sipos and von Heljne (Sipos et al., 1993), using the GES hydrophobicity scale (Engelmen et al., 1986) and a trapezoid sliding window (a full window of 21 residues and a core window of 11 residues), as recommended. Candidate transmembrane domains were selected on the basis of their average hydrophobicity  $\langle H \rangle$ , and were classified as certain ( $\langle H \rangle \geq 1.0$ ) or putative (0.6,  $\langle H \rangle <1$ ).

The best topology for the protein was predicted on the basis of three different criteria: a) the net charge difference between the 15 N-terminal and the 15 C-terminal residues flanking the most N-terminal transmembrane segment (Hartmann et al., 1989); b) the difference in positively charged residues between the two sides of the membrane in loops smaller than 60 residues, and c) the analysis of the overall amino acid composition of loops longer than 60 residues by the compositional distance method (Nakashima et al., 1992). Using the above criteria the TopPred II program (Sipos wt al., 1993) calculated all the possible topologies of the proteins including the certain transmembrane segments

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and either included or excluded each of the putative segments to determine the most likely structure.

#### PKD1 Protein Purification

The PKDl protein may be purified according to conventional protein purification procedures well known in the art. Alternatively, the protein may be purified from cells harboring a plasmid containing an expressible PKDl gene. For example, the protein may be expressed in an E.coli expression system and purified as follows.

Cells are grown in a 10 liter volume in a Chemap Fermentor (Chemapec, Woodbury, NY) in 2% medium. Fermentation temperature may be 37°C, pH 6.8, and air as provided at 1 vvm. Plasmid selection may be provided using ampicillin for a plasmid containing an ampicillin resistance gene. Typical yield (wet weight) is 30 g/l.

For cell lysis, 50g wet cell weight of E.coli containing the recombinant PKD1 plasmid may be resuspended in a final volume of 100ml in 50 mM Tris-HC1 pH 8.0, 5 mM EDTA, 5mM DTT, 15 mM mercaptoethanol, 0.5% triton X-100, and 5 mM PMSF. 300 mg lysozyme is added to the suspension, and incubated for 30 min at room temperature. The material is then flyzed using a BEAD BEATER (R) (Biospec Products, Bartlesville, OK) containing an equal volume of 0.1-0.15 um glass beads. The liquid is separated from the beads and the supernatant removed, the pellet dissolved in 20 mM Tris-C1 pH 8.0.

The protein may be purified from the supernatant using DEAE chromatography, was is well known in the art.

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Preparation of Antibodies.

Antibodies specific for PDK1 protein or a fragment thereof are prepared as follows. A peptide corresponding to at least 8 amino acid residues of the PKD1 sequence of Fig. 15, are synthesized. Coupling of the peptide to carrier protein and immunizations is performed as described (Dymecki, S.M., J. Biol. Chem 267:4815-4823, 1992). Rabbit antibodies against this peptide are raised and sera are titered against peptide antigen by ELISA. The sera exhibiting the highest titer (1:27,000) are most useful.

Techniques for preparing monoclonal antibodies are well known, and monoclonal antibodies of this invention may be prepared by using the synthetic polypeptides of this invention, preferably bound to a carrier, as the immunogen as was done by Arnheiter et al., Nature, 294, 278-280 (1981).

Monoclonal antibodies Tare typically obtained from hybridoma tissue cultures or from ascites fluid obtained from animals into which the hybridoma tissue was introduced.

Nevertheless, monoclonal antibodies may be described as

being "raised to" or "induced by" the synthetic polypeptides of this invention or their conjugates with a carrier.

Antibodies are utilized along with an "indicating group" also sometimes referred to as a "label". The indicating group or label is utilized in conjunction with the antibody as a means for determining whether an immune reaction has taken place, and in some instances for determining the extent of such a reaction.

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The indicating group may, be a single atom as in the case of radioactive elements such as iodine 125 or 131, hydrogen 3 or sulfur 35, or NMR-active elements such as fluorine 19 or nitrogen 15. The indicating group may also be a molecule such as a fluorescent dye like fluorescein, or an enzyme, such as horseradish peroxidase (HRP), or the

The terms "indicating group" or "label" are used herein to include single atoms and molecules that are linked to the antibody or used separately, and whether those atoms or molecules are used alone or in conjunction with additional reagents. Such indicating groups or labels are themselves well-known in immunochemistry and constitute a part of this invention only insofar as they are utilized with otherwise 15 novel antibodies, methods and/or systems. Detection of PKD1 and Subcellular Localization.

Another embodiment of this invention relates to an assay for the presence of PKD1 protein in cells. Here an above-described antibody is raised and harvested. antibody or idiotype-containing polyamide portion thereof is then admixed with candidate tissue and an indicating group. The presence of the naturally occurring amino acid sequence is ascertained by the formation of an immune reaction as signaled by the indicating group. Candidate tissues include any tissue or cell line or bodily fluid to be tested for the presence of PKD1. . .

Metabolic labeling : immunoprecipitation, immunolocalization assays are performed in cells

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described previously (Furth, M.E., et al., Oncogene 1:47-58, 1987; Laemmli, U.K., Nature 227:680-685, 1970; Yarden, Y., et al., EMBO J. 6:3341-3351, 1987; Konopka, J.B., et al., Mol. Cell. Biol. 5:3116-3123, 1985). For immunoblot 5 analysis, total lysates are prepared (using Fruth's lysis buffer) (Fruth, M.E., et-al., Oncogene, 1:47-58, 1987). Relative protein concentrations are determined with a colorimetric assay kit (Bio-Rad) with bovine serum albumin the standard. A protein of lysate containing 10 approximately 0.05 mg of protein is mixed with an equal sample buffer containing 2 volume of 2 SDS. mercaptoethanol, boiled for 5 min., fractioned on 10% polyacrylamide-SDS gels b(Konopka, J.B., et al., J.Virol., 51:223-232, 1984): and (stransferred to immunobilon polyvinyldine difluoride (Millipore Corp., Bedford, MA) 15 Protein blots lare treated with specific antipeptide antibodies (see below). Primary binding of the PKD1-specific antibodies is detected using anti-IgG second antibodies conjugated to horseradish peroxidase subsequent chemiluminescence development 20 blotting system (Amersham International).

For metabolic labeling, 10<sup>6</sup> cells are labeled with 100 µCi of <sup>35</sup>S-methionine in 1 ml of Dulbecco's modified Eagles medium minus methionine (Amersham Corp.) for 16h. Immunoprecipitation of PKDl protein from labeled cells with antipeptide antiserum is performed as described (Dymecki, S.M., et al., supra). Portions of lysates containing 10<sup>7</sup> cpm of acid-insoluble <sup>35</sup>S-methionine are incubated with 1 µg of

the antiserum in 0.5 ml of reaction mixture.

Immunoprecipitation samples are analyzed by SDSpolylarcylamide gel electrophoresis and autoradiography.

For immunolocalization studies, 10<sup>7</sup> CMK cells are resuspended in 1 ml of sonication buffer (60mM Tris-HCl, pH 7.5, 6 mM EDTA, 15 mM EGTA, 0.75M sucrose, 0.03% leupeptin 12mM phenylmethylsulfonyl fluoride, 30 mM 2-mercaptoethanol). Cells are sonicated 6 times for 10 seconds each and centrifuged at 25,000 xg for 10 min at 4°C.

The pellet is dissolved in 1 ml of sonication buffer and centrifuged at 25,000 x g for 10 min at 4°C.

The pellet (nucleus fraction) is resuspended in 1 ml of sonication buffer and added to an equal volume of 2 x SDS sample buffer. The supernatant obtained above (after the 15 first sonication) is again centrifuged at 100,000 x g for 40 min at 4°C. The supernatant (cytosolic fraction) is removed and added to an equal volume of 2 x concentrated SDS sample buffer. The remaining pellet (membrane fraction) is washed and dissolved in sonication buffer and SDS sample buffer as Protein samples are analyzed by described above. electrophoresis on 10% polyacrylamide gels, according to the -Laemmli method (Konopka, J.B., supra). The proteins are transferred from the gels on a 0.45-µm polyvinylidine difluoride membrane for subsequent immunoblot analysis. Primary binding of the PKD1 specific antibodies is detected. using anti-IgG second antibodies conjugated to horseradish peroxidase.

- For immunohistochemical localization of PKD1 protein,

CMK cells or U3T3 are grown on cover slips to approximately 50% confluence and are washed with PBS (pH 7.4) after removing the medium. The cells are prefixed for 1 min at 37°C in 1% paraformaldehyde containing 0.075% Triton X-100, rinsed with PBS and then fixed for 10 min with 4% paraformaldehyde. After the fixation step, cells are rinsed in PBS, quenched in PBS with o.1 and finally rinsed again in PBS. For antibody staining, the cells are first blocked with a blocking solution (3% bovine serum albumin in PBS) 10 and incubated for 1 h at 37°C. The cells are then incubated for 1 h at 37°C with antiserum (1:100 dilution or with preimmune rabbit serum (1:100). After the incubation with the primary antibody, the cells are washed in PBS containing 3% bovine and serum albumin and 0.1% Tween 20 and incubated for 1 h at 37 C in fluorescein-conjugated donkey anti-rabbit IgGs (Jackson Immunoresearch, Maine) diluted 1:100 in blocking solution. and the second of the

is added to each coverslip before mounting on glass slides and sealing with clear nail polish. All glass slides examined with a Zeiss Axiophot microscope.

An indicating group or label is preferably supplied along with the antibody and may be packaged therewith or packaged separately. Additional reagents such as hydrogen peroxide and diaminobenzideine may also be included in the system when an indicating group such as HRP is utilized. Such materials are readily available in commerce, as are many indicating groups, and need not be supplied along with

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the diagnostic system. In addition, some reagents such as hydrogen peroxide decompose on standing, or are otherwise short-lived like some radioactive elements, and are better supplied by the end-user.

Pharmaceutical Compositions of the Invention; Dosage and Administration

Pharmaceutical formulations comprising PKD1 nucleic acid or protein, or mutants thereof, can be prepared by For example, procedures well known in the art. injectables, e.g., liquid solutions or suspensions. Solid forms for solution in, or suspension in, a liquid prior to injection also can be prepared. Optionally, the preparation also can be emulsified. The active ingredient can be mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredient. For example, water, saline, dextrose, glycerol, ethanol, etc. or combinations thereof. Also useful are wetting or emulsifying agents, pH buffering agents or adjuvants. PKDl protein or DNA can be administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. each case, the active protein or the nucleic acid will be present in the range of about 0.05% to about 10%, preferably in ther ange of about 1-2% by weight. Alternatively, the active protein or the nucleic acid will be administered at a dosage of about 10mg-2kg/kg body weight, preferably 50mg-400mg/kg/body weight. Administration may be daily, weekly,

or in a single dosage, as determined by the physician.

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### OTHER EMBODIMENTS

Other embodiments will be evident to those of skill in the art. It should be understood that the foregoing detailed description is provided for clarity only and is merely exemplary. The spirit and scope of the present invention are not limited thereto, being defined by the claims set forth below.

#### REFERENCES

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#### CLAIMS

- 1. An isolated nucleic acid sequence comprising:-
  - (a) a PKD1 gene or its complementary strand,
- (b) a sequence substantially homologous to a substantial portion of a molecule defined in (a) above, or
  - (c) a fragment of a molecule defined in (a) or (b) above.
  - 2. A sequence according to claim 1, wherein the PKD1 gene has the nucleic acid sequence according to Figure 15.
- 10 3. A sequence according to claim 1, wherein the PKD1 gene has the partial nucleic acid sequence according to Figure 7.
  - 4. A sequence according to claim 1, wherein the PKD1 gene has the partial nucleic acid sequence according to Figure 10.
- 15 5. An isolated nucleic acid selected from the group consisting of:
  - (a) [OX114] a nucleic acid including a deletion of 446 base pairs between residues 1746-2192 as defined in Figure 7;
- 20 (b) [OX32] a nucleic acid including a deletion of 135 base pairs between residues 3696-3831 as defined in Figure 7;
  - (c) [OX875] a nucleic acid wherein about 5.5kb flanked by the two Xbal sites shown in Figure 3a are deleted and the

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EcoRl site separating the CW10 (41kb) and JH1 (18kb) fragments is thereby absent;

- (d) (WS-53) a nucleic acid including a deletion of about 100kb encompassing the PKD1 gene, wherein the 3' end of the deletion lies between the JH1 and CW21 fragments and the 5' end of the deletion lies between the SM6 and JH17 fragments shown in Figure 6;
- (e) (461) a nucleic acid wherein about 18 base pairs are deleted in the 75 base pair intron amplified by the primer pair 3A3C insert at position 3696 of the 3' sequence as shown in Figure 11;
- (f) (OX1054) a nucleic acid wherein about 20 base pairs are deleted in the 75 base pair intron amplified by the primer pair 3A3C insert at position 3696 of the 3' sequence as shown in Figure 11;
- (g) (WS-212) a nucleic acid including a deletion of about 75kb downstream of the PKDl gene and located between fragments SM9 and CW9 distal of the PKDl gene and the PKDl 3'UTR proximal to the PKDl gene as shown in Figure 12;
- 20 (h) (WS-215) a nucleic acid including a deletion of about 160kb encompassing the PKD1 gene, wherein the deletion extends 3' of the PKD1 gene to within fragment CW15 and 5' of the PKD1 gene to between fragments CW10 and CW36 as shown in Figure 12;
- 25 (i) (WS-227) a nucleic acid including a deletion of about 50kb encompassing the PKD1 gene, wherein the deletion extends 3' of the PKD1 gene to within fragment CW20 and 5' of the PKD1 gene to within fragment JH11 as: shown in Figure

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12;

- (j) (WS-219) a nucleic acid including a deletion of about 27kb encompassing a portion of the PKDl gene, wherein the deletion extends 3' of the PKDl gene within fragment JHl and into the PKDl gene to within fragment JH6 as shown in Figure 12;
  - (k) (WS-250) a nucleic acid including a deletion of about 160kb encompassing the PKDl gene, wherein the deletion extends 3' of the PKDl gene to within fragment CW20 and 5' of the PKDl gene to within fragment BLu24 as shown in Figures la and 12; and
  - (1) (WS-194) a nucleic acid including a deletion of about 65kb encompassing the PKDl gene, wherein the deltion extends 3' of the PKDl gene to within fragment CW20 and 5' of the PKDl gene to within fragment CW10.

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- 6. An isolated nucleic acid according to any preceding Claim, wherein the molecule is an RNA transcript comprising a sequence complementary to the coding region of the nucleic acid sequence according to Fig. 15 and comprising a length of about 14 KB.
- 7. An isolated nucleic acid according to claim 5 comprising an RNA transcript.

8. An isolated nucleic acid according to claim 6 comprising an RNA transcript.

9. A nucleic acid probe comprising 10 nucleotides complementary to 10 consecutive nucleotides of the PKD1 sequence according to Figure 15.

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- 10. A nucleic acid probe according to claim 9 wherein said probe is between 15 nucleotides and 14 kb in length.
  - 11. A nucleic acid probe according to claim 10, said probe being between 100 nucleotides and 5 kb in length.

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12. A recombinant expression vector comprising the isolated nucleic acid according to claim 10.

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10 13. A host cell comprising the vector of claim 12.

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- 14. A recombinant expression vector comprising the isolated hucleic acid according to claim 5.
  - 15. A recombinant expression vector comprising the isolated nucleic acid according to claim 7.
- 15 16. An isolated polypeptide comprising a PKD1 protein having the amino acid sequence according to Fig. 15.
  - 17. An isolated polypeptide comprising a PKDl protein fragment having the amino acid sequence according to Fig. 7.
  - 18. An isolated polypeptide comprising a PKD1 protein

fragment having the amino acid sequence according to Fig.

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- 19. An isolated polypeptide comprising a PKD1 protein fragment having an amino acid sequence comprising the amino acid sequence according to Fig. 7 and the amino acid residue deletions defined by the nucleotide deletions of claim 5, parts (a), (b) and (j).
- 20. An immunoglobulin molecule having specificity for PKDl protein, said protein comprising the amino acid sequence according to any one of Figures 7, 10 or 15.

- 21. A method for screening a subject to determine whether said subject is a PKD1-associated disorder carrier or has a PKD1-associated disorder, which method comprises detecting the presence or absence of PKD1 nucleic acid in a biological sample from said subject; wherein detection of a mutant or absent PKD1 nucleic acid is indicative of a PKD1-associated disorder.
- 22. A method for screening a subject to determine whether said subject is a PKDl-associated disorder carrier or has a PKDl-associated disorder, which method comprises detecting the presence or absence of PKDl polypeptide in a biological sample from said subject, wherein detection of a mutant or absent PKDl polypeptide is indicative of a PKDl-associated disorder.

23. A method according to claim 21, comprising detecting a genomic fragment comprising the PKD1 gene or a portion thereof, a genomic fragment comprising a flanking region of the PKD1 gene or PKD1 RNA.

- 5 24. A method according to claim 23, wherein said detection comprises hybridizing a PKDl nucleic acid probe to nucleic acid from said biological sample and comparing the results thereof with results obtained using a biological sample from a subject who is not a carrier of a PKDl-associated disorder.
  - 25. A method according to claim 25, wherein said detection includes applying a nucleic acid amplification process to said nucleic acid to amplify a fragment of the PKD1 nucleic acid.

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26. A method according to claim 26, wherein said nucleic acid amplification process comprises amplifying a fragment of PKD1 nucleic acid utilizing a set of primers selected from the group consisting of:-

AH3 F9 : AH3 B7 \* 5

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20 3A3 C1 : 3A3 C2

2 AH4 F2 : JH14 B3.

27. A method according to claim 24 wherein said detection step comprises digesting nucleic acid from said biological sample to EcoRl fragments and hybridising with a DNA probe

which hybridises to the restriction fragment in Figure 3(a) or 12.

- 28. A method according to claim 27, wherein nucleic acid from said biological sample is digested with EcoR I and said DNA probe is selected from the group consisting of the probes CW10, JH14, JH5, JH6, JH4, JH13, JH8, JH11 and CW36 identified in Figures 3a and 12.
- 29. A method according to claim 28 which comprises digesting said nucleic acid to provide BamH I fragments and hybridising with a DNA probe which hybridises to the BamH I fragment identified (B) in Figure 3(a).
  - 30. A method according to claim 30, wherein said DNA probe comprises the DNA probe 1A1HO.6 identified herein.

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31. A method of treating a patient afflicted with a PKD15 associated disorder comprising administering a nucleic acid
sequence according to any of claims 1 to 8.

- 32. A method of treating or preventing a PKD1-associated disorder which method comprises administering to a patient in need thereof a PKD1 gene having the sequence according to Figure 15 so as to permit expression of PKD1 protein.
  - 33. A method of treating or preventing a PKD1-associated disorder which method comprises administering to a patient

in need thereof a mutated PKDl gene isolated from WS212 DNA so as to permit expression of PKDl protein.

- 34. A diagnostic kit for amplifying a portion of the PKD1 gene, comprising a pair of nucleic acid primers complementary to a portion of the PKD1 nucleic acid sequence according to Fig. 15, and packaging means therefore.
- 35. A diagnostic kit according to claim 34, wherein the nucleic acid primers comprise one or more of the following sets:

10 AH3 F9 : AH3 B7;

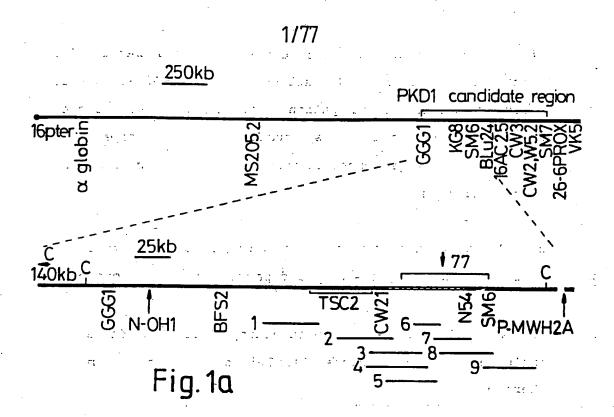
3A3 C1 : 3A3 C2; and

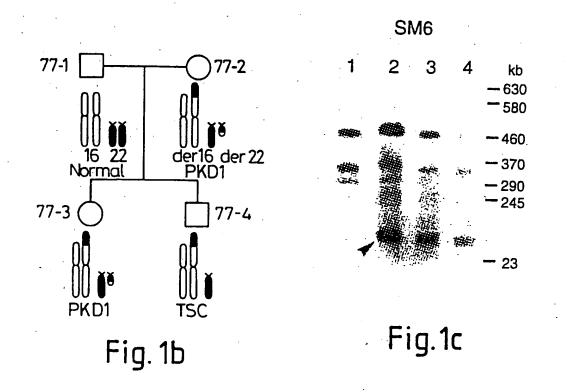
Kubur Nil Seli i Bashka jir katemati ni memberat ke mademi

AH4 F2 : JH14 B3.

- 36. A diagnostic kit for carrying out a method for determining whether said subject is a PKD1-associated disorder carrier or a patient having a PKD1-associated disorder, which kit includes a nucleic acid probe capable of hybridising to a sequence according to claim 1.
- 37. A diagnostic kit for carrying out a method for determining whether said subject is a PKD1-associated disorder carrier or a patient having a PKD1-associated disorder, which kit includes a nucleic acid probe capable of hybridising to a sequence according to claim 6 and packaging means therefore.

- 38. A diagnostic kit for carrying out a method for determining whether said subject is a PKD1-associated disorder carrier or a patient having a PKD1-associated disorder, which kit includes a nucleic acid probe capable of hybridising to a sequence according to claim 5 and packaging means therefore.
  - 39. A diagnostic kit for detecting PKD1 nucleic acid, including the DNA probe CW10 and packaging means therefore.
- 40. A diagnostic kit for detecting PKD1 nucleic acid,
  10 including the DNA probe 1A1HO.6 and packaging means
  therefore.





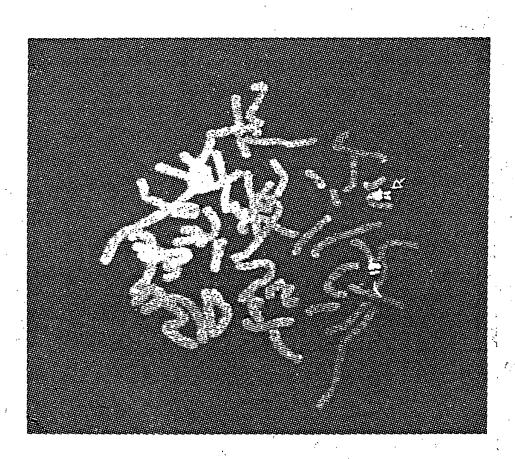
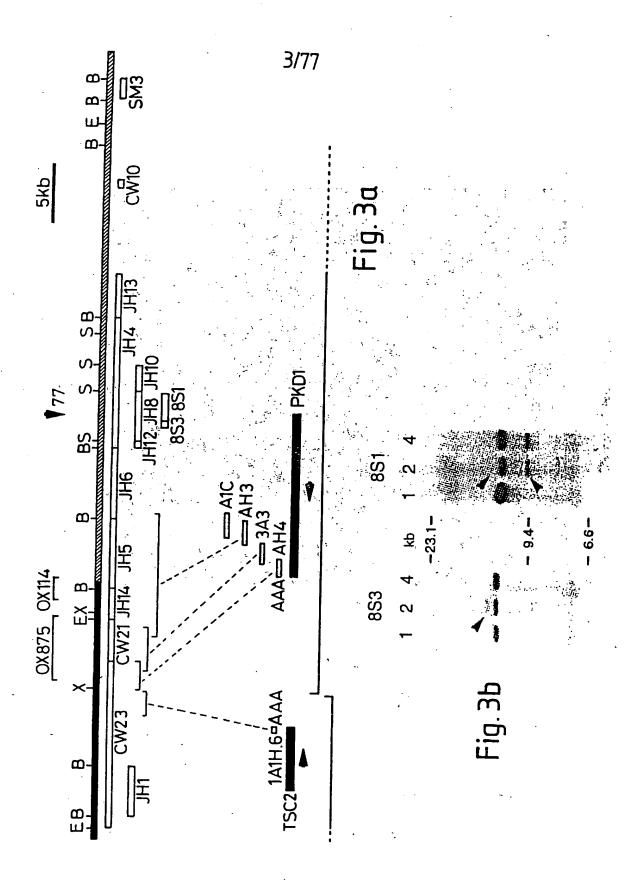


Fig. 2



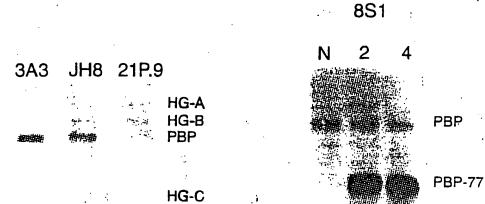
SUBSTITUTE SHEET (RULE 26)

3A3

1 2 3 4 5 6 7 8 9 kb

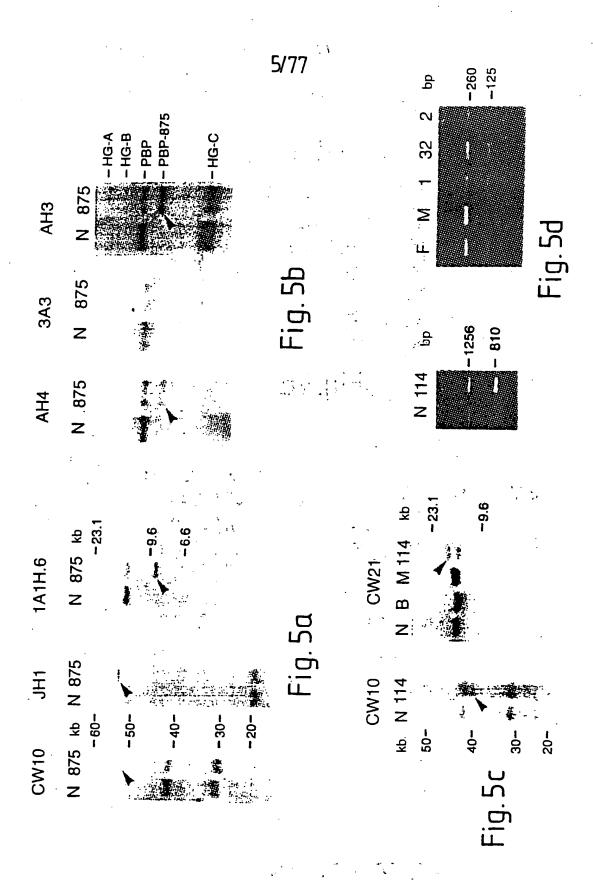


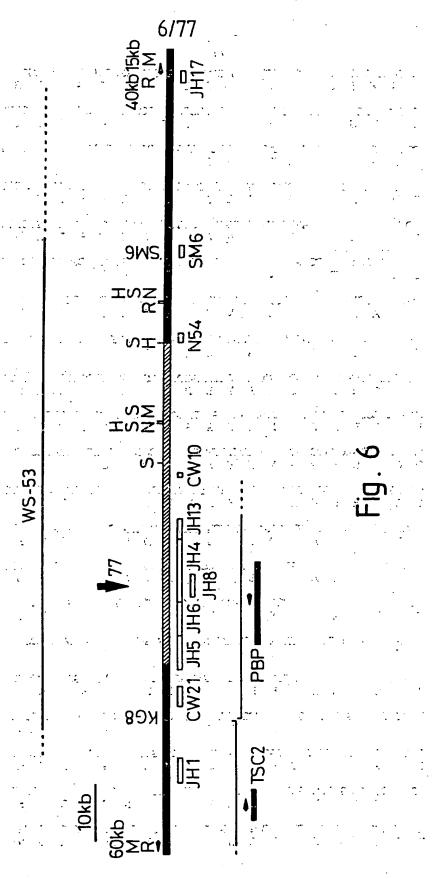
Fig. 4a



\* Fig.4b -

Fig.4c





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1	CTCAACCAGGAGCCCCTGACCCTGGCCGGCGAGGATCCTTGGCCCAGGGCAAGCGCTCG	60
1	LNEEPLTLAGEEIVAQGKRS	20
61 21	GACCOGOGAGOCTGCTGTGCTATGGCGGCCCACGCCCACGCCCTGCCTGCC	120 40
121	CCCCAGGCTTTCAGCGGGCCCTGGCCAACCTCAGTGACGTGGTGCAGCTCATCTTTCTG	180
41	P E A F S G A L A N L S D V V Q L I F L	60
181 61	GIGGACTCCAATCCCTTTCCCTTTGGCTATATCAGCAACTACACCGTCTCCACCAAGGTG V D S N P F P F G Y I S N Y T V S T K V	240 80
241 81	GOCTOGATGGCATTCCAGACACAGGCCGCCCGCCCAGATCCCCATCGAGCGGCTGGCCTCA A S M A F Q T Q A G A Q I P I E R L A S	300 100
301	GAGOCCCCATCACOTTGAACGTGCCCAACAACTCCGACTGGGCTGCCCGGGCCACCGC	360
101	ERAITVKVPNNSDWAARGHR	120
361 121	AGCTCCGCCAACTCCGCCAACTCCGTTGTGTGCCAGCCCCAGGCCTCCGTCGGTGCTGTG S S A N S A N S V V V Q P Q A S V G A V	420 140
421	GTCACCCTGGACAGCAACCCTGCGGCCGGCTGCATCTGCAGCTCAACTATACGCTG	480
141	V T L D S S N P A A G L H L Q L N Y T L	160
481 161	CTGGACGCCACTACCTGTCTGAGGAACCTGAGCCCTACCTGCAGTCTACCTAC	540 180
541 181	GAGCCCCGCCCAATGAGCACACTGCTCGCCTAGCAGGAGGATCCGCCCAGAGTCACTC EPRPNEHNCSASRRIRPESL	600 200
601	CAGGGTGCTGACCACCGGCCCTACACCTTCTTGATTTCCCCGGGGGGGG	660
201	Q G A D H R P Y T F F I S P G S R D P A	220
661	GGGAGTTACCATCTGAACCTCTCCAGCCACTTCCGCTGGTCGCGCGTGTCCGTG G S Y H L N L S S H F R W S A L Q V S W	720 240
221		
721 241	GCCCIGIACACGICCCCIGICCCAGIACITICAGCGACGACACACGACGACACAGAG G L Y T S L C Q Y F S E E D M V W R T E	780 260
781 261	GGGCTGCTGCCCCTGGAGGAGACCTCGCCCCGCCACCTC G L L P L E E T S P R Q A V C L T R H L	840 280
341	ACCECCTTCEGCCCAGCCTCTTCGTGCCCCCAĀĠCCATGTCCGCTTTGTGTTTCCTGAG	900
281	TAFGASLFVPPSHVRFVFPE	300
901 301	COGACAGOGGATGIAAACIACATOGTCATGCTGACATGTGCTGTGTGCCTGGTGACCIAC PTADVNYIV	960 320
961	ATEGTCATEGCCCCATCCTCCACAACCTCGACCACTTCGATCCCACCCCCCCC	1020
321	M V M A A I L H K L D Q L D A S R G R A	340
021 341	ATCCCTTICTGTGGGCAGGGGGGGCTTCAAGTAGGAGATCCTGGTCAAGACAGGCTGG I P F C G Q R G R F K Y E I L V K T G W	1080 360
081	GCCCCCCCTCAGGTACCACCCCCACGTCGCCATCATCCTGTATGCGGTGGACAGCCCGG	1140
361	G R G S G T T A H V G I M L Y G V D S R	380
141 381	AGOSGCCACOGGCACCTGGACGCGACAGCCTTCCACCGCACAGCCTGGACATCTTC S G H R H L D G D R A F H R N S L D I F	1200 400
201	COCATOCCCACCOCCACACCCTGGGTACCCTTGTGGAACATCCCAGTGTGGCACCACACAC	1260
401	RIATPHSLGSVWK,IR,VWHDN	420

	0///	
1261 421	Figure 7 AAAGGGCTCAGCCCTGCTGCTGCAGCAGGTCATGGTCAGGGACCTGCAGAGGGCA K G L S P A W F L Q H V I V R D L Q T A	1320 440
1321 441	CSCAGOSCCITCTTCCTGGTCAATGACTGGCTTTCCGTGGAGACGGCCAACGGGGCCAACGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGCCAACGGGGGG	1380 460
1381 461	CIGGIGGAGAGGAGGIGCIGCCCGGGGAGGCAGCCCITTTGCGCTTCCCGCCCCTG	140 480
1441 481	CICGICGCICAGCICCAGCICGCTICITICACAAGCACATCICGCTCTCCATATCCGAC L V A E L Q R G F F D K H I W L S I W D	1500 500
1501 501	CCCCCCCTCGTAGCCCTTTCACTCCCATCCAGAGGGCCACCTGCTGCGTTCTCCTCATC	1560 520
1561 521	TGCCTCTTCCTGGGGCCCAACGCCGTGTGGGTACGGGCCTGTTGGCGACTCTGCCTACAGC C L F L G A N A V W Y G A V G D S A Y S	1620 540
1621 541	ACGGGGCATGIGICCAGGCIGAGCCCGGIGAGCGICGACACAGICGCTGITGGCCTCGIG T G H V S R L S P L S V D T V A V G L V	1680 560
1681 561	TOCAGORIGICIATOCOGICIACCIGGOCATCCTTTTTCTCTTCCCCATCTCCCCCC S S V V V Y P V Y L A I L F L F R M S R	1740 580
1741 581	AGCAAGGTGGCTGGGAGCCCCGAGCACACTGCCGGGCAGCAGGTGCTGGACATCGAC S K V A G S P S P T P A G Q Q V L D I D	1800 <b>600</b>
1801 601	AGCIGCCIGGACIGGICGGICGACAGCICCTTCCICAGGITCTCAGGCCTCCAGGCT S C L D S S V L D S S F L T F S G L H A	1860 620
1861 621	GACCCCTTTGTTCGACAGATGAAGAGTGACTTGTTTCTCGATGATTCTAAGAGTCTCGTG E A F V G Q M K S D L F L D D S K S L V	1920 640
1921 641	TGCTGGCCCTCCGGCAACGCTAGTTGGCCGCACCTGCTCAGTGACCCGTCCATT C W P S G E G T L S W P D L L S D P S I	1980 660
1981 661	CTCGCTAGCAATCTGCGCCAGCTGCCACGCGCCCAGGCCCAGGGCCCAGGCCAGGCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCAGGCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCAGGCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCCAGGCCAGGCCCAG	2040 680
2041 681	GAGGACGCTTCTCCCTGGCCAGCCCCTCCTCGCCTGCCAAATCCTTCTCAGCATCAGAT E D G F S L A S P Y S P A K S F S A S D	2100 700
2101 701	GAAGACCIGATCCAGGAGGICCTIGCCGAGGGGGGGGGGG	2160 720
2161 721	ACCCACATEGAAACEGACCIECICAGCAGCCIGICCAGCACTCCIEGGGAGAAGACAGAG T H M E T D L L S S L S S T P G E K T E	2220 740
2221. 741.	ACCCTGGCGCTGCAGACGCTGGGGGAGCTGGGGCCACCCCAGGCCTGAACTGGGAA T L A L O R L G E L G P P S P G L N W E	2280 760
2281 761	CAGCCCCAGGCAGGCTGTCCAGGACAGGACTGGTGGAGGGTCTGGGAAGGGCCTGQPQAARAGGACAGGACAGGACTGGTGGAGGGCTCTGGGAAGGGCCTG	2340 780
2341 781	CTGCCCGCCTGGTGTGCCTCCCTGGCCCACCGGTCAGCCTGCTCCTGGTGGCTGTGGCT L P A W C A S L A H G L S L L L V A V A	2400 800
2401 801	GIGGCIGICICAGGGIGGGIGGGIGGGGIGGGGGGGGGG	2460 820
2461 821	CIGIOCAGCAGCCAGCITCCIGGCCTCATTCCTCGGCTGGGAGCCACTGAAGGTCITG	2520 840

	Figure 7 soils a
l L	CTGGAAGCCCTGTACTTCTCACTGGTGGCCAAGCGCGTGCACCGGATGAAGATGACACC LEALYFSLVAKRLHPDEDT
	CTGGTAGAGCCCGGCTGTGAGCCCTGTGAGCCCACGTGTGCCCCGGGTACGCCACCC L V E S P A V T P V S A R V P R V R P P
-	CACGCCTTTGCACTCTTCCTGGCCAAGGAGGAGGCCGCAAGGTCAAGAGGCTACATGGC H G F A L F L A K E E A R K V K R L H G
	ATCCTCCCCACCCTCCTCCTCTTACATCCTTTTTCTCCTC
ļ L	GGGGATGCCTCATGCCCATGGGCACGCCTACCGTCTGCAAAGCGGCCATCAAGCAGGAGCTG
Ĺ	GDASCHGHAYRLQSAIKQEL
î 1	CACAGCOGGGCCTTCCTGGCCATCACGCGGTCTGAGGAGGTCTCGCCATGGATGG
	GIGCIGCIGCCCTACGTCCACGGAACCAGTCCAGCCCAGAGCTGGGGCCCCACGGCTG
1 1	V L L P Y V H G N Q S S P E L G P P R L
	R Q V R L Q E A L Y P D P P G P R V H T
L.	TGCTCGGCCCAGGAGGCCTTCAGCACCAGCGATTACGACGTTGGCTGGGAGAGTCCTCAC
•	C.S.A.A.G.G.F.S.T.S.D.Y.D.V.G.W.E.S.P.H.
	AATGGCTGGGGGATGTGGGGCCTATTCAGGGCGGATCTGCTGGGGGCATGGTCCTGGGGC N G S G T W A Y S A P D L L G A W S W G
	TOCTIFICOCTIFICAÇÃCACOCCCCCCTACCTCCACGACCTCCCCCTGACCCTGACCTCGACGAC
•	SCAVYDSGGYVQELGLSLEE
	S R D R L R F L Q L H N W L D N R S R A
	GTGTTCGTGGAGGTCAGGGCTACAGCCGGGGGGGGGGGG
	V F L E L T R Y S P A V G L H A A V T L
•	R L E F P A A G R A L A A L S V R P F A
L L	CIGOCOCCICACOCOCCICIOCICCICCICCICCICCICCICCICCICCIC
	TTOSCOTTGCACTTOSCOTTGGCOGAGGCCOGTACTTGGCACAGGGAAGGGCGCTGGCGC
	FAVHFAVAEARTWHREGRWR
	V L R L G A W A R W L L V A L T A A T A
	CIGGTACCCTCCCCACCTCCGTCCCCCTCACCCCAGTCGACCCGTTTCGTCCCCCCC
	LVRLAQLGAADRQWTRFVRG
	R P R R F T S F D Q V A H V S S A A R G
	CTGCCGCCTCCTCTTCCTGCTTTTGGTCAAGGCTGCCCAGCAGGTAGGCTTCGTG LAAS LLF LL V KAAQ H V R F V
	OCCAGTOGTOCTTTTGGCAAGACATTATGCCGAGCTCTGCCAGAGCTCCTGGGGGTC
	POWSVFGKTICCAGGGGGGC

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5101	Figure 7 C	<u>ont'd</u> ccccrcccc	• •		CCIGCCCIICC	ragg 516
5161	TGTGGTGGCG	TTATGGCAGCCC	EGCIGCIGCIT	GGATGCGAGCIT	GCCTTCCCCC	GGIG 522
5221	CIGGGGGCAC	AGCTGTCTGCC	AGGCACTCTCAT	CACCCCAGAGGC	CTÍGICATCCI	528
5281					ATCAGGTCTGG	
5341	CTAGCAGGAC	TAGGCATGTCAG	AGGACCCCAGG	GTGGTTAGAGGA	AAAGACTCCTC	CTGG 540
5401	: ':			• • • • • • • • • • • • • • • • • • • •	GIGGGGGGG	
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5461		. ",				
5521				·	CCCCCCAACCCC	CCC 558
5581	ACCAAGCAGA	CAAAGICAATAA	AAGAGCTGTCT	GACTGCAAAAAA	AAAAAA 5631	
				• •		
	1A1H0.6				1 .	: : :
1 61 121 181 241 301 361 421 481 541	AAGCITICGCA TIAOGAGTICGCA AGCGTIGGCCA CACGCAAAITA TOCAAGTICGA GOOGCTTACT GCACAGACTIC TOCTCGGTICG CITTIGGACGGT GAGGCACAGA	AGATOSTGTC TIGGCCTCACA TIGGCCGGCT CCAACCCCAG CAGCCGAGCC AGGACTTCAC ATTGCCTGTC	COSCACATC CCIACCICIG CACACCIGGC CACACTIGIG	TTTIGTOCAGE AGGANAGACA CTGCCCTTGG AGGGGCTCCA AAGGGGCTCC GTGCACCCTC TATICAGGTGG TCAGGCGGG ATAAAAGTCCT	TIGATUSTICAC TIGGAGGGCT TIGGCCCCACA ACCURACICA ACCURACICATA GCCACACACA GCCACACACACACACACACACACACAC	CCCCCTGGAC TGTGGACACC GATGGCCCTG TATCTACCCC CTGCGAGGAA CAAAGCCCCT GCGCCTCATC CTGCACTGGC CACAGACATA
•	Figure 8	) )	, , , , , , , , , , , , , , , , , , ,			
	CW10L	*			: :	
1 61 121 181	GIGGGGIG GGIGGGGIG GGIGGGGIG	GCACGTACGC GTACGTCCTC GCTCAGTGCC TG	TICIOGIGIG ACICCITITIG COCCUCATG	TGICACACGT TTCTTTTCAC TGGCACCCCC	GCCCATTCIT	GAACTOTTOG GACTOCCACT GCTGTTAGGT
•	<u>CW10R</u>					
1		CCCCCACGAG	CAGGGGAGAG	GCACCCAAGG	<b>T.</b> 12 11	
	Figure 9		•			

	Figure 7 cont'd 10///	
3781 1261	ACCITICGCCCICGICGICCICGCGGTAGCCTAGCCCAGCIGGCCATCCIGCICGICTCT T L G L V V L G V A Y A Q L A I L L V S	3840 1280
3841 1281	TCCTGTGTGGACTCCCTCTGGAGGGTGGCCCAGGCCCTGTTGGTGCTGTGCCCTGGGACT	3900 1300
3901 1301	GGCTCTCTACCCTGTGTCCTCCCGGGGGGCACCTGTCACCCCTGCTGTGTGTG	3960 1320
3961 1321	CTCTGGGCACTGGGGGGCTGTGGGGGGCTGTTATTCTCGGCTGGGGCLL W A LL R L W G A L R L G A V I L R W R	4020 1340
4021 1341	TACCACCCTTGCGTGGAGAGCTGTTACCGGCCGGCCTGGGAGCCCCACGACTACGAGATG Y H A L R G E L Y R P A W E P Q D Y E M	4080 1360
4081 1361	GIGGAGITGITCCIGCGCAGGCIGCGCCTCTGGATGGCCCTCAGCAAGGICAAGGAGITC V E L F L R R L R L W M G L S K V K E F	4140 1380
4141 1381	CGCCACAAAGICCCCITTCAAGGGATGGAGCCCCTGCCCTCTCCCTCCCAGGGGCTCC R H K V R F E G M E P L P S R S S R G S	4200 1400
4201 1401	AAGGTATCCCCGGATGTGCCCCCCCACCCACCCACCCACC	4260 1420
4261 1421	TOCTOCAGCCAGCTGGATGGGCTGAGGCTGGGCCCGGGACAAGGTGTGAG S S S Q L D G L S V S L G R L G T R C E	4320 1440
4321 1441	CCTCACCCTCCCACCCTCCTCACCCAGTTTGACCGACTC P.E.P.S.R.L.Q.A.V.F.E.A.L.L.T.Q.F.D.R.L	4380 1460
4381.: <sup>7</sup> 1461	AACCAGGCCACAGAGGAGCTCTACCAGCTGGAGCAGCTGCACAGCCTGCAAGGCCGC N Q A T E D V Y Q L E Q Q L H S L Q G R	4440 1480
4441 1481	AGGAGCAGCOGGGGGCCGGGCCAGCA: R S S R A P A G S S R G P S P G L R P A	4500 1500
4501 1501	CIGCCCAGCCCCTTGCCCCGGCCAGTCGGGGTTGTGGACCTGGCCCCAGCAGG L P S R L A R A S R G V D L A T G P S R	4560 1520
4561 1521	ACACCITOGGGCCAAGAACAAGGTCCACCCCAGCAGCACITAGICCITCCITCCTGGCGGG T P S G Q E Q G P P Q Q H L V L L P G G	4620 1540
4621 1541	GETEGGCOETGGAGTOGGACACOGCTCAGTATTACTTTCTGCOGCTGTCAAGGCC  G G P W S R S G H R S V L L S A A V K A	4689 0 1560
4681 1561	CACCCCCACGCAGAATCCCTCCACGTACCTTCCCCAGACACCACGCCACCCCCCATCTCTCT E G Q A E W L H V G S P E S R Q G H L S	4740 1580
4741 1581	GICTGTGGGCTTCAGCACTTTAAAGAGGCTGTGTGGCCAACCAGGACCCAGGGTCCCCTC  V C G L Q H F K E A V W P T R T Q G P L	4800 1600
4801 1601	CCCAGCTCCCTTGGGAAGGACACAGCAGTATTGGACGGTTTCTAGCCTCTGAGATGCTAA P S S L G K D T A V L D G F	4860 1620
4861	TITATITCCCCGAGTCCTCAGGTACAGCGGGGGGGGGCCCCCCCGGGCAGAT	4920
4921	GIOCCCACIGCTAAGGCTGCTGGCTTCAGGGAGGGTTAGCCTGCACCGCCCACCCTG	4980
4981	CCCCTAAGTTATTACCTCTCCAGTTCCTACCGTACTCCCTGCACCGTCTCACTGTGTGTC	5040
5041		5100

		EVY III	3.870	DECC	DT 778	TON-	cm	TD :		•	<b></b>		<b>.</b>	• •		
	•			DESC							_		_			
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					ı Arı										o val	
TO: Se:	C CA	C AA s As	C CN n Le 3	u Lei	ı Arı	G GCC g Ala	G CIO	GA( 1 Ası 40	C GIT o Val	Gly	CTO Lea	CTO Lea	G GO J Ala 4!	G AAC a Asi 5	CIC Leu	142
TO	G GO r Ala	G CT a Le 5	u Ala	A GAC a Glu	Le	g gat I Ast	T ATA	Sea	C AAC ASD	AAC AST	AAC Lys	F ATT	T TC: Sei	r ACC	TTA Leu	190
GAV Glu	GA: 1 G1: 6:	i Cl	A ATZ	A TTT e Phe	GC Ala	AAT AST 70	TTA Leu	TTT Phe	'AAT Asn	TTA Leu	AGI Ser 75	Glu	ATA 1 Ile	A AAC e Asn	CTG Leu	238
AGT Ser 80	Gly	AA( Asi	o cox	G TTT D Phe	GAC Glu 85	Cys	GAC Asp	TGI	Gly	CIG Leu 90	Ala	TCC Trp	CIC	Pro	CGA Arg 95	286
Trp	GCC Ala	GAC Glu	G GAC	G CAG 1 Gln 100	Glr	GIG Val	CGG Arg	GIG Val	GIG Val 105	Gln	Pro	GAG Glu	GCA Ala	Ala 110	ACG Thr	334
TGI Cys	GCI Ala	GC Gly	Pro 115	GCC Gly	TCC Ser	CIG	CCT Ala	GGC Gly 120	Gln	CT Pro	CIG	CIT	GQC Gly 125	ATC	OCC Pro	382
TIG	Leu	GAC Asp 130	Ser	Gly	TGT Cys	GT Gly	GAG Glu 135	GAG Glu	TAT Tyr	GIC Val	Ala	TCC Cys 140	CIC	CT Pro	GAC Asp	430
AAC Asn	AGC Ser 145	Ser	Gly	ACC Thr	GIG Val	GCA Ala 150	GCA Ala	GTG Val	TCC Ser	TTT Phe	TCA Ser 155	OCT Ala	CC Ala	CAC His	GAA Glu	478
CC Gly 160	CIG	Leu	CAG Gln	CCA Pro	GAG Glu 165	Ala	TGC Cys	AGC Ser	CC Ala	TTC Phe 170	TCC Cys	TTC Phe	TCC Ser	ACC Thr	GC Gly 175	526
CAG Gln	GC Gly	CTC	GCA Ala	Ala 180	CIC Leu	TCG Ser	GAG Glu	CAG Gln	GC Gly 185	TGG Trp	TGC Cys	CTG Leu	TGT Cys	GGG Gly 190	‱ Ala	574
CCC Ala	CAG Gln	Pro	TCC Ser 195	AGT Ser	GCC Ala	TCC Ser	TTT Phe	CCC Ala 200	TCC Cys	CTG Leu	TCC Ser	CTC Leu	TGC Cys 205	TOC Ser	Gly	622
OCC Pro	ccc Pro	CCA Pro 210	Pro	CCT Pro	CCC Ala	ccc Pro	ACC Thr 215	TGT Cys	AGG Arg	CCC Gly	Pro	ACC Thr 220	CTC	CTC Leu	CAG ( Gln	670
CAC	GTC Val 225	TTC Phe	CCT Pro	CCC Ala	TCC Ser	CCA Pro 230	GCG Gly	CCC Ala	ACC Thr	CIG Leu	GTG Val	GCG Gly	OCC Pro	CAC His	GGA Gly	718

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OCI Pro 240	Leu	Ala	C TCI Ser	Gly	CAG Gln 245	Leu	GCA Ala	Ala	TTC Phe	CAC His 250	Ile	Ala	Ala	Pro	Leu 255		766
CCI Pro	GIC Val	ACI Thr	Ala	ACA Thr 260	Arg	Trp	GAC Asp	Phe	GGA Gly 265	Asp	Gly	TOC Ser	Ala	GAG Glu 270	val	•	814
GAT Asp	Ala	Ala	GGG Gly 275	Pro	Ala	Ala	Ser	His	Arg	Tyr	Val	CIG Leu	Pro 285	ÇG1y	Arg		862
TAT Tyr	CAC	GIG Val 290	Thr	Ala	GTG Val	Leu	Ala 295	Leu	Gly	Ala	Gly	Ser	Ala	Leu	CIG Leu	• € .	910
		Asp	Val	Gln		Glu	,Ala					Leu			GIG Val	· :	. 958 :
	Pro								Ser			CTC Leu					1006
					Gly			Ala							CIG		1054
		Glu		Ala		Ala	Val	His		Leu	Cys	OCC Pro			ACG Thr		1102
			Pro					Cys				GIG Val 380			AAG Lys		1150
												TGG Trp					1198
									Val			TTC Phe		Val		• •	1246
CCG Arg	GTC Val	ACC Thr	Arg	AGC Ser 420	CTA Leu	GAC Asp	GIG Val	TCG Trp	ATC Ile 425	GJA CCC	TTC Phe	TCG Ser	ACT Thr	GIG Val 430	CAG Gln	<u>.</u> .	1294
		Glu		Gly			Pro		Gly			TTC Phe			Glu,		1342
												CCC Ala 460				;.	1390
							Thr					ACC Thr			TGC Cys	•	1438

#### 14/77 TCA GOG COG CAC AGC TAC GTC TGC GAG CTG CAG COC GGA GGC CCA GTG 1486 Ser Ala Pro His Ser Tyr Val Cys Glu Leu Gln Pro Gly Gly Pro Val 485 490 480 CAG GAT GOO GAG AAC CTC CTC GTG GGA GOO COO AGT GOO GAC CTG CAG 1534 Gln Asp Ala Glu Asp Leu Leu Val Gly Ala Pro Ser Gly Asp Leu Gln 500 505 510 OGA COO CTG AÓG COT CTG OCA CAG CAG GAC GOC CTC TCA GOC COG CAC 1582 Gly Pro Leu Thr Pro Leu Ala Gln Gln Asp Gly Leu Ser Ala Pro His 520 525 GAG COC GTG GAG GTC ATG GTA TTC COG GGC CTG OGT CTG AGC OGT GAA 1630 Glu Pro Val Glu Val Met Val Phe Pro Gly Leu Arg Leu Ser Arg Glu 535 GOO THE CHE ACE ACE GOO GAA THI GOO ACE CAG GAG CHE COO COO COO 1678 Ala Phe Leu Thr Thr Ala Glu Phe Gly Thr Gln Glu Leu Arg Arg Pro . 545 550 GOC CAG CTG CGG CTG CAG CTG TAC CGG CTC CTC AGC ACA GCA GGG ACC 1726 Ala Gln Leu Arg Leu Gln Val Tyr Arg Leu Leu Ser Thr Ala Gly Thr 565 1774 OOG GAG AAC GGC AGC GAG OCT GAG AGC AGG TOO OOG GAC AAC AGG ACC Pro Glu Asn Gly Ser Glu Pro Glu Ser Arg Ser Pro Asp Asn Arg Thr 580 585 CAG CTG GOO GOO TOO ATG OOA GOO GGA COO TGG TGC OOT GGA GOO 1822 Gln Leu Ala Pro Ala Cys Met Pro Gly Gly Arg Trp Cys Pro Gly Ala 600 605 595 AAC ATC TGC TTG CGG CTG GAC GCC TCT TGC CAC CCC CAG GCC TGC GCC 1870 Asn Ile Cys Leu Pro Leu Asp Ala Ser Cys His Pro Gln Ala Cys Ala 615 620 1918 AAT GGC TGC ACG TCA GGG CCA GGG CTA CCC GGG GCC CCC TAT GCG CTA Asn Gly Cys Thr Ser Gly Pro Gly Leu Pro Gly Ala Pro Tyr Ala Leu 630 TOG AGA GAG TITC CITC TITC TOO GIT GOO GOG GOG COO GOO GAG TAC 1966 Trp Arg Glu Phe Leu Phe Ser Val Ala Ala Gly Pro Pro Ala Gln Tyr 645 650 TOG GTC ACC CTC CAC GGC CAG GAT GTC CTC ATG CTC OCT GGT GAC CTC 2014 Ser Val Thr Leu His Gly Gln Asp Val Leu Met Leu Pro Gly Asp Leu 660 665 670 2062 GIT GOC TIG CAG CAC GAC GCT GOC CCT GOC GCC CTC CTG CAC TGC TCC Val Gly Leu Gln His Asp Ala Gly Pro Gly Ala Leu Leu His Cys Ser 675 680 COG GCT COC GGC CAC CCT GGT COC CAG GCC COG TAC CTC TCC GCC AAC 2110 Pro Ala Pro Gly His Pro Gly Pro Gln Ala Pro Tyr Leu Ser Ala Asn 690 695 COC TOG TOA TOG CTG COC CAC TTG COA GOC CAG CTG GAG GGC ACT TGG 2158 Ala Ser Ser Trp Leu Pro His Leu Pro Ala Gln Leu Glu Gly Thr Trp 710 715

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	Cys					Leu					Ala			CAG Gln		`.	2206
										Gly				Pro 750		*2	2254
			Val											CAC His			2302
CIC	TCC Ser	TGC Cys 770	AGC Ser	Phe	GAC Asp	Val	GIC Val 775	Ser	CCA Pro	GTG Val	Ala	GGG Gly 780	CIG Leu	OGG Arg	GTC Val		2350
														AAC Asn		٠	2398
				Leu										CC Ala			2446
CCT Ala	CCC Arg	TGG Trp	CCT Pro	CCC Gly 820	GC Gly	AGT Ser	GIC Val	Ser	GCC Ala 825	œc Arg	TTT Phe	GAG Glu	AAT ASD	GTC Val 830	Cys		2494
Pro	CCC Ala	CIG Leu	GIG Val 835	∞ Ala	ACC Thr	TTC Phe	GIG Val	000 Pro 840	G Gly	TCC Cys	œ Pro	TCC	GAG Glu 845	ACC Thr	AAC Asn	ar ser s	25 <b>4</b> 2
GAT Asp	ACC Thr	CTG Leu 850	TTC Phe	TCA Ser	GIG Val	GTA Val	GCA Ala 855	CTG Leu	œ Pro	TGG Trp	CIC Leu	AGT Ser 860	GAG Glu	GCG Gly	GAG Glu	5x 	2590
CAC His	GTG Val 865	GTG Val	GAC Asp	GTG Val	GTG Val	GIG Val 870	GAA Glu	AAC Asn	AGC Ser	Ala	AGC Ser 875	ŒG Arg	GCC Ala	AAC Asn	CIC Leu	· ·	2638
AGC Ser 880	CIG Leu	CCG Arg	GTG Val	ACG Thr	CCG Ala 885	GAG Glu	GAG Glu	CC Pro	ATC Ile	TGT Cys 890	Gly	CIC Leu	CCC Arg	CCC Ala	ACG Thr 895	• •	<b>268</b> 6
Pro	AGC Ser	CCC Pro :	GAG. Glu	GCC Ala 900	OGT Arg	GTA Val	CTG Leu	Gl'n	GGA Gly 905	Val GTC	CTA Leu	GIG Val	Arg	TAC Tyr 910	AGC Ser	•	2734
Pro	GTG Val	Val	GAG Glu 915	Ala	GJA CCC	TCG Ser	GAC Asp	ATG Met 920	A97 ĈIC	TTC Phe	ŒG Arg	TCG Trp	ACC Thr 925	ATC Ile	AAC Asn	• •	2782
GAC Asp	aag Lys	CAG Gln 930	TCC Ser	CTG Leu	ACC Thr	Phe	CAG Gln 935	Asn	GIG Val	orc Val	TTC Phe	AAT Asn 940	Val	ATT Ile	TAT Tyr		2830
CAG Gln	AGC Ser 945	œ Ala	ccc Ala	GTC' Val	TTC Phe	AAG Lys 950	CTC Leu	TCA Ser	CTĞ Leu	ACG Thr	CCC Ala 955	TCC Ser	AAC Asn	CAC His	GÎĞ Val	7.	2878

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		AC GTA ACC GTG	GAG CCG ATG AAC Glu Arg Met Asn	
			GTG CTG TCC CCC Val Leu Ser Pro 990	
Ala Thr Leu Va			GAC TCA GCT GTG Asp Ser Ala Val 1005	
	eu Trp Asn Phe Gl		CAG GCC CTC CAC Gln Ala Leu His 1020	
			CCA GAC CCC TCG Pro Asp Pro Ser 1035	
			ACC TAC CCT CCC Thr Tyr Ala Ala O	
			GCC TTC GAG AAC Ala Phe Glu Asn 1070	Leu
Thr Gln Gln Va			CTG CCC TCC GTG Leu Pro Ser Val. 1085	
	er Asp Gly Val Le		OG CC GTC ACC Arg Pro Val Thr 1100	
TAC CCG CAC CC Tyr Pro His Pr 1105	OC CTG CCC TCG CC TO Leu Pro Ser Pr 1110	n GGG GGT GTT TO Gly Gly Val	CTT TAC ACG TGG Leu Tyr Thr Trp 1115	GAC 3358 Asp
			CAG CCG CCT CCC Gln Pro Ala Ala )	
CAC ACC TAT GO His Thr Tyr Al	C TOG AGG GGC AC a Ser Arg Gly Th 1140	C TAC CAC GTG F Tyr His Val 1145	CCC CTG GAG GTC Arg Leu Glu Val 1150	Asn
Asn Thr Val Se	oc eer ees ees ee er Gly Ala Ala Al 155	C CAG GOG GAT a Gln Ala Asp 1160	GIG COC GIC TIT Val Arg Val Phe 1165	GAG 3502 Glu
GAG CIC CGC GG Glu Leu Arg Gl 1170	y Leu Ser Val As	C ATG AGC CTG op Met Ser Leu 75	CCC GTG GAG CAG Ala Val Glu Gln 1180	GCC 3550
CCC CCC GTG GT Ala Pro Val Va 1185	NG GTC AGC GCC GC al Val Ser Ala Al 1190	G GTG CAG ACG a Val Gln Thr	GGC GAC AAC ATC Gly Asp Asn Ile 1195	ACG : 3598 Thr

SUBSTITUTE SHEET (RULE 26)

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TC Tri 120	Thr	Phe	GAC Asp	ATG Met	Gly 120	Asp	Gly	ACC Thr	GIG Val	CIG Leu 1210	Ser	GJA GCC	Pro	GAG Glu	GCA Ala 1215		3646
	GIG Val				Tyr					Asn					Val		3694
	C CCC Ala			Pro					Äla					Val		,	3742
	TTC Phe		Leu					Val					Cys				3790
	CAG Gln 126	Pro					Thr					Gly			CCC Ala	• •	3838
	TAC Tyr O					Thr					Ser						.3886
	OGG Arg				Thr					Phe					The	5.1	3934
TTC	Pro	CIG Leu	CCC Ala 131	Leu	GTG Val	CIG Leu	Ser.	AGC Ser 1320	Arg	GIG Val	AAC Asti	AGG Arg	GCG Ala 1325	His	TAC Tyr	9, 1 12,	3982
	ACC Thr		Ile					Glu					Thr			<i>:</i> `	4030
	GAG Glu 134	Arg					Leu					Trp					4078
	cc Ala O					Pro					Trp						4126
GAG Glu	GAA Glu	CCC Ala	GCC Ala ;	CC Pro 1380	Thr.	OGT Arg	CCC Ala	Arg	GC Gly 1385	Pro	GAG Glu	Val Val	ACG Thr	TTC Phe 1390	Ilè		<b>4174</b>
TAC	CGA Arg	GAC Asp	CCA Pro 1395	Gly	TCC Ser	TAT Tyr	CIT Leu	GTG Vál 1400	Thr	GTC Val	ACC Thr	CCG Ala	TCC Ser 1405	yzu	yżu Yyc	· .·	4222
	TCT Ser		Ala					Leu					Glu			· .	<b>4270</b>
CIG Leu	GIC Val 1425	Thr	AGC Ser	ATC. Ile	aag Lys	GIC Val 1430	Asn	Gly Gly	TCC Ser	CIT Leu	666 Gly 1435	Leu	GJu GAG	CTG Leu	CAG Gln	,	4318

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	Pro					Ala		Gly			Arg				TAC Tyr 1455	5	4366
			CIG Leu		Asp			Trp		Glu						.· •	4414
				Ser					Thr					Gly	TCG		4462
			Ser					Trp					Val		Arg		4510
		Arg	Gly				Asn			Arg		Val			CIG Leu	• •	4558
	Gly		Val			Ser					Ala					٠	4606
CCC Arg	TAT Tyr	TCC Ser	TCG Trp	GIG Val 1540	Leu	Cvs	GAC Asp	Arg	TGC Cys 1545	Thr	ccc Pro	ATC Ile	CCT Pro	GGG Gly 1550	Gly	'.	4654
CCT Pro	ACC Thr	Ile	TCT Ser 1555	Tyr	ACC Thr	TTC Phe	CCC Arg	TCC Ser 1560	Val.	Gly	ACC Thr	TTC Phe	AAT Asn 156!	Ile	ATC Ile	7.	.4702
GIC Val	ACG Thr	Ala	GAG Glu )	AAC Asn	GAG Glu	Val	Gly	TCC Ser	CCC Ala	CAG Gln	Asp	AGC Ser 1580	Ile	TTC Phe	GIC Val	, , <b></b> ,	4750
TAT Tyż	CIC Val 158	Leu	CAG Gln	CIC Leu	ATA Ile	GAG Glu 1590	Gly	CIG Leu	Gln	GTG Val	GIG Val 1595	Gly	Gly	GC Gly	OGC Arg		<b>4798</b>
	Phe		ACC Thr			Thr	Val		Leu		Ala					ŕ	4846
			GIC Val		Tyr					Trp			Arg		Pro		4894
		Ala	GGC Gly 1635	Ser.		Lys			Ser					Glu			4942 ·
		Tyr	CAT His	Val		Leu		Ala					Gly			. *·.	4990
		Asp	TGC Çys				Phe]			Pro		Gly				<i>i</i> •	5038

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		l Thi					Pro					Thr				Leu 1695		5086
•						Gly					Val			Trp		Leu O		5134
					Ser					Glu					His	AGC Ser	•	5182
	TTC Phe	Pro	ACA Thr 173	Pro	GGC Gly	CIG	CAC	TTG Leu 173	Val	ACC Thr	ATG Met	ACG Thr	GCA Ala 174	Gly	AAC Asr	Pro		5230
			Ser					Val					Gln			GIG Val		5278
		Gly					Ala					Gly				.ccc Ala 1775	vii Vii	5326
						Pro					Leu					AAT Asn O		5374
	GIG Val	AGC Ser	TCG Trp	TGC Cys 179	Trp	GCT Ala	GIG. Val	OC. Pro	GC Gly 1800	Gly	AGC Ser	AGC Ser	aag Lys	OGT Arg 180	Gly	CCT Pro	3	5422
	CAT His	GTC Val	ACC Thr 1810	Met	GTC Val	TTC Phe	OCG. Pro	GAT ASP 1815	Ala	GC Gly	ACC Thir	TTC Phe	TCC Ser 1820	Ile	OG Arg	CIC		5470
			Ser					Trp					Tyr			ACG Thr		5518
	GCG Ala 1840	Glu	GAG Glu	OC. Pro	ATC Ile	GTG Val 1845	Gly	CIG Leu	GIG Val	Leu	TGG Trp 1850	Ala	AGC. Ser	AGC Ser	aag Lys	GIG : Val 1855		5566
			OCC Pro			Leu					Ile					Gly		5614
			GTC Val		Phe					Gly:					Glu			5662
	CTC Leu	CCC Pro	Gly 1890	Pro	CCT Arg	TTC Phe	Ser	CAC His 1895	Ser	TIC Phe	er Pro	CCC. Arg	GIC Val 1900	Gly	GAC Asp	CAC His	.•	5710
			AGC Ser			Gly		Asn			Ser		Ala				ςi.	5758

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	GIG Val 1920	Arg	ATC Ile	GTG Val	GIG Val	CIG Leu 192	Glu	CCC Ala	GIG Val	AGT Ser	GGG Gly 1930	Leu	CAG Gln	ATG Met	Pro	AAC Asn 1935		5806
•	TGC Cys	TGC Cys	GAG Glu	Pro	GGC Gly 1940	Ile	CCC Ala	ACG Thr	GGC Gly	ACT: Thr 1945	Glu	AGG Arg	'AAC Asn	TTC Phe	ACA Thr 1950	Ala Ala		5854
	CCC Arg	val GTG	CAG Gln	Arg 195	Gly	TCT Ser	œ Arg	GTC Val	ccc Ala 1960	Tyr	GCC Ala	TCG Trp	TAC Tyr	TTC Phe 196	Ser	CTG Leu		5902
-	CAG Gln	AAG Lys	GTC Val 1970	Gln	Gly	GAC Asp	TCG Ser	CIG Leu 1975	Val	ATC	CTG Leu	TCG Ser	GC Gly 1980	Arg	GAC Asp	GTC Val	c	5950
	ACC Thr	TAC Tyr 1985	Thr	CCC Pro	GTG Val	Ala	GCG Ala 1990	Gly	CIG Leu	TTG Leu	Glu	ATC Ile 1995	Gln	GIG Val	CCC Arg	‱″ Ala,	,	5998
٠	TTC Phe 2000	Asn	CCC Ala	CTG Leu	GC Gly	AGT Ser. 2005	Glu.	AAC Asn	CCC Arg	ACG Thr	CIG Leu 2010	Val	CTG Leu	GAG Glu	GTT Val	CAG Gln 2015		6046
	GAC Asp	GCC Ala	GIC Val	CAG Gln	TAT Tyr 2020	Val	GCC Ala	CIG Leu	CAG Gln	AGC Ser 2025	Gly	CC Pro	TCC Cys	TTC Phe	ACC Thr 2030	AAC Asn )	. •	6094
•	CCC Arg	TCG Ser	CCG Ala	CAG Gln 203		GAG Glu	CCC Ala	CCC Ala	ACC Thr 2040	Ser	OCC Pro	AGC Ser	ccc Pro	CGG Arg 2045	Arg	GIG Val		6142
	CCC Ala	TAC Tyr	CAC His 2050	Trp	GAC Asp	TTT Phe	GGG Gly	GAT Asp 2055	Gly	TCG Ser	OCA Pro	GCG Gly	CAG Gln 2060	Asp	ACA Thr	CAT Asp		<b>6190</b>
	GAG Glu	CCC Pro 2065	Arg	GCC Ala	GAG Glu	CAC His	TCC Ser 2070	Tyr	CIG Leu	AGG Arg	CCT Pro	GGG Gly 2075	Asp	TÁC Tyr	CGC Arg	GIG Val		6238
	CAG Gln 2080	Val	AAC Asn	GCC Ala	TCC Ser	AAC Asn 2085	Leu	CTG Val	AGC Ser	TTC Phe	TTC Phe 2090	Val	CCG Ala	CAG Gln	GCC Ala	ACG Thr 2095		6286
	GTG Val	ACC Thr	GTC Val	CAG Gln	GTG Val 2100	Leu	çcc Ala	TGC Cys	CCG Arg	GAG Glu 2105	Pro	GAG Glu	GIG Val	GAC Asp	GIG Val 2110	Val		6334
	CTG Leu	CCC Pro	CTĠ Leu	CAG Gln 211	GiG Val	ČTG Leu	ATG Met	CGG Arg	CGA Arg 2120	Ser	G]ru CÀG	OCC Arg	aac Asn	TAC Tyr 2125	Leu	GAG Glu	. ,	6382
₹	GCC Ala	CAC His	GTT Val 2130	Asp	CTG Leu	OCC Arg	GAC Asp	TGC Cys 2135	Vai	ACC Thr	TAC Tyr	CAG Gln	ACT Thr 2140	Glu	TAC Tyr	OCC Arg	:	6430
	TGG Trp	GAG Glu 2145	Val	TAT Tyr	CCC Arg	ACC Thr	CCC Ala 2150	Ser	TGC Cys	CAG Gln	CCG Arg	OG Pro 2155	Gly	OCC Arg	OCA Pro	Ala		6478

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	Ar						y Va					g Pro				G CIG 1 Leu 217		6526	
•					a Le						Cys					C GIG 1. Val 90		6574	
-				y Ası						ı Ser					ı ya.	G AOG 1 Thr		6622	
•	GT( Val	GCC Ala	22:	o Glu	ı Arç	CIO J. Lei	GTC 1.Val	Pro 221	Ile	ATT : Ile	GAC Glu	GGI Gly	Gly 222	Ser	TAC Tyr	C CCC C Arg		6670	
, ,	GIC Val	Trp 222	Sea	A GAC	C. ACA	Arg	GAC J. Asp 223	Leu	GIG Val	CIG Leu	GAT Asp	G1y 223	Ser	GAC Glu	TOX Ser	TAC Tyr		6718	
	GAC Asp 224	Pro	AAC Asr	CTG Leu	GAG Glu	GAC Asp 224	Gly	GAC Asp	CAG Gln	AOG Thr	Pro 225	Leu	AGT Ser	TTC	CAC His	TGG Trp 2255		6766	
ė., '	Ala	TGI Cys	Val	. Ala	TOG Ser 226	Thr	CAG Gln	Arg	Glu	GCT Ala 226	Gly	Gly	TGT Cys	CCC Ala	Leu 227	AAC Asn O	z, 7	6814	
	TTT	GGG Gly	Pro	227	-Gly	AGC Ser	AGC Ser	Thr	GIC Val 228	Thr	ATT Ile	Pro	OGG Arg	GAG G1u 228	Arg	CIG:	i tiri idd	6862	
.13	.çcc Ala	GCT Ala	GCC Gly 229	Val	GAG Glu	TAC	ACC Thr	TTC Phe 229	Ser	CIG Leu	ACC Thr	GIG Val	10G Trp 2300	Lys	Ala	Gly	·	6910	
•	Arg	AAG Lys 230	Glu	GAG .Glu	GCC Ala	ACC Thr	AAC Asn 2310	Gln	ACG Thr	GTG Val	CTG Leu	ATC Ile 231	Arg	AGT Ser	GC GLy	Arg œG		6958	
	GIG Val 2320	Pro	ATT	GIG Val	TCC Ser	TTG Leu 232	Glu	TGT Cys	GTG Val	TCC Ser	TGC Cys 2330	Lys	CCA Ala	CAG Gln	CCC Ala	GIG Val 2335	•	7006	
٠						Ser				TAC Tyr 2345	Leù					Leu		7054	
	AAT Asn	TCC Cys	AGC Ser	AGC Ser 235	Gly	TCC Ser	AAG Lys	CGA Arg	00G Gly 2360	Arg	TGG Trp	CCT Ala	OCA Ala	OGT Arg 2365	Thr	TTC . Phe		7102	
	AGC Ser	AAC Asn	AAG Lys 237(	Thr	CTG Leu	GIG Val	Leu	GAT Asp 2375	Glu	ACC Thr	ACC Thr	Thr	TCC Sér 2380	Thr	CC Cly	AGT Ser	` <b>.</b>	. 7150	
	GCA Ala	GGC Gly 2385	Met	CGA Arg	CTG Leu	GIG Val	CIG Leu 2390	Arg	CCG Arg	GC Gly	GIG Val	CIG Leu 2395	Arg	GAC Asp	ĆĵÀ ŒĈ	GAG Glu	• -	7198	

GGA TAC ACC TTC ACG CTC ACG GTG CTG GGC CGC TCT GGC GAG GAG GAG GAG GAG GAG GAG GAG GAG	<b>7246</b>
GCC TGC GCC TGC ATC GCC CTG TGC GCC AAC GCC GCG GCC GCly Cys Ala Ser Ile Arg Leu Ser Pro Asn Arg Pro Pro Leu Gly Gly 2420 2425 2430	7294
TCT TCC CCC CTC TTC CCA CTG CCC CCT GTG CAC GCC CTC ACC ACC AAG Ser Cys Arg Leu Phe Pro Leu Gly Ala Val His Ala Leu Thr Thr Lys 2435 2440 2445	7342
GTG CAC TTC GAA TGC AGG GGC TGG CAT GAC GGG GAG GAT GCT GGC GCC Val His Phe Glu Cys Thr Gly Trp His Asp Ala Glu Asp Ala Gly Ala 2450 2455 2460	<b>7390</b>
CCC CTG GTG TAC GCC CTG CTG CTG CGG CGC TGT CGC CAG GGC CAC TGC Pro Leu Val Tyr Ala Leu Leu Leu Arg Arg Cys Arg Gln Gly His Cys 2465 2470 2475	7438
GAG GAG TTC TGT GTC TAC AAG GGC AGC CTC TGC AGC TAC GGA GCC GTG Glu Glu Phe Cys Val Tyr Lys Gly Ser Leu Ser Ser Tyr Gly Ala Val 2480 2485 2490 2495	<b>74</b> 86
CTG CCC CCG GGT TTC AGG CCA CAC TTC GAG GTG GGC CTG GCC GTG GTG Leu Pro Pro Gly Phe Arg Pro His Phe Glu Val Gly Leu Ala Val Val 2500 2510	.7534
GTG CAG GAC CAG CTG GGA GCC GCT GTG GTC GCC CTC AAC AGG TCT TTG Val Gln Asp Gln Leu Gly Ala Ala Val Val Ala Leu Asn Arg Ser Leu 2515 2520 2525	7582
GOC ATC ACC CTC CCA GAG COC AAC GGC AGC GCA ACG GGG CTC ACA GTC Ala Ile Thr Leu Pro Glu Pro Asn Gly Ser Ala Thr Gly Leu Thr Val 2530 2535 2540	7630
TGG CTG CAC GGG CTC ACC GCT AGT GTG CTC CCA GGG CTG CTG CGG CAG Trp Leu His Gly Leu Thr Ala Ser Val Leu Pro Gly Leu Leu Arg Gln 2545 2550 2555	7678
GCC GAT CCC CAG CAC GTC ATC GAG TAC TCG TTG GCC CTG GTC ACC GTG Ala Asp Pro Gln His Val Ile Glu Tyr Ser Leu Ala Leu Val Thr Val 2560 2570 2575	7726
CTG AAC GAG TAC GAG COG GCC CTG GAC GTG GCG GCA GAG CCC AAG CAC Leu Asn Glu Tyr Glu Arg Ala Leu Asp Val Ala Ala Glu Pro Lys His 2580 2585 2590	7774
GAG CGG CAG CAC CGA GCC CAG ATA CGC AAG AAC ATC ACG GAG ACT CTG Glu Arg Gln His Arg Ala Gln Ile Arg Lys Asn Ile Thr Glu Thr Leu 2595 2600 2605	7822
GTG TCC CTG AGG GTC CAC ACT GTG GAT GAC ATC CAG CAG ATC GCT GCT Val Ser Leu Arg Val His Thr Val Asp Asp Ile Gln Gln Ile Ala Ala 2610 2615 2620	7870
CCC CTG CCC CAG TCC ATG CCC CCC ACC ACC CAG CTC GTA TCC CCC TCG Ala Leu Ala Gln Cys Met Gly Pro Ser Arg Glu Leu Val Cys Arg Ser 2625 2630 2635	7918

TOC CTG AAG CAG ACG CTG CAC AAG CTG GAG GOC ATG ATG CTC ATC CTG 7966 Cys Leu Lys Gln Thr Leu His Lys Leu Glu Ala Met Met Leu Ile Leu 2650 ... 2645<sub>:::</sub> CAG GCA GAG ACC ACC GCG GCC ACC GTG ACG CCC ACC GCC ATC GCA GAC 8014 Gln Ala Glu Thr Thr Ala Gly Thr Val Thr Pro Thr Ala Ile Gly Asp 2670 2660 77.0 **2665** AGC ATC CTC AAC ATC ACA GGA GAC CTC ATC CAC CTG GOC AGC TOG GAC 8062 Ser Ile Leu Asn Ile Thr Gly Asp Leu Ile His Leu Ala Ser Ser Asp 2685 GTG CGG GCA CCA CAG CCC TCA GAG CTG GGA GCC GAG TCA CCA TCT CGG 8110 Val Arg Ala Pro Gln Pro Ser Glu Leu Gly Ala Glu Ser Pro Ser Arg 2690 2695 2700 3 11 4 ATG GTG GOG TOC CAG GOC TAC AAC CTG ACC TCT GOC CTC ATG OGC ATC 8158 Met Val Ala Ser Gln Ala Tyr Asn Leu Thr Ser Ala Leu Met Arg Ile 2705 **2710** (2) 2 15 1 (2) 27**15** 3 16 CTC ATG COC TOC COC GTG CTC. AAC GAG GAG COC CTG. ACG CTG GOC GGC 8206 Leu Met Arg Ser Arg Val Leu Asn Glu Glu Pro Leu Thr Leu Ala Gly 2730 8254 GAG GAG ATC GTG GOC CAG GGC AAG CGC TOG GAC COG CGG AGC CTG CTG Glu Glu Ile Val Ala Gln Gly Lys Arg Ser Asp Pro Arg Ser Leu Leu TICK THAT GOD GOD GOD COLA GOG COT GOD TOC CALL THE TOO LATE CODE GAG 8302 Cys Tyr Gly Gly Ala Pro Gly Pro Gly Cys His Phe Ser Ile Pro Glu .... **2760** €... 2755 GCT TTC AGC GGG GCC CTG GCC AAC CTC AGT GAC GTG GTG CAG CTC ATC 8350 Ala Phe Ser Gly Ala Leu Ala Asn Leu Ser Asp Val Val Gln Leu Ile TIT CIG GIG GAC TOO AAT COO TIT COO TIT GGC TAT ATC AGC AAC TAC 8398 Phe Leu Val Asp Ser Asn Pro Phe Pro Phe Gly Tyr Ile Ser Asn Tyr 2790 2795 ACC GTC TOO ACC AAG GTG GOO TOG ATG GOA TTC CAG ACA CAG GOO GOO 8446 Thr Val Ser Thr Lys Val Ala Ser Met Ala Phe Gln Thr Gln Ala Gly 2810 2800 - 2805 SOC CAG ATC COC ATC GAG COG CTG SOC TCA GAG COC GOC ATC ACC GTG 8494 Ala Gln Ile Pro Ile Glu Arg Leu Ala Ser Glu Arg Ala Ile Thr Val 2825 AAG GTG COC AAC AAC TOG GAC TOG GCT GCC COG GGC CAC COC AGC TOC 8542 Lys Val Pro Asn Asn Ser Asp Trp Ala Ala Arg Gly His Arg Ser Ser 2840 2835 2845 GOO AAC TOO GOO AAC TOO GIT GIG GIC CAG GOO CAG GOO TOO GIC GGT 8590 Ala Asn Ser Ala Asn Ser Val Val Val Gln Pro Gln Ala Ser Val Gly 2855 2860 GCT GTG GTC ACC CTG GAC AGC AGC AAC CCT GCG GCC GGG CTG CAT CTG  $\cdot$ 8638 Ala Val Val Thr Leu Asp Ser Ser Asn Pro Ala Ala Gly Leu His Leu 2865 2870 2875

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CAG Gln 288	Leu	AAC Asn	TAT Tyr	ACG Thr	CTG Leu 288	CTG Leu 5 '	GAC Asp	Gly	CAC His	TAC Tyr 289	Leu	TCT Ser	GAG Glu	GAA Glu	CCT Pro 2895	8686
GAG Glu	Pro	TAC	CTG Leu	GCA Ala 2900	Val	TAC Tyr	CTA Leu	CAC His	TCG Ser 290	Glu	OCC Pro	CCG Arg	Pro	AAT Asn 291	Glu	8734
CAC	AAC Asn	TGC Cys	TCG Ser 291	Ala	ACC Ser	AGG Arg	AGG Arg	ATC Ile 292	Arg	CCA Pro	GAG Glu	TCA Ser	CIC Leu 292	Gln	GT Gly	8782
			Arg			ACC Thr		Phe			Pro		Ser		GAC Asp -	8830
CCA Pro	GCG Ala 294	Gly	AGT Ser	TAC Tyr	CAT His	CTG Leu 2950	Asn	CTC Leu	TCC Ser	Ser	CAC His 295	Phe	CCC Arg	TCG Trp	TCG Ser	8878
	Leu					GC Gly					Leu					8926
					Val	TCG Trp		Thr		Gly			Pro		Glu	8974
		Ser		Arg		CCC Ala	Val	Cys		Thr		His		Thr		9022
			Ser			GIG Val		Pro			Val		Phe			<b>9070</b>
		$\mathbf{Pro}$			Asp.	GTA Val 3030	Asn		Ile	Val		Leu				9118
	Cys			Thr		ATG Met			Ala		Ile			Lys		9166
					Ser	CCG Arg				Ile			Cys		Gln	9214
				Lys		GAG Glu			Val					Gly		9262
			Thr			CAC His		Gly					Gly			9310
		Ser				CAC His 3110	Leu			Asp		Ala				9358

9934

10030

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9406 AAC AGC CTG GAC ATC TTC CGG ATC GCC ACC CGG GAC AGC CTG GGT AGC Asn Ser Leu Asp Ile Phe Arg Ile Ala Thr Pro His Ser Leu Gly Ser 3130 3135 3125 3120 GTG TOG AAG ATC OGA GTG TOG CAC GAC AAC AAA GOG CTC AGC OCT GOC .9454 Val Trp Lys Ile Arg Val Trp His Asp Asn Lys Gly Leu Ser Pro Ala 3145 3150 3140 TOG THE CTG CAG CAC GTC ATC GTC AGG GAC CTG CAG AGG GCA CGC AGC 9502 Trp Phe Leu Gln His Val Ile Val Arg Asp Leu Gln Thr Ala Arg Ser 3165 3155 3160 00 GOO THE THE CHE GHE AAT GAC TOG CHT TOG GHE GAG ACE GAG GOO AAC Ala Phe Phe Leu Val Asn Asp Trp Leu Ser Val Glu Thr Glu Ala Asn 3170 3180 3175 GGG GGC CTG GTG GAG AAG GAG GTG CTG GGC GGG AGC GAC GCA GGC CTT 9598 Gly Gly Leu Val Glu Lys Glu Val Leu Ala Ala Ser Asp Ala Ala Leu ... 3195 3190 3185 TTG CCC TTC CCG CCC CTG CTG CCT CAG CTG CAG CCT CCC TTC TTT ~9646 Leu Arg Phe Arg Arg Leu Leu Val Ala Glu Leu Gln Arg Gly Phe Phe 3205 3210 GAC AAG CAC ATC TOG CTC TOC ATA TOG GAC COG CCC CCT AGC CCT .... Asp Lys His Ile Trp Leu Ser Ile Trp Asp Arg Pro Pro Arg Ser Arg 3225 3230 3220 TTC ACT OSC ATC CAG AGG GOC AGG TGC TGC GTT CTC CTC ATC TGC CTC TAGG Phe Thr Arg Ile Gln Arg Ala Thr Cys Cys Val Leu Leu Ile Cys Leu 3240 TTC CTG GGC GCC AAC GCC GTG TGG TAC GGG GCT GTT GGC GAC TCT GCC 9790 Phe Leu Gly Ala Asn Ala Val Trp Tyr Gly Ala Val Gly Asp Ser Ala 3260 3250 3255 TAC AGC AGG GGG CAT GTG TOC AGG CTG AGC CTG AGC GTC GAC ACA Tyr Ser Thr Gly His Val Ser Arg Leu Ser Pro Leu Ser Val Asp Thr

3330 3335 3340

CAC GCT GAG GCC TITT GTT GGA CAG ATG AAG AGT GAC TITG TITT CTG GAT 10078

His Ala Glu Ala Phe Val Gly Gln Met Lys Ser Asp Leu Phe Leu Asp

3345 3350 3355

3270

3285

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GIC GCT GIT GGC CIG GIG TOC AGC GIG GIT GIC TAT COC GIC TAC CIG Val Ala Val Gly Leu Val Ser Ser Val Val Val Tyr Pro Val Tyr Leu

GOC ATC CIT TIT CTC TTC CGG ATG TOC CGG AGC AAG GTG GCT GGG AGC

Ala Ile Leu Phe Leu Phe Arg Met Ser Arg Ser Lys Val Ala Gly Ser

COC AGC COC ACA CCT GOC GOG CAG CAG GTG CTG GAC ATC GAC AGC TGC Pro Ser Pro Thr Pro Ala Gly Gln Gln Val Leu Asp Ile Asp Ser Cys

CTG GAC TOG TOC GTG CTG GAC AGE TOC THE CTC AGG THE TCA GGC CTC

Leu Asp Ser Ser Val Leu Asp Ser Ser Phe Leu Thr Phe Ser Gly Leu

3290

3305

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GAT Ast 336	TCT Ser 50	AAG Lys	AGT Ser	CTG Leu	GIG Val 336	Cys	TGG Trp	Pro	TCC Ser	GC Gly 3370	Glu	GGA Gly	ACG Thr	CIC	AGT Ser 3375	· ·	10126 ·
TC	Pro	GAC Asp	CIG Leu	CIC Leu 338	Ser	GAC Asp	CCG Pro	TCC Ser	ATT Ile 338	Val	GIY	AGC Ser	AAT Asn	CIG Leu 339	Arg		10174
CAC Glr	CTG Leu	GCA Ala	03G Arg 339	Gly	CAG Gln	GCG Ala	CCC Gly	CAT His 340	Gly	CTG Leu	GCC Gly	CCA Pro	GAG Glu 340	Glu	GAC Asp		10222
GCL	TTC Phe	TCC Ser 3410	Leu	CCC Ala	AGC Ser	ccc Pro	TAC Tyr 341	Şer	CCT Pro	cc Ala	AAA Lys	TCC Ser 3420	Phe	TCA Ser	GCA Ala		10270
Ser	GAT Asp 342	Glu	GAC Asp	CTG Leu	ATC Ile	CAG Gln 3430	Gln	GTC Val	CIT Leu	Ala	GAG Glu 343	Gly	GIC Val	AGC Ser	AGC Ser		10318
	OCC Ala				Asp		His				Asp						L0366
CIG	TCC Ser	AGC Ser	ACT Thr	CCT Pro 3460	Gly	GAG Glu	AAG Lys	Thr	GAG Glu 3465	Thr	CIG Leu	ccc Ala	Leu	CAG Gln 3470	Arg	1	LO414
CIG	Gly	GAG Glu	CIG Leu 3475	Gly	CCA Pro	cc Pro	AGC Ser	CCA Pro 3480	Gly	CIG Leu	AAC Asn	Trp	GAA Glu 3485	Gln	OCC Pro		10462
CAG Gln	GCA Ala	Ala 3490	Arg	CTG Leu	TCC Ser	AGG Arg	ACA Thr 3495	Gly	CIG Leu	GIG Val	Glu	GGT Gly 3500	Leu	CCG Arg	aag Lys	1	10510
	CIG Leu 350	Leu					Ala					Gly				]	10558
CIC Leu 352	CTG Leu 0	GTG Val	CCT Ala	GIG Val	GCT Ala 3525	Val	CCT Ala	GIC Val	TCA Ser	000 Gly 3530	Trp	GTG Val	OCT Gly	ccc Ala	AGC Ser 3535	. 1	10606
TTC	e Pro	OCG Pro	Gly	GTG Val 3540	Ser	GTT Val	ccc Ala	TCG Trp	CTC Leu 3545	Leu	TCC Ser	AGC Ser	Ser	CCC Ala 3550	Ser	. 1	10654
TTC	CTG Leu	CCC Ala	TCA Ser 355	Phe	CTC Leu	Gĵy	TCG Trp	GAG Glu 3560	Pro	CIG Leu	AAG Lys	GTC Val	TTG Leu 3565	Leu	GAA Glu		10702
GCC Ala	CTG Leu	TAC Tyr 3570	Phe.	TCA Ser	CIG Leu	GTG Val	CCC Ala 3575	Lys	CGG Arg	CTG Leu	His	003 Pro 3580	Asp.	GAA Glu	GAT Asp	1	10750
Asp	ACC Thr 358	Leu	GTA Val	GAG Glu	ACC Ser	000 Pro 3590	Ala	olg Vaj	ACG Thr	CCT Pro	GIG Val 3595	Ser	CCA Ala	CCT Arg	GIG Val	1	10798

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,	Pro 360	Arg	GTA Val	Arg	Pro	Pro 360	His	G1y	TITI Phe	GCA Ala	CIC Leu 361	Phe	Leu	GCC Ala	AAG Lys	GAA Glu 3615		10846
,	GAA Glu	Ala	Arg	AAG Lýs	GIC Val 362	Lys	AGG Arg	CTA Leu	His	GGC Gly 362	Met	CIG Leu	Arg	ACC Ser	CIC Leu 363		-	10894
	OIG Val	TAC	ATG	CTT Leu 363	Phe	CIG	CTG Leu	GIG Val	ACC Thr 364	Leu	CIG	QCC Ala	ACC	TAT Tyr 364	Gly	GAT Asp		10942
				CAT His O					Arg			Ser		Ile			•	10990
			His	AGC Ser			Phe		Ala		Thr		Ser			CTC <sub>1</sub> : Leu		11038
		Pro		ATG Met	Ala		Val					Val					•	11086
				GAG Glu		Gly		Pro	Årg		Arg					Gln		11134
				TAC Tyr 3715	Pro					Pro			His		Cys			11182
	Ala	GCA Ala	GGA Gly 3730	GIY	TTC Phe	AGC Ser	ACC Thr	AGC Ser 3735	Asp	TAC Tyr	GAC Asp	GIT Val	GC Gly 3740	Trp	GAG Glu	AGT Ser		11230
			Asn	Gly				Trp					Pro					11278
(		Ala		TCC Ser	Trp		Ser		Ala	Val		Asp			Gly		••	11326
1	/al	Gln	Glu	CTG Leu	Gly 3780	Leu	Ser	Leu :	Glu	Glu 3785	Ser	Arg	<b>Asp</b>	Arg	Leu 3790	Arg		11374
I	Phe	Leu	Gln	CIG Leu 3795	His	Asn	Trp	Leu	Asp 3800	Asn	Arg	Ser	Arg	Ala 3805	Val	Phe		11422
I	eu	Glu	Leu 3810		Arg	Tyr	Ser	Pro 3815	Ala	Val	Gly	Leu	His 3820	Ala ) .	Ala	Val		11470
	hr		Arg	CIC Leu		Phe		Ala			Arg .		Leu				•	11518

								25	) <i>   </i>	•							
AGC Ser 3840	Val	CCC Arg	Pro	TTT Phe	CCG Ala 3845	Leu	OCC Arg	CCC Arg	CIÇ Leu	AGC Ser 3850	Ala	GIŸ	CTC Leu	TCG Ser	CIG Leu 3855	115	66
OCT Pro	CIG Leu	CTC Leu	ACC Thr	TCG Ser 3860	Val	TGC Cys	CTG Leu	Leu	CTG Leu 3865	Phe	ECC Ala	GTG Val	CAC His	TTC Phe 387(	Ala	116	14
GTG Val	CC Ala	GAG Glu	CC Ala 387	Arg	ACT Thr	<u>1</u> rp 103	CAC His	AGG Arg 3880	Glų	Gly	CCC Arg	TGG Trp	CCC Arg 3885	Val	CIG Leu	116	62
CGG Arg	CIC Leu	GGA Gly 3890	Ala	TGG Trp	CCC Ala	Arg	TCG Trp 3895	Leu	ÇIĞ Leu	Val Val	CCG Ala	CTG Leu 3900	Thr	CCG Ala	Ala	117	10
ACG Thr	GCA Ala 3905	Leu	GTA Val	ŒC Arg	CTC Leu	GCC Ala 3910	Ģln	CIG Leu	GCT Gly	∝ Ala	GCT Ala 391	Aśp	CCC Arg	CAG Gln	TGG Trp	117	58
ACC Thr 3920	Arg	TTC Phe	GIG Val	Arg Arg	GC Gly 3925	Arg	ccc Pro	CCC Arg	CGC Arg	TTC Phe 3930	Thr	AGC Ser	TTC Phe	GAC Asp	CAG Gln 3935	118	06
GIG Val	GCG Ala	CAC His	Val	AGC Ser 394	Ser	CCA Ala	CCC Ala	CGT Arg	GGC Gly 394	CIG Leu 5	CCC Ala	Ala	TCG Ser	CIG Leu 395	Leu	118	54
TTC Phe	CTG Leu	CTT Leu	TTG Leu 395	Val	AAG Lys	GCT Ala	œc Ala	396 Gln CAG	His	GTA Val	Arg	TTC Phe	GIG Val 396	Arg	CAG Gln	119	02 .
TCG Trp	TCC Ser	GIC Val 397	Phe	Gly	AAG Lys	ACA Thr	TTA Leu 397	Cys	CCA Arg	OCT Ala	CIG Leu	CCA Pro 398	Glu	CIC	CIG Leu	119	50
Gly	GTC Val 398	Thr	TTG	Gly	CIG Leu	GIG Val 399	Val	CIC	Gly	GTA Val	Ala 399	Tyr	CCC Ala	CAG Gln	CIG L <u>ệ</u> u	, 119	98
Ala 400	Ile	CTG Leu	CTC	GIG Val	TÇT Ser 400	Ser	TGT Cys	GTG Val	GAC Asp	Ser 401	Leu	TCG	AGC Ser	GIG Val	CCC Ala 4015	., 120	<b>14</b> 6
CAG Gln	GCC Ala	CIG	TTG Leu	Val Val 402	Leu	TGC Cys	CCT Pro	Gly	ACT Thr 402	Gly	-CTC	-TCT Ser	ACC Thr	CIG Leu 403	Cys Cys	120	94
CCT Pro	GCC Ala	GAG Glu	TCC Ser 403	Trp	CAC His	CIG	TCA Ser	Pro 404	Leu	CIG Leu	TGI Cys	GIG Val	GGG Gly 404	Leu	TCG	121	142
CCA Ala	CIG	Arg 405	Leu	TGG	Gly	Ala	CTA Leu 405	Arg	CIC Leu	GCG Gly	GCI Ala	Val 406	Ile	CIC Leu	CCC Arg	12:	190
TCC	Arg 406	Tyr	CAC His	C GCC	Leu	Arg 407	Gly	GAC Glu	CIC Leu	TAC 1 Tyr	Arg 407	Pro	Ala	Tri	GAG Glu	12	238

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Pro 4080	Gln	GAC Asp	TAC Tyr	GAG Glù	ATG Met 4085	Val	GAG Glu	TTG Leu	TTC Phe	CIG Lëu 4090	Arg	AGG Arg	CIG Leu	OCC Arg	CTC Leu 4095	12286
TCG Trp	ATG Met	G Gly	CIC	ACC Ser 4100	Lys	GTC Val	aag Lys	GAG Glu	TIC Phe 410	Arg	CAC His	aaa Lys	GTC Val	Arg 4110	Phe 👵	12334
GAA Glu	Gly Gly	ATG Met	GAG Glu 4115	Pro	CTG Leu	OCC Pro	TCT Ser	Arg 4120	Ser	TCC Ser	AGG Arg	GCC Gly	TCC Ser 4125	Lys	GTA Val	12382
TCC Ser	œ Pro	GAT Asp 4130	Val	CC Pro	CCA Pro	CC Pro	AGC Ser 413	Ala	Gly	TCC Ser	GAT Asp	CCC Ala 414(	Ser	CAC His	CCC (	, <b>12430</b> · ·
Ser	ACC Thr 4145	Ser	TCC Ser	AGC Ser	CAG Gln	CTG Leu 4150	Asp	GG Gly	CTG Leu	ÄGC Ser	GIG Val 4155	Ser	CIG Leu	GC Gly	CCG Arg	12478
CIG Leu 4160	Gly	ACA Thr	AGG Arg	TGT Cys	GAG Glu 4165	Pro	GĀĞ Glu	OCC Pro	TCC Ser	OSC Arg 4170	Leu	CAA Gln	CCC Ala	Val Val	TTC Phe 4175	12526
GAG Glu	CCC Ala	CIG Leu	CIC Leu	ACC Thr 4180	Gln	TTT Phe	GAC Asp	OGA Arg	CIC Leu 418	Àsn	CAG Gln	ÖCC Ala	Thr	GAG Glu 4190	Asp ·	12574
GIC Val	TAC Tyr	CAG Gln	CTG Leu 419	Glu	CAG Gln	CAG Gln	ĆIĠ Leu	CAC His 4200	Ser	CIG Leu	CAA Gln	Gly	OSC Arg 4205	Ārģ	AGC Ser	12622
AGC Ser	CCG Arg	GOG Ala 4210	Pro	ССС Ala	GCA Gly	TCT Ser	TCC Ser 421	Arg	GC Gly	CCA Pro	TCC Ser	OG Pro 4220	Gly	CIG	CCG . Arg .	<b>12670</b>
		Leu					Ala					Gly			CTG Leu	12718
600 Ala 4240	Thr	GC Gly	Pro	AGC Ser	AGG Arg 424	Thr	CCT Pro	TCG Ser	GC	CAA Gln 4250	Glu	CÂA Gl n	CCT Cly	CCA Pro	Pro 4255	12766
CAĞ Gln	CAG Gln	CAC His	Leu	GIC Val 4260	Leu	CIT	CCT Pro	Gly	GGG Gly 426	Gly	GGG Gly	Pro	Trp	AGT Ser 427	OGG Arg D	12814
AGT Ser	GGA Gly	CAC His	Arg 427	Ser	(Val	TTA Leu	CTT Leu	TCT Ser 428	Ala	GCT Ala	GIC Val	AAG Lys	CCC Ala 428	Glu	Gly	12862
			Trp					Ser					Gln		CAT His	12910
CTG Leu	TCT Ser 430	Val	TGT Cys	Gly	ÇIT Leu	CAG Gln 431	His	TTT Phe	AAA Lys	GAG Glu	GCT Ala 431	Val	TCG	CCA Pro	ACC Thr	12958

AGG ACC CAG GGT CCC CTC CCC AGC TCC CTT CGG AAG GAC ACA GCA GTA Arg Thr Gln Gly Pro Leu Pro Ser Ser Leu Gly Lys Asp Thr Ala Val 4320 4335	13006
TTG GAC GGT TTC TAGCCTCTGA GATGCTÄATT TATTTCCCCG AGTCCTCAGG	13058
Leu Asp Gly Phe	. 1
TACAGOGGC TGTGCCCCGC CCCACCCCCT GGGCAGATGT CCCCCACTGC TAAGGCTGCT	13118
GOCTICAGGG AGGGITAGGC TGCACCGCCG CCACCCTGGC CCTAAGTTAT TACCTCTCCA	13178
GTTCCTACCE TACTCCCTCC ACCEPTCAC TGTGTGTCTC GTGTCAGTAA TTTATATGGT	13238
GTTAAAATGT GTATATTTTT GTATGTCACT ATTTTCACTA GGGCTGAGGG GCCTGGGGCCC	13298
AGAGCIGGCC TCCCCCAACA CCTGCTGCGC TTGGTAGGTG TGGTGGCGTT ATGGCAGCCC	13358
GOCTOCTOCT TOCATOCCAG CITOCCCTTG GOCCOCACAG CIGTCTGCCA	13418
GOCACICICA TCACCCAGA GOCCITGICA TOCICOCITG COCCAGOCCA GOTAGCAAGA	13478
GAGCAGCCC CAGCCCTCCT GCCATCAGGT CTGCGCAAGT AGCAGGACTA GCCATGTCAG	13538
AGGACCCCAG GETGETTAGA GGAAAAGACT CCTGCTGGGG GCTGGCTCCC AGGGTGGAGG	13598
ANGERGACIG TERETETETE TERETECCOC COCCACCOCC CAGINETICCIC TATECCOCCAG	13658
GCAGCCICAA GGCCCICGGA GCIGGCIGIG CCIGCTICIG TGTACCACTT CIGIGGGCAT	13718
GGCCGCTTCT AGAGCCTCGA CACCCCCCA ACCCCGCAC CAAGCAGACA AAGTCAATAA	13778
AAGAGCTGTC TGACTGCAAA AAAAAAAAA	13807
(ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ.ID NO: 2:	r
Gly Ala Ala Cys Arg Val Asn Cys Ser Gly Arg Gly Leu Arg Thr Leu 1 10 15	
Gly Pro Ala Leu Arg Ile Pro Ala Asp Ala Thr Ala Leu Asp Val Ser 20 25 30	. 3
His Asn Leu Leu Arg Ala Leu Asp Val Gly Leu Leu Ala Asn Leu Ser 35 40 45	
Ala Leu Ala Glu Leu Asp Ile Ser Asn Asn Lys Ile Ser Thr Leu Glu 50 55 60	
Glu Gly Ile Phe Ala Asn Leu Phe Asn Leu Ser Glu Ile Asn Leu Ser 65 70 75 80	•
Gly Asn Pro Phe Glu Cys Asp Cys Gly Leu Ala Trp Leu Pro Arg Trp 85 90 95	
Ala Glu Glu Gln Gln Val Arg Val Val Gln Pro Glu Ala Ala Thr Cys 100 105 110	•

	130		. •	. <i>i</i>	•	133		٤. '	Val		7.40	٠.,	٠.	1	••			•		
145				٠.	150	•	-	:	• •	Ĭ	( ,,		•-	·					., - ;	٠
Leu	Leu	Gln	Pro	Glu 165	Ala	Cys	Ser	Ala	Phe 170	Cys	Phe	Ser	Thr	Gly 175	Gl	n			•	
Gly	Leu	Ala	Ala 180	Leu	Ser	Glu	Gln	Gly 185	Trp	Cyś	<u>Leu</u>	Cys	Gly 190	Ala	Al	: :	2		T,	
Gln	Pro	Ser 195	Şer	Ala	Ser	Phe	Ala 200	Cys	Leu	Ser	Leu	Cys 205	Ser	GJA	Pr	<b>.</b>		· C	٠.	
	: Pro 210	Pro	Pro	Aĺa	Pro	Thr 21,5	Cys	Arg	Gly	Pro	Thr 220	Leu	Leu	Gln	Hi	s .	· -	 		
Val: 225	Phe	Pro	Ala	Ser	Pro 230	Gly	Ala	Thr	Leu.	Val 235	Gly	Pro	His	Gly	24	Q		•	5%	i.
Leu	Ala	Ser	Gly	Gln 245	Leu	Ala	Ala	Phe	His 250	Ile	Ala	Ala	Pre	255	ı Pr	Ö	· ·	<b>`</b>	ارون. ار	-in.
Val	Thr	Ala	. Thr 260	Arg	Trp	Asp	Phe	Gly 265	Asp	Gly	Ser	Ala	Glu 270	val	As	, sb			ÇAÇ v	
Ala	Ala	Gly 275		Ala	a Ala	Ser	His 280	Arg	Tyr	Val	Teu	Pro 285	Gly	Arc	τ <u>η</u> 	γ <b>τ</b> .	•		*	Fernanda 1 Jun
His	Val 290	Thr	Ala	val	Leu	1 Ala 295	Leu	Gly	Ala	Gly	Ser 300	Ala	Let	ı Lei	G	ly	, 	ė	;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;;	
Thi: 305	_	Val	l Gli	n Val	1 Glu 310		Ala	Pro	Ala	Ala 315	Leu	Gli	Lea	υ Va	1 .C 3	ys 20		, · ·		•
Pro	Ser	: Sei	r Va	1 Gli 32	n Sei 5	c Ast	Glu	. Ser	1 éu 330	Asp	Lev	ı Sei	c Ile	e Gl: 33	n A 5	sn.			, , , , ,	
Arg	Gl3	, Gl	y Se:	r Gl	ÿ Lei	u Glu	ı Ala	Ala 345	Туг	Ser	Ile	e Va	1 Al 35	a Le O	u G	ly	•			
Glu	ı Glı	2 Pro 35		a Ar	g Ala	 a Va	1 His	s Pro	Lev	. Суз	Pro	Se: 36	r As 5	p Th	r G	lu				
Ile	Ph:	e Pro	o G1	y As	n Gl	у Ні: 37	s Cys 5	s Tyr	r Axg	Leu	va. 38	i Va O	1 <sup>'</sup> G1	u Ly	s A	la		• .		
Ala 38		p Le	u Gl	n Al	.a G1 39	n Gl	u Gl	n Cy	s Glr	Ala 395	ā Trī	p <sub>.</sub> Al	a Ġl	y A1	a A	100		• •	€'	٠
Le	u Al	a Me	t Va	1 As 40	sp Se )5	r Pr	o Al	a Va	1 Glr 410	n Arī	g Ph	e Le	u. Va	11 Se 41	er A L5	Arg		ř		
Va	l Th	r Ar	g Se 42		eu As	sp Va	l Tr	p II 42	e Gl 5	y Ph	e Se	r Tr	ir Va 43	30 श G	ln (	31ý		٠		
Va	ı Gl	u Va 43		ly Pi	ro Al	la Pr	o G1 44	n Gl 10	y Gl	u Al	a Ph	e Se 44	er Le 15	eu G	ļu '	Ser	•			
Cy	rs GJ 45		en`T	က်က	eu Pa	ro G 45	ly G1 55	u Pr	ю нт	s PI	ο Al 46	ia 11 60	nr A	la G	lu Ì	His	; ·			

Cys 465	Val	Arg	Leu	Gly	Pro 470		Gly	Trp	Cys	Asn 475	Thr	Asp	Leu	Cys	Ser 480
Ala	Pro	His	Ser	Tyr 485	Val	_	Glu >- \		Gln 490	Pro	Gly	Gly :	Pro	Val 495	
Asp	Ala	Glu	Asn 500	Leu	Leu	Val		Ala 505	Pro	Ser	Gly	Asp :	Leu 510	Gln	Gly
Pro	Leu	Thr 515	Pro	.Leu	Ala	Gln	Gln 520	Asp	Gly	Leu	Ser	Ala 525	Pro	His	Glu
Pro	Val 530	Glu	Val	Met	<u>V</u> al	Phe 535	Pro	⁄Gly	Leu	Arg	Leu 540	Ser	Arg	Glu	Ala .
Phe 545	Leu	Thr	Thr	Ala	Glu 550	Phe	Gly	Thr	Gln	Glu 555	Leu	Arg	Arg	Pro	Ala 560
Gln	Leu	Arg	Leu	Gln 565	Val	Ţyr	Arg		Leu 570	Ser	Thr	Ala	Gly.	Thr 575	Pro ·
Glu	Asn	Gly	Ser 580	Glu	Pro	Glu	Ser	Arg 585	Ser	Pro	Asp	Asn	Arg 590	Thr	Gln
Leu	Ala	Pro 595	Ala	Cys	Met	Pro	Gly 600	Gly	Arg	Trp	Cys	Pro 605	Gly	Ala	Asn
Ile	Cys 610		Pro	Leu	Asp	Ala 615	Ser	Cys	His	Pro	Gln 620		Cys	Ala.	Asn∷.
Gly 625	Cys	Thr	Ser	Gly	Pro 630	Gly	Leu	Pro	Gly	Ala 635	Pro	Tyr	Ala	Leu-	Trp :: 640
Arg	Glu	Phe	Leu	Phe 645	Ser	Val	Ala	Ala	G1y 650	Pro	Pro	Ala	Gln	Tyr 655	Ser '
Val	Thr	Leu	His 660	Gly	Gln 	Āsp	Val	Leu 665	Met	Leu	Pro	Gly	Asp 670	Leu	Val
Gly	Leu	Gln 675	His	Asp	Ala	Gly	Pro 680	Gly	Ala	Leu	Leu ···	His 685	Cys	Ser	Pro.
Ala	Pro 690	Gly	His	Pro	Gly	Pro 695		Ala	Pro	Tyr	Leu 700	Ser	Ala	Asn	Ala
Ser 705	Ser	Trp	Leu 	Pro.	His 710	Leu	Pro	Ala	Gln	Leu 715	Glu	Gly	Thr	Trp	Ala 720
Cys	Pro	Ala		Ala 725	Leu	Arg	Leu	-Leu	Ala 730	Ala	Thr	Glu	Gln	Leu 735	Thr -
Val	Leu	Leu	Gly 740	Leu	Arg	Pro	Asn	Pro 745	Gly	Leu	Arg	Met	Pro 750	Gly.	Arg
Tyr	Glu	Val 755	Arg	Ala	Glu	Val	Gly 760	Asn	Gly	Val	Ser	A <del>rg</del> 765	His	Asn	Leu
Ser	Cys 770		Phe	Asp	Val	.Val 775	Ser	:Pro	Val	Ala	Gly 780	Leu	Arg	Val	Ile
Tyr 785	Pro	Ala	Pro :	-Arg	Asp 790	Gly	_	·Leu ·	Tyr	Val 795	Pro	Thr	Asn	Gly	Ser 800

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Ala	Leú	Val	Leu -	Gln 805	Val	Asp :	Ser	Gly	Ala 810	Asn	Ala	Thr	Ala	Thr 815	Ala		•
Arg	Trp	Pro	Gly 820	Gly	Ser	Val	Ser	Ala 825	Arg	Phe	Glu	Asn	Val 830	Cys	Pro		
Ala	Leu	Val 835	Ala	Thr	Phe	Val	Pro 840	Gly	Cys	Pro	Trp	Glu 845	Thr	ÁŚN	Asp		
Ihr	Leu 850	Phe	Ser	Val	Val	Ala 855	Leu	Pro	Trp	Leu	Ser 860	'Glu	Gly	.Glu	His		ř
Val 865	Val	Ásp	Val	Val	Val 870	Glu	Asn '	Ser	Ala	Ser 875	Arg	Ala	Asn	Leu	Ser 880	. :	
Leu	Arg	Val	Thr	Ala 885	Glu '	Glu	Pro	Ile	Cys 890	Gly	ľéu	Ar <del>g</del>	Ala	Thr 895	Pro		ers US
Ser	Pro		Ala: 900	Arg	Val	Leu	Gln	Gly 905	Val:	Leu '	Val	Aig	Tyr 910	Seir	Pro	•	·· ·;·
Val	Val	Glu 915	Ala	ΟΊУ	Ser	Asp	Met 920	Val	Phe	Arg	Trp	Thr 925	lle	Ąsn	Asp	`	á. ú.
Lys	Gln 930	Ser:	Leu	.Thr	Phe	Gln 935	Asn	Val	Vàl	Phe	Asn 940	Val	Ile	Tyr	Gln -	. *	٠.
Ser 945	Ala	Ala:	Val <sup>-</sup>	Phe	Lys 950	Leu	Sér	Léù	Thr	Ala 955	Ser	'Asn	His	Val	Ser . 960 :	: 1	***
Asn	Val	Thr	Val	Asn 965	Tyr	Asn	Val	Thr	Val 970	Glu '	Arg	Met	Asn	Arg 975	Met .	, <u>;</u> (1,	1977 1872 1872
Gln.	Gly	Leu`	Gln 980	Val	Ser	Thr	Val	Pro 985	Ala	Val	Leu		990	'Asn	Ala	•	
I'hr	Leu'	Vál 995	Leu	The	Gly	Gly	Val 1000		Val	Asp	Ser	Ala 1005		Glu	Val		
Ala	Phe 1010		Tip	Asn	Phe	Gly 1015		Gly	Glu		Ala 1020		His	Gln	Phe -	t p.*	
Gln 1025		Pro	Tyr	Asn	Glu 1030		Phe	Pro	.Val	Pro 1035		Pro	Ser	Val	Ala 1040		
Gln	Val <sub>,</sub>	Léu	Val	Glu 1045		Aśn	Val	Met	His 1050		Tyr	Ala L	Ala	Pro 1055	Gly:	• ;-	
Glu	Tyr		Leu 1060		.Val	Leu	Ala	Ser 1065		Ala	Phe	Glu	Asn 107(		Thr -	٠.	V <sub>a</sub> v
Gln	Gln	Val 1075		Val	Ser	Val	Arg 1080	_	Ser	Leu	Pro	Ser 108	_	Ala	Val	٠.	,
Gly	Val. 1090		.Asp		Val	Leu 109		Ala	Gly	Arg	Pro 1100		Thr	:Phe	Tyr	• •	
Pro 1105		Pro	Leu	Pro	Ser 1110	~	.Gly	Gly	Val	Leu 111!	_	Thr	Trp	Asp	Phe 1120	. •	
Gly	Asp	Gly	Ser	Pro 112		Leu	Thr	:Gln	Ser 113		Pro	'Ala	Ala	Asin 113	:His'	•	:.

### SUBSTITUTE SHEET (RULE 26)

								.54	+/ / /	1					
Thr	Tyr	Ala	Ser 114		Gly	Thr	Tyr	His 114		Arg	Leu	Glu	Val 115	Asn O	Asn
Thr	Val	Ser 115		Ala	Ala	Ala	Gln 116		Asp	Val	Arg	Val 116		Glu	Glu
Leu	Arg 1170		Leu	Ser	Val	Asp 117		Ser	Leu	Ala	Val 118	<u>-</u>	Gln	Gly	Ala
Pro 118		Val	Val	Ser	Ala 1190		Val	Gln	Thr	Gly 119	Asp 5	Asn	Ile	Thr	Trp 120
Thr	Phe	Asp	Met	Gly 120	Asp 5	Gly	. Libić	Val	Leu 121		Gly	Pro	Glu	Ala 121	
Val	Glu	His	Val 1220		Leu	Arg	Ala	Gln 122		Cys	Thr	Val	Thr 123	_	Gly
Ala	Ala	Ser 123		Ala	Gly	His	Leu 1240	_	Arg	Ser	Leu	His 124	_	Leu	Val
Phe	Val 1250		Glu	Val	Leu	Arg 125		Glu	Pro	Ala	Ala 1260		Ile	Pro	Thr
Gln 1265	Pro	Asp	Ala	Arg	Leu 1270		Ala	Tyr	Val	Thr 1275		Asn	Pro	Ala	His 1280
Tyr	Leu	Phe :	Asp	Trp 1285	Thr	Phe	Gly	Asp	Gly 1290	Ser )	Ser	Asn	Thr	Thr 1295	
Arg	Gly	Cys	Pró 1300		Val	Thr	His	Asn 1305		Thr	Arg	Ser	Gly 1310	_	Phe
Pro	Leu	Ala 1315		Val	Leu	Ser 	Ser 1320		Val	Asn	Arg	Ala 132		Tyr	Phe
Thr	Ser 1330	Ile )	Cys	Val	Glu (	Pro 1335		Val	Gly	Asn	Val 1340		Leu	Gln	Pro
Glu 1345	Arg	Gln	Phe		Gln 1350		Gly	Asp	Glu ;	Ala 1355		Leu	Val	Ala ;	Cys 1360
Ala	Trp	Pro	Pro	Phe 1365	Pro	Tyr	Arg	Tyr	Thr 1370	_	Asp	Phe	Gly	Thr 1375	-
Glu	Ala	Ala ,.	Pro 1380		Arg	Ala	Arg	Gly 1385		Glu	Val	Thr	Phe 1390		Tyr
Arg	Asp	Pro 1395		Ser	Tyr	Leu	Val 1400		Val	Thr	Ala	Ser 140		Asn :	Ile
Ser	Ala 1410		Asn	Asp	Ser	Ala 1415		Val	Glu 	Val	Gln 1420		Pro	Val	Leu
Val 1425		Ser 	Ile	Lys	Val 1430		Ğly	Ser 	Leu ,	Gly 1435	-	Glu	Leu	Gln	Gln 144(
Pro	Tyr	Leu	Phe	Ser	Ala	Val	Gly	Arg	Gly	Arg	Pro	Ala	Ser	Tyr	Leu

Trp	Asp	Leu	Gly 1460		Gly	Gly	Trp	Leu 1465		Gly	Pro	Glu	Val 1470		His		
Ala	Tyr	Asn 1475		Thr	Gly	Asp	Phe 1480		Val	Arg	Val	Ala 1485	Gly	Trp	Asn	•	
Glu	Val 1490		Arg.	Ser	Glu	Ala 1495		Leū	Asn	Vaĺ	Thr 1500		Lys 	Arg	Arg	•	
Val 1505	_	Gly	Leu	Val	Val 1510		Ala	Ser	Arg	Thr 1515	Val	Val	Pro	Leu	Asn 1520	2	•
Gly	Ser	Val	Ser ,	Phe 1525		Thr	Ser	Leu	Glu 1530		Gly	Ser	Asp	Val 1535		•- * ,	
		¥	_					_•									'
Tyr	Ser	Trp	Val 1540		Cys	Asp	Arg	Cys 1545	Thr	Pro	Ile		G1y 1550		Pro	27	٠.
Thr	Ile	Ser 1555		Thr	Phe	Arg	Ser 1560	•	Gly ;	Thr	Phe	Asn 1565		Ile	Val	gra mangan	:
Thr	Ala 1570		Asn	Glu	Val	Gly 1575	-	Ala :	Gln	Asp	Ser 1580		Phe	Val	Tyr		
Val 1585		Gln	Leu		Glu 1590					Val 1595	Gly	Gly	Gly		1600 1600	147.7 14. 14. 14.	•
Phe	Pro	Thr	Asn	His 1605		Val		Leu co			Val	Val	Arg	Asp 1615		e Vojak i je	•
Thr	Asn	Val	Ser 1620		Ser	Trp		Ala 1625		Arg	Asp	Arg	Gly 1630		Ala	٠.	
Leu	Ala	Gly 1635		Gly	Lys	Gly	Phe 1640	•	Leu		Val	Leu 1645		Ala	Gly		
Thr	Tyr 1650		Val	Gln		Arg 1655		Thr	Asn	Met 	Leu 1660	_	Ser :	Ala	Trp	• • •	
Ala 1665		Cys	Thr	Met	Asp 1670		Val	Glu	Pro	Val 1675	Gly	Trp	Leu	Met	Val 1680	7	•
Thr	Ala	Ser	Pro	Asn 1685	-	Ala	Ala		Asn 1690		Ser		Thr	Leu 169	-	•.	
Ala	Glu	Leu	Ala 1700	Gly )	Gly	Ser	Gly	Val 1705		Tyr	Thr	Trp	Ser 1710		Glu		
Glu	Gly	Leu 1715		Trp	Glu		Ser 1720	_	Pro	Phe	Thr	Thr 1725		Ser			
Pro	Thr 1730		Gly	Leu		Leu 173					Ala 1740		Asn	Pro	Leu 	•	•
Gly 1745	_	Ala	Asn		Thr 1750		Glu		Asp		Gln 5	Val	Pro	Val	Ser 1760	•	
Gly	Leu	Ser	Ile	Arg 176		Ser	Glu	Pro	Gly 1770		Ser	Phe	Val	Ala 177		•	

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- Gly Ser Ser Val Pro Phe Trp Gly Gln Leu Ala Thr Gly Thr Asn Val 1780 1785 1790
- Ser Trp Cys Trp Ala Val Pro Gly Gly Ser Ser Lys Arg Gly Pro His 1795 1800 1805
- Val Thr Met Val Phe Pro Asp Ala Gly Thr Phe Ser Ile Arg Leu Asn 1810 1815 1820
- Ala Ser Asn Ala Val Ser Trp Val Ser Ala Thr Tyr Asn Leu Thr Ala 1825 1830 1835 1840
- Glu Glu Pro Ile Val Gly Leu Val Leu Trp Ala Ser Ser Lys Val Val 1845 1850 1855
- Ala Pro Gly Gln Leu Val His Phe Gln Ile Leu Leu Ala Ala Gly Ser 1860 1865 1870
- Ala Val Thr Phe Arg Leu Gln Val Gly Gly Ala Asn Pro Glu Val Leu 1875 1880 1885
- Pro Gly Pro Arg Phe Ser His Ser Phe Pro Arg Val Gly Asp His Val 1890 1895 1900
- Val Ser Val Arg Gly Lys Asn His Val Ser Trp Ala Gln Ala Gln Val 1905 1910 1915 1920
- Arg Ile Val Val Leu Glu Ala Val Ser Gly Leu Gln Met Pro Asn Cys 1925 1930 1935
- Cys Glu Pro Gly Ile Ala Thr Gly Thr Glu Arg Asn Phe Thr Ala Arg. 1940 1945 Ag 1950
- Val Gln Arg Gly Ser Arg Val Ala Tyr; Ala Trp Tyr Phe Ser Leu Gln 1955 1960 1965
- Lys Val Gln Gly Asp Ser Leu Val Ile Leu Ser Gly Arg Asp Val Thr. 1970 1975 1980
- Tyr Thr Pro Val Ala Ala Gly Leu Leu Glu Ile Gln Val Arg Ala Phe 1985 1990 1995 2000
- Asn Ala Leu Gly Ser Glu Asn Arg Thr Leu Val Leu Glu Val Gln Asp 2005 2010 2015
- Ala Val Gln Tyr Val Ala Leu Gln Ser Gly Pro Cys Phe Thr Asn Arg 2020 2025 2030
- Ser Ala Gln Phe Glu Ala Ala Thr Ser Pro Ser Pro Arg Arg Val Ala 2035 2040 2045
- Tyr His Trp Asp Phe Gly Asp Gly Ser Pro Gly Gln Asp Thr Asp Glu 2050 2055 2060
- Pro Arg Ala Glu His Ser Tyr Leu Arg Pro Gly Asp Tyr Arg Val Gln 2065 2070 2075 2080
- Val Asn Ala Ser Asn Leu Val Ser Phe Phe Val Ala Gln Ala Thr Val: 2085 2090 2095

### ` 37*177*

Thr Val Gln Val Leu Ala Cys Arg Glu Pro Glu Val Asp Val Val Leu 2100 2105 2110

Pro Leu Gln Val Leu Met Arg Arg Ser Gln Arg Asn Tyr Leu Glu Ala 2115 2120 2125

His Val Asp Leu Arg Asp Cys Val Thr Tyr Gln Thr Glu Tyr Arg Trp 2130 2135 2140

Glu Val Tyr Arg Thr Ala Ser Cys Gln Arg Pro Gly Arg Pro Ala Arg 2145 2150 2155 2160

Val Ala Leu Pro Gly Val Asp Val Ser Arg Pro Arg Leu Val Leu Pro 2165 2170 2175

Arg Leu Ala Leu Pro Val Gly His Tyr Cys Phe Val Phe Val Val Ser 2180 2185 2190

Phe Gly Asp Thr Pro Leu Thr Gln Ser Ile Gln Ala Asn Val Thr Val 2195 2200 2205

Ala Pro Glu Arg Leu Val Pro Ile Ile Glu Gly Gly Ser Tyr Arg Val 2210 2215 2220

Trp Ser Asp Thr Arg Asp Leu Val Leu Asp Gly Ser Glu Ser Tyr Asp 2225 2230 2235 2240

Pro Asn Leu Glu Asp Gly Asp Gln Thr Pro Leu Ser Phe His Trp Ala 2245 2250 2255

Cys Val Ala Ser Thr Gln Arg Glu Ala Gly Gly Cys Ala Leu Asn Phe 2260 2265 2270

Gly Pro Arg Gly Ser Ser Thr Val Thr Ile Pro Arg Glu Arg Leu Ala 2275 2280 2285

Ala Gly Val Glu Tyr Thr Phe Ser Leu Thr Val Trp Lys Ala Gly Arg 2290 2295 2300

Lys Glu Glu Ala Thr Asn Gln Thr Val Leu Ile Arg Ser Gly Arg Val 2305 2310 2315 2320

Pro Ile Val Ser Leu Glu Cys Val Ser Cys Lys Ala Gln Ala Val Tyr 2325 2330 2335

Glu Val Ser Arg Ser Ser Tyr Val Tyr Leu Glu Gly Arg Cys Leu Asn 2340 2345 2350

Cys Ser Ser Gly Ser Lys Arg Gly Arg Trp Ala Ala Arg Thr Phe Ser 5 2355 2360 2365

Asn Lys Thr Leu Val Leu Asp Glu Thr Thr Thr Ser Thr Gly Ser Ala 2370 2380

Gly Met Arg Leu Val Leu Arg Arg Gly Val Leu Arg Asp Gly Glu Gly 2385 2390 2395 2400

Tyr Thr Phe Thr Leu Thr Val Leu Gly Arg Ser Gly Glu Glu Glu Gly 2405 2410 2415

Cys Ala Ser Ile Arg Leu Ser Pro Asn Arg Pro Pro Leu Gly Gly Ser 2420 - 2425 2430

Cys Arg Leu Phe Pro Leu Gly Ala Val His Ala Leu Thr Thr Lys Val 2435 2440 2445

His Phe Glu Cys Thr Gly Trp His Asp Ala Glu Asp Ala Gly Ala Pro 2450 2455 2460

Leu Val Tyr Ala Leu Leu Leu Arg, Arg Cys Arg Gln Gly His Cys Glu 2465 2470 2475 2480

Glu Phe Cys Val Tyr Lys Gly Ser Leu Ser Ser Tyr Gly Ala Val Leu 2485 2490 2495

Pro Pro Gly Phe Arg Pro His Phe Glu Val Gly Leu Ala Val Val Val 2500 2505 2510

Gln Asp Gln Leu Gly Ala Ala Val Val Ala Leu Asn Arg Ser Leu Ala 2515 2520 2525

Ile Thr Leu Pro Glu Pro Asn Gly Ser Ala Thr Gly Leu Thr Val Trp 2530 2535 2540

Leu His Gly Leu Thr Ala Ser Val Leu Pro Gly Leu Leu Arg Gln Ala 2545 2550 2555 2560

Asp Pro Gln His Val Ile Glu Tyr Ser Leu Ala Leu Val Thr Val Leu 2565 2570 2575

Asn Glu Tyr Glu Arg Ala Leu Asp Val Ala Ala Glu Pro Lys His Glu-2580 2585 2590

Arg Gln His Arg Ala Gln Ile Arg Lys Asn Ile Thr Glu Thr Leu Val 2595 2600 2605

Ser Leu Arg Val His Thr Val Asp Asp Ile Gln Gln Ile Ala Ala Ala 2610 2615 2620

Leu Ala Gin Cys Met Gly Pro Ser Arg Glu Leu Val Cys Arg Ser Cys 2625 2630 2635 2640

Leu Lys Gln Thr Leu His Lys Leu Glu Ala Met Met Leu Ile Leu Gln 2645 2650 2655

Ala Glu Thr Thr Ala Gly Thr Val Thr Pro Thr Ala Ile Gly Asp Ser 2660 2665 2670

Ile Leu Asn Ile Thr Gly Asp Leu Ile His Leu Ala Ser Ser Asp Val 2675 2680 2685

Arg Ala Pro Gln Pro Ser Glu Léu Gly Ala Glu Ser Pro Ser Arg Met 2690 2695 2700

Val Ala Ser Gln Ala Tyr Asn Leu Thr Ser Ala Leu Met Arg Ile Leu 2705 2710 2715 2720

Met Arg Ser Arg Val Leu Asn Glu Glu Pro Leu Thr Leu Ala Gly Glu 2725 2730 2735

Glu Ile Val Ala Gin Gly Lys Arg Ser Asp Pro Arg Ser Leu Leu Cys 2740 2745 2750

Tyr Gly Gly Ala Pro Gly Pro Gly Cys His Phe Ser Ile Pro Glu Ala 2755 2760 2765

Phe Ser Gly Ala Leu Ala Asn Leu Ser Asp Val Val Gln Leu Ile Phe 2770 2775 2780

Leu Val Asp Ser Asn Pro Phe Pro Phe Gly Tyr Ile Ser Asn Tyr Thr 2785 2790 2795 2800

Val Ser Thr Lys Val Ala Ser Met Ala Phe Gln Thr Gln Ala Gly Ala 2805 2810 2815

Gln Ile Pro Ile Glu Arg Leu Ala Ser Glu Arg Ala Ile Thr Val Lys 2820 2825 2830

Val Pro Asn Asn Ser Asp Trp Ala Ala Arg Gly His Arg Ser Ser Ala 2835 2840 2845

Asn Ser Ala Asn Ser Val Val Val Gln Pro Gln Ala Ser Val Gly Ala 2850 2855 2860

Val Val Thr Leu Asp Ser Ser Asn Pro Ala Ala Gly Leu His Leu Gln 2865 2870 2875 2880

Leu Asn Tyr Thr Leu Leu Asp Gly His Tyr Leu Ser Glu Glu Pro Glu 2885 2890 2895

Pro Tyr Leu Ala Val Tyr Leu His Ser Glu Pro Arg Pro Asn Glu His 2900 2905 2910

Asn Cys Ser Ala Ser Arg Arg Ile Arg Pro Glu Ser Leu Gln Gly Ala 2915 2920 2925

Asp His Arg Pro Tyr Thr Phe Phe Ile Ser Pro Gly Ser Arg Asp Pro 2930 2935 2940

Ala Gly Ser Tyr His Leu Asn Leu Ser Ser His Phe Arg Trp Ser Ala 2945 2950 2955 2960

Leu Gln Val Ser Val Gly Leu Tyr Thr Ser Leu Cys Gln Tyr Phe Ser 2965 2970 2975

Glu Glu Asp Met Val Trp Arg Thr Glu Gly Leu Leu Pro Leu Glu Glu 2980 2985 2990

Thr Ser Pro Arg Gln Ala Val Cys Leu Thr Arg His Leu Thr Ala Phe 2995 3000 3005

Gly Ala Ser Leu Phe Val Pro Pro Ser His Val Arg Phe Val Phe Pro 3010 3015 3020

Glu Pro Thr Ala Asp Val Asn Tyr Ile Val Met Leu Thr Cys Ala Val 3025 3030 3035 3040

Cys Leu Val Thr Tyr Met Val Met Ala Ala Ile Leu His Lys Leu Asp 3045 3050 3055

- Gln Leu Asp Ala Ser Arg Gly Arg Ala Ile Pro Phe Cys Gly Gln Arg 3060 3065 3070
- Gly Arg Phe Lys Tyr Glu Ile Leu Val Lys Thr Gly Trp Gly Arg Gly 3075 3080 3085
- Ser Gly Thr Thr Ala His Val Gly Ile Met Leu Tyr Gly Val Asp Ser 3090 3095 3100
- Arg Ser Gly His Arg His Leu Asp Gly Asp Arg Ala Phe His Arg Asn 3105 3110 3115 3120
- Ser Leu Asp Ile Phe Arg Ile Ala Thr Pro His Ser Leu Gly Ser Val 3125 3130 3135
- Trp Lys Ile Arg Val Trp His Asp Asn Lys Gly Leu Ser Pro Ala Trp 3140 3145 3150
- Phe Leu Gln His Val Ile Val Arg Asp Leu Gln Thr Ala Arg Ser Ala 3155 3160 3165
- Phe Phe Leu Val Asn Asp Trp Leu Ser Val Glu Thr Glu Ala Asn Gly 3170 3175 3180
- Gly Leu Val Glu Lys Glu Val Leu Ala Ala Ser Asp Ala Ala Leu Leu 3185 3190 3195 3200
- Arg Phe Arg Arg Leu Leu Val Ala Glu Leu Gln Arg Gly Phe Phe Asp 3205 3210 3215
- Lys His Ile Trp Leu Ser Ile Trp Asp Arg Pro Pro Arg Ser Arg Phe 3220 3225 3230
- Thr Arg Ile Gln Arg Ala Thr Cys Cys Val Leu Leu Ile Cys Leu Phe 3235 3240 3245
- Leu Gly Ala Asn Ala Val Trp Tyr Gly Ala Val Gly Asp Ser Ala Tyr 3250 3255 3260
- Ser Thr Gly His Val Ser Arg Leu Ser Pro Leu Ser Val Asp Thr Val 3265 3270 3275 3280
- Ala Val Gly Leu Val Ser Ser Val Val Val Tyr Pro Val Tyr Leu Ala 3285 3290 3295
- Ile Leu Phe Leu Phe Arg Met Ser Arg Ser Lys Val Ala Gly Ser Pro 3300 3305 3310
- Ser Pro Thr Pro Ala Gly Gln Gln Val Leu Asp Ile Asp Ser Cys Leu 3315 3320 3325
- Asp Ser Ser Val Leu Asp Ser Ser Phe Leu Thr Phe Ser Gly Leu His 3330 3335 3340
- Ala Glu Ala Phe Val Gly Gln Met Lys Ser Asp Leu Phe Leu Asp Asp 3345 3350 3355 3360
- Ser Lys Ser Leu Val Cys Trp Pro Ser Gly Glu Gly Thr Leu Ser Trp 3365 3370 3375

Pro Asp Leu Leu Ser Asp Pro Ser The Val Gly Ser Asn Leu Arg Gln 3380 3385 3390

Leu Ala Arg Gly Gln Ala Gly His Gly-Leu Gly Pro Glu Glu Asp Gly 3395 3400 3405

Phe Ser Leu Ala Ser Pro Tyr-Ser Pro Ala Lys Ser Phe Ser Ala Ser 3410 3415 3420

Asp Glu Asp Leu Ile Gln Gln Val Leu Ala Glu Gly Val Ser Ser Pro 3425 3430 3435 3440

Ala Pro Thr Gln Asp Thr His Met Glu Thr Asp Leu Leu Ser Ser Leu
3445 3450 3455

Ser Ser Thr Pro Gly Glu Lys Thr Glu Thr Leu Ala Leu Gln Arg Leu 3460 3470

Gly Glu Leu Gly Pro Pro Ser Pro Gly Leu Asn Trp Glu Gln Pro Gln 3475 3480 3485

Ala Ala Arg Leu Ser Arg Thr Gly Leu Val Glu Gly Leu Arg Lys Arg 3490 3495 3500

Leu Leu Pro Ala Trp Cys Ala Ser Leu Ala His Gly Leu Ser Leu Leu 3505 3510 3515 3520

Leu Val Ala Val Ala Val Ala Val Ser Gly Trp Val Gly Ala Ser Phe 3525 3530 3535

Pro Pro Gly Val Ser Val Ala Trp Leu Leu Ser Ser Ser Ala Ser Phe 3540 3540 3550

Leu Ala Ser Phe Leu Gly Trp Glu Pro Leu Lys Val Leu Leu Glu Ala 3555 3560 3565

Leu Tyr Phe Ser Leu Val Ala Lys Arg Leu His Pro Asp Glu Asp Asp 3570 3575 3580

Thr Leu Val Glu Ser Pro Ala Val Thr Pro Val Ser Ala Arg Val Pro 3585 3590 3595 3600

With the transfer of the state of the state of

Arg Val Arg Pro Pro His Gly Phe Ala Leu Phe Leu Ala Lys Glu Glu 3605 3610 3615

Ala Arg Lys Val Lys Arg Leu His Gly Met Leu Arg Ser Leu Leu Val 3620 3625 3630

Tyr Met Leu Phe Leu Leu Val Thr Leu Leu Ala Ser Tyr Gly Asp Ala 3635 3640 3645

Ser Cys His Gly His Ala Tyr Arg Leu Gln Ser Ala Ile Lys Gln Glu 3650 3660

Leu His Ser Arg Ala Phe Leu Ala Ile Thr Arg Ser Glu Glu Leu Trp 3665 3670 3680

Pro Trp Met Ala His Val Leu Pro Tyr Val His Gly Asn Gln Ser 3685 3690 3695

Ser Pro Glu Leu Gly Pro Pro Arg Leu Arg Gln Val Arg Leu Gln Glu 3700 3705 3710

Ala Leu Tyr Pro Asp Pro Pro Gly Pro Arg Val His Thr Cys Ser Ala 3715 3720 3725

Ala Gly Gly Phe Ser Thr Ser Asp Tyr Asp Val Gly Trp Glu Ser Pro 3730 3735 3740

His Asn Gly Ser Gly Thr Trp Ala Tyr Ser Ala Pro Asp Leu Leu Gly 3745 3750 3755 3760

Ala Trp Ser Trp Gly Ser Cys Ala Val Tyr Asp Ser Gly Gly Tyr Val 3765 3770 3775

Gln Glu Ieu Gly Ieu Ser Ieu Glu Glu Ser Arg Asp Arg Ieu Arg Phe 3780 3785 3790

Leu Gln Leu His Asn Trp Leu Asp Asn Arg Ser Arg Ala Val Phe Leu 3795 3800 3805

Glu Leu Thr Arg Tyr Ser Pro Ala Val Gly Leu His Ala Ala Val Thr 3810 3815 3820

Leu Arg Leu Glu Phe Pro Ala Ala Gly Arg Ala Leu Ala Ala Leu Ser 3825 3830 3835 3840

Val Arg Pro Phe Ala Leu Arg Arg Leu Ser Ala Gly Leu Ser Leu Pro 3845 3850 3855

Leu Leu Thr Ser Val Cys Leu Leu Phe Ala Val His Phe Ala Val 3860 3865 3870

Ala Glu Ala Arg Thr Trp His Arg Glu Gly Arg Trp Arg Val Leu Arg 3875 3880 3885

Leu Gly Ala Trp Ala Arg Trp Leu Leu Val Ala Leu Thr Ala Ala Thr 3890 3895 3900

Ala Leu Val Arg Leu Ala Gln Leu Gly Ala Ala Asp Arg Gln Trp Thr 3905 3910 3915 3920

Arg Phe Val Arg Gly Arg Pro Arg Arg Phe Thr Ser Phe Asp Gln Val 3925 3930 3935

Ala His Val Ser Ser Ala Ala Arg Gly Leu Ala Ala Ser Leu Leu Phe 3940 3945 3950

Leu Leu Val Lys Ala Ala Gln His Val Arg Phe Val Arg Gln Trp 3955 3960 3965

Ser Val Phe Gly Lys Thr Leu Cys Arg Ala Leu Pro Glu Leu Leu Gly 3970 3980

Val Thr Leu Gly Leu Val Val Leu Gly Val Ala Tyr Ala Gln Leu Ala 3985 3990 3995 4000

Ile Leu Leu Val Ser Ser Cys Val Asp Ser Leu Trp Ser Val Ala Gln 4005 4010 4015

Ala Leu Leu Val Leu Cys Pro Gly Thr Gly Leu Ser Thr Leu Cys Pro 4020 4025.

Ala Glu Ser Trp His Leu Ser Pro Leu Leu Cys Val Gly Leu Trp Ala 4035 4040 4045

Leu Arg Leu Trp Gly Ala Leu Arg Leu Gly Ala Val Ile Leu Arg Trp 4050 4055

Arg Tyr His Ala Leu Arg Gly Glu Leu Tyr Arg Pro Ala Trp Glu Pro 4065 4070 4075 4080

Gln Asp Tyr Glu Met Val Glu Leu Phe Leu Arg Arg Leu Arg Leu Trp 4085 4090 4095

Met Gly Leu Ser Lys Val Lys Glu Phe Arg His Lys Val Arg Phe Glu 4100 4105 4110

Gly Met Glu Pro Leu Pro Ser Arg Ser Ser Arg Gly Ser Lys Val Ser 4115 4120 4125

Pro Asp Val Pro Pro Pro Ser Ala Gly Ser Asp Ala Ser His Pro Ser 4130 4135 4140

Thr Ser Ser Ser Gln Leu Asp Gly Leu Ser Val Ser Leu Gly Arg Leu 4145 4150 4160

Gly Thr Arg Cys Glu Pro Glu Pro Ser Arg Leu Gln Ala Val Phe Glu
4165 4170 4175

Ala Leu Leu Thr Gln Phe Asp Arg Leu Asn Gln Ala Thr Glu Asp Val 4180 4185

Tyr Gln Leu Glu Gln Gln Leu His Ser Leu Gln Gly Arg Arg Ser Ser 4195 4200 4205

Arg Ala Pro Ala Gly Ser Ser Arg Gly Pro Ser Pro Gly Leu Arg Pro 4210 4215 4220

Ala Leu Pro Ser Arg Leu Ala Arg Ala Ser Arg Gly Val Asp Leu Ala 4225 4230 4235 4240

Thr Gly Pro Ser Arg Thr Pro Ser Gly Gln Glu Gln Gly Pro Pro Gln
4245
4250
4255

Gln His Leu Val Leu Leu Pro Gly Gly Gly Gly Pro Trp Ser Arg Ser 4260 4265 4270

Gly His Arg Ser Val Leu Leu Ser Ala Ala Val Lys Ala Glu Gly Gln 4275 4280 4285

Ala Glu Trp Leu His Val Gly Ser Pro Glu Ser Arg Gln Gly His Leu 4290 4295 4300

Ser Val Cys Gly Leu Gln His Phe Lys Glu Ala Val Trp Pro Thr Arg 4305 4310 4315 4320

Thr Gln Gly Pro Leu Pro Ser Ser Leu Gly Lys Asp Thr Ala Val Leu 4325 4330 4335

Asp Gly Phe

Figure 10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3: (Compare Figure 7) CTC AAC GAG GAG CCC CTG ACG CTG GCC GAG GAG ATC GTG GCC CAG Leu Asn Glu Glu Pro Leu Thr Leu Ala Gly Glu Glu Ile Val Ala Gln 4355 4350 **434**5 GOC AAG COC TOO GAC COC COC AGC CTG CTG TGC TAT GOC GOC GOC A Gly Lys Arg Ser Asp Pro Arg Ser Leu Leu Cys Tyr Gly Gly Ala Pro 4365 4360 GGG CCT GGC TGC CAC TTC TCC ATC CCC GAG GCT TTC AGC GGG GCC CTG Gly Pro Gly Cys His Phe Ser Ile Pro Glu Ala Phe Ser Gly Ala Leu 4380 4375 GOC AAC CTC AGT GAC GTG GTG CAG CTC ATC TIT CTG GTG GAC TGC AAT 192 Ala Asn Leu Ser Asp Val Val Gln Leu Ile Phe Leu Val Asp Ser Asn 4395 4390 COC TIT COC TIT GGC TAT ATC AGC AAC TAC ACC GTC TOC ACC AAG GTG Pro Phe Pro Phe Gly Tyr Ile Ser Asn Tyr Thr Val Ser Thr Lys Val 4410 4415 GOO TOG ATG GOA TTC CAG ACA CAG GOO GOO GOO CAG ATC COO ATC GAG Ala Ser Met Ala Phe Gln Thr Gln Ala Gly Ala Gln Ile Pro Ile Glu 4430 4425 OGG CTG GOO TOA GAG GOO GOO ATC ACC GTG AAG GTG COO AAC AAC TOG Arg Leu Ala Ser Glu Arg Ala Ile Thr Val Lys Val Pro Asn Asn Ser 4445 4440 **44**50 CAC TOG OCT OCC OGG OGC CAC OGC AGC TOC OCC AAC TOC OCC AAC TOC Asp Trp Ala Ala Arg Gly His Arg Ser Ser Ala Asn Ser Ala Asn Ser 4465 4455 432 GIT GIG GIC CAG CCC CAG GCC TCC GIC GGT GCT GIG GIC ACC CIG GAC Val Val Gln Pro Gln Ala Ser Val Gly Ala Val Val Thr Leu Asp 4480 4475 AGC AGC AAC OCT GOG GOC GOG CTG CAT CTG CAG CTC AAC TAT ACG CTG 480 Ser Ser Asn Pro Ala Ala Gly Leu His Leu Gln Leu Asn Tyr Thr Leu 4490 CTG GAC GGC CAC TAC CTG TCT GAG GAA OCT GAG COC TAC CTG GCA GTC 528 Leu Asp Gly His Tyr Leu Ser Glu Glu Pro Glu Pro Tyr Leu Ala Val 4510 4505 TAC CTA CAC TOG GAG COC COG COC AAT GAG CAC AAC TGC TOG GCT AGC 576 Tyr Leu His Ser Glu Pro Arg Pro Asn Glu His Asn Cys Ser Ala Ser 4525 4520 624 AGG AGG ATC COC CCA GAG TCA CTC CAG GGT GCT GAC CAC CGG CCC TAC Arg Arg Ile Arg Pro Glu Ser Leu Gln Gly Ala Asp His Arg Pro Tyr 4545 4535 4540 ACC TTC TTC ATT TOC COG GGG AGC AGA GAC CCA GGG GGG AGT TAC CAT 672 Thr Phe Phe Ile Ser Pro Gly Ser Arg Asp Pro Ala Gly Ser Tyr His 4555 4550 . . CTG AAC CTC TOO AGO CAC TTC COO TOG TOG GOG CTG CAG GTG TOO GTG 720 Leu Asn Leu Ser Ser His Phe Arg Trp Ser Ala Leu Gln Val Ser Val

4570

e e e e e e e e e e e e e e e e e e e	45/77		
Gly Leu Tyr-Thr Ser 4580	CTG TGC CAG TAC TTC AG Leu Cys Gln Tyr Phe S 4585	590 4	595
Trp Arg Thr Glu Gly 460	· ± .	4610	
Ala Val Cys Leu Thr 4615	OCC CAC CTC ACC GCC T Arg His Leu Thr Ala P 4620	4625	
Val Pro Pro Ser His 4630	GIC OSC TITT GIG TITT O Val Arg Phe Val Phe P 4635	4640	
Val Asn Tyr Ile Val 4645	ATG CTG ACA TGT GCT G Met Leu Thr Cys Ala V 4650	4655	
ATG GTC ATG GCC GCC Met Val Met Ala Ala 4660	C ATC CTG CAC AAG CTG G A 11e 1eu His Lys 1eu A 4665	AC CAG TIG GAT GOC A Sp Gln Leu Asp Ala 9 1670	AGC - 1008 Ser 1675
CCC CCC CCC ATT Arg Gly Arg Ala III	C CCT TIC TGT GGG CAG C Pro Phe Cys Gly Gln A 30 4685	4690	The orbitals
GAG ATC CTC GTC AA Glu Ile Leu Val Ly 4695	3 ACA GGC TGG GGC CGG C s Thir Gly Trp Gly Arg C 4700	SCC TCA GGT ACC ACG Gly Ser Gly Thr Thr 4705	300 1104
	G CTG TAT GGG GTG GAC. 1 t Leu Tyr Gly Val Asp 9 4715	AGC CGG AGC GGC CAC: Ser Arg Ser Gly His	00G 1152 Arg
CAC CTG GAC GOC GA His Leu Asp Gly As 4725	C AGA GOC TIC CAC GOC p Arg Ala Phe His Arg 4730	AAC AGC CTG GAC ATC Asn Ser Leu Asp Ile 4735	TTC 1200 Phe
OGG ATC GCC ACC CO Arg Ile Ala Thr Pr 4740	G CAC AGC CTG GGT AGC TO His Ser Leu Gly Ser 4745	GTG TOG AAG ATC CGA Val Trp Lys Ile Arg 4750	GIG 1248 Val 4755
Tro His Aso Aso Ly	A GGG CTC AGC CCT GCC rs Gly Leu Ser Pro Ala 760 . 47,65	TIP FIE DEG OET 1220	. —
ATC GTC AGG GAC C Ile Val Arg Asp L 4775	rg CAG ACG GCA CGC AGC eu Gln Thr Ala Arg Ser 4780	GCC TITC TITC CTG GTC Ala Phe Phe Leu Val 4785	AAT 1344 Asn
mai am mar d	ic cac ace cac coc aac al Glu Thr Glu Ala Asn 4795	GGG GGC CTG GTG GAG Gly Gly Leu Val Glu 4800	AAG 1392 Lys
GAG GTG CTG GCC G Glu Val Leu Ala A 4805	CG AGC GAC GCA GCC CIT la Ser Asp Ala Ala Leu 4810	THE COC THE COS COC Leu Arg Phe Arg Arg 4815	Leu
CTG GTG GCT GAG C Leu Val Ala Glu I 4820	TG CAG CGT GGC TIC TIT eu Gln Arg Gly Phe Phe 4825	GAC AAG CAC ATC TOS Asp Lys His Ile Try 4830	CIC 1488 Leu. 4835

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TCC / Ser :	ATA Ile	Trp Trp	GAC Asp	CCG. Arg 4840	Pro.	CT Pro	Arg Arg	Ser	OGT Arg 4845	Phe	ACT Thr	CCC Arg	ATC Ile	CAG Gln 4850	Arg		1536
CC A	ACC Ihr	TGC Cys	TGC Cys 4855	Val	CTC Leu	CTC Leu	ATC Ile	TGC Cys 4860	Leu	TIC Phe	CIG Leu	GC Gly	Ala 4865	Asn	CCC Ala		1584
GIG :	rcc Irp	TAC Tyr 4870	Gly	CCT Ala	GTT Val	Gly	GAC Asp 4875	Ser	CC Ala	TAC Tyr	AGC Ser	ACG Thr 4880	Gly	CAT His	GIG Val	;	1632
TCC-7	ACG Arg 4885	Leu	AGC Seir	OCG Pro	CTG Leu	AGC Ser 4890	Val	GAC Asp	ACA Thr	val GIC	CCT Ala 4895	Val	GC Gly	CIG Leu	GTG Val	· • ·	1680
TCC 7 Ser 5 4900	Ser	GTG Val	GTT Val	Val	TAT Tyr 4905	Pro	GTC Val	TAC Tyr	CIG Leu	CCC Ala 4910	Ile	CTT Leu	TTT Phe	CIC	TTC Phe 4915	<b>.</b>	1728
CCG A	ATG Met	TCC Serj	yrg œ	AGC Ser 4920	Lys	Val Val	CCT Ala	Gly	AGC Ser 4925	Pro	AGC Ser	Pro	Thr	CCT Pro 4930	Ala	-	1 <b>7</b> 76
Gly (	CAG Gln	CAG Gln	GIG Val 4935	Leu	GAC Asp	ATC Ile	Asp Asp	Ser	TGC Cys )	Leu	GAC Asp	TOG Ser	Ser	GIG Val		1 28	1824
GAC A	AGC Ser	TCC Ser 4950	Phe	CIC Leu	ACG Thr	TTC Phe	TCA Ser 495	Gly	CIC Leu!	CAC His	CCT Ala	GAG Glu 4960	Ala	TTT Phe	GIT Val		1872
Gly (		Met					Phe					Lys			GTG:		1920
TGC ' Cys ' 4980	Trp	CCC Pro	TCC Ser	Gly	GAG Glu 498	Gly	ACG Thr	CIC	AGT Ser	TCG Trp 4990	Pro	GAC Asp	CIG Leu	CIC Leu	AGT Ser 4995		1968
GAC ( Asp )	OG Pro	TCC Ser	Ile	GTG Val 5000	Gly	AGC Ser	AAT Asn	Leu	Arg 500	Gln	CIG Leu	GCA Ala	Arg	GC Gly 501	Gln		2016
Ala	GC Gly	CAT His	GGG Gly 501	Leu	Gly	CCA Pro	GAG Glu	GAG Glu 502	Asp	Gly	TTC	TCC Ser	CIG Leu 502	_Ala	AGC Ser		2064
œ Pro	TAC Tyr	TCG Ser 503	Pro	Ala	aaa Lys	TCC Ser	TTC Phe 503	Ser	GCA Ala	TCA Ser	GAT Asp	GAA Glu 504	Asp	CIG Leù	ATC Ile	•	2112
Gln	CAG Gln 504	Val	CIT	∞ Ala	GAG Glu	GGG Gly 505	Val	AGC Ser	AGC Ser	CA Pro	Ala 505	-Pro	ACC Thr	CAA Gln	GAC Asp	. •	2160
2ACC Thr 5060	His	C AT Met	G GA Glu	A AC	G GA Asp 506	Leu	G CI Leu	C AG Ser	C AG Ser	C CT Leu 507	Ser	C AG Ser	C AC	T CC Pro	T GGG Gly 5075		2208

									7/7	7									
GA G1	G AA( u Ly:	G AC	A GAO	ACC Thi 508	Leu	G Ala	CIO Leu	CAC Glr	ACC Arg 508	Leu	GC	GAC Glu	CIO Leu	G GC G15	Pro	A.		2256	
8Ci Pro	C AC Sez	C C Pro	CA GO Gly 509	7 Lèi	NG AA I Asn	C TG	G GA Glü	A CA Glr 510	Pro	C CA	G GC Ala	A GC Ala	G AG Arg 510	Leu	G TO Ser	r	•	2304	
AG	G ACA y Thi	GG2 Gly 511	CIC Leu .0	GIC Val	GAG Glu	Gly	CIG Leu 511	Arfg	AAG	Arg	CIG	CIG Leu 512	Pro	Ala	TI	; ) -		2352	
TG Cys	GCC 5 Ala 512	Ser	CÍG	Ala	CAC His	G1y 513	Leu	AGC Ser	CIG	CIC	CIG Leu 513	Val	GCT Ala	GIG Val	GCI Ala		- :	2400	
GTC Val 514	. Ala	GIC Val	TCA Sei	Gly	TGG Trp 514	Val	GIY	∞ Ala	AGC Ser	TTC Phe 515	Pro	OCG Pro	GC Gly	GIG Val	AGT Ser 515			2448	\$ 34%
Gin Val	CCG Ala	TGG	CIC	CIG Leu 516	Ser	AGC Ser	AGC Ser	Ala	AGC Ser 516	Phe	CIG Leu	CCC Ala	TCA Ser	TTC Phe 517	Leu	70		2496	: # :#:
Gly	TYP	GAG Glu	CCA Pro 517	Leu	AAG Lys	GIC Val	TTG Leu	Leu	GAA Glu )	Ala	CIG Leu	TAC	Phe	TCA Ser	ĆIG Leu	• •		2544	· 集
GIG Val	GCC Ala	AAG Lys 519	OGG Arg O	CIG Leu	CAC His	CCG Pro	GAT Asp 5195	Glu	CAT Asp	GAC Asp	ACC Thr	CIG Leu 5200	Val	GAG Glu	ACC Ser			2592	
Pro	GCT Ala 520	var.	ACG Th <u>i</u> r	CCT Pro	GIG: Val	AGC Ser 5210	Ala	CGT: Arg	GIG Val	OCC Pro	OGC Arg. 5215	Val:	Arg	CCA Pro	occ Pro	А	•	2640	
CAC His 522	GIY	TTT Phe	GCA Ala	CIC Leu	TTC Phe 5225	Leu	CCC Ala	AAG Lys	GAA Glu	GAA Glu 5230	Ala	Arg	AAG Lys	GTC Val	AAG Lys 523	- ,		2688-	
AGG Arg	CTA Leu	CAT His	GLY	ATG Met 5240	Leu,	CCG Arg	AGC Ser	CIC Leu	CIG Leu 5245	Val	TAC Tyr	ATG: Met	CIT Leu	TTT Phe 5250	Leu	tage et a		2736	
CIG	Val GTG	'ACC Thr	CIG Leu 5255	Leu	CCC Ala	ACC Ser	Tyr	GGG Gly 5260	Asp	GCC: Ala	TCA Ser	CA2 .	CAT His 5265	Gly	CAC His	·		2784	
GCC Ala	TAC Tyr	CGT Arg 5270	CTG Leu )	CAA Gln	AGC Ser	Ala	ATC Ile 5275	Lys	CAG CAG	GAG Glu	CTG Leu	CAC His 5280	Ser /	OGG Arg	CCC Ala	: 5 :	., 2	2832	
TTC Phe	CTG Leu 5285	ΑТФ	ATC Ile	ACG Thr	Arg.	TCT Ser 5290	Glu	GAG Glu	CIC Leu	Trp.	CA Pro 5295	Trp	ATG Met	CC Ala	CAC His	<b>~</b> .	2	2880	
GIG Val 5300	Leu	CIG Leu	Pro ,	IÄL	GTC Val 5305	HIS	Gly Gly	AAC Asn	Gln;	TCC Ser 5310	Ser.	CCA Pro	GAG Glu	Leu	GG Gly 5315	,··	2	2928	
$\infty$	ŒΆ	œ	CTG	œ	CAG	GIG	œ	CIG	CAG	GAA	CCA	CIC	TAC	CCA.	GAC		2	2976	

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		CCG Arg			Gln		Airg		GĽn	Glu					Asp	2976
CCT Pro	Pro	Gly	Pro 533	Arg	GTC Val	CAC His	Thr	TGC Cys 534	Ser	Ala	GCA Ala	Gly	GC Gly 534	Phe	AGC Ser (	3024
		GAT Asp 5350	Tyr			Gly		Glu			His		Gly		Gly	3072
		ecc Ala 5			Ala		Asp			Gly		Trp				3120
	Cys	∞ Ala				Ser.					Gln					3168
AGC Ser	CTG Leu	GAG Glu	GAG Glu	AGC Ser 5400	Arg	GAC Asp	CCG Arg	Leu.	Arg	TTC Phe	Leu	Gln	CIG Leu	His		3216
TGG Trp	CTG Leu	GAC Asp	AAC Asn 5415	Arg.	AGC Ser	Arg	Ala	.Val.	Phe	CIG Leu	Glu	Leu	Thr	Arg	TAC Tyr	3264
AGC	œ	œ	CITC	222	CHIC.	030	~~	~~	~~~				~~~	C3.C		3312
Ser	Pito	Ala 5430	Val'	Gly	Leu	His	Ala	Ala	Val	Thr	Leu	Arg 5440	Leu	Glu	Phe	3312
Ser	Pitto GCG	Ala 5430 CC Ala	occ val	CCC Gly	Leu CC Ala	His CTG Leu	Ala 5435 CCC Ala	Ala ccc	Val	Thr	Leu GIC	Arg 5440 CC Arg	Leu ) ccc	Glu	Phe Coo	3360
Ser CG Pro	ecc Ala 5445 ecc Arg	Ala 5430 CC Ala	CIC GJA GC AJ	COC Arg	CCC Ala	CTG Leu 5450 GGC Gly	Ala 5435 GCC Ala )	Ala CCC Ala TCG	CTC Leu CTG Leu	Thr AGC Ser	CTC Val 5455 CTG Leu	Arg 5440 CC Arg	CC Pro	Glu TTT Phe TCG	Phe Ala GIG	
Ser  CIG Leu 5460	CTG	Ala 5430 GCC Ala GCC Arg	Val´ ) GGC Gly CTC Leu CTG Leu	Gly CCC Arg ACC Ser	CCC Ala S465	CTG Leu 5450 GGC Gly GTG Val	Ala 5435 CCC Ala ) CIC Leu	Ala  CCC Ala  TCC Ser  TTC Phe	CTC Leu CTG Leu	AGC Ser CCT Pro 5470 GTG Val	GIC Val 5455 CIG Leu	Arg 5440 CCC Arg CTC Leu GAG Glu	Leu Pro ACC Thr	Glu TTT Phe TCG Ser CGT Arg	COG Ala GIG Val 5475 ACT Thr	3360
Ser CCG Pro CTG Leu 5460 TCC Cys	GCC Ala 5445 CCC Arg CTC Leu	Ala 5430 Ala CCC Arg CTG Leu AGG	Val' ) CCC Gly CTC Leu CTG Leu GAA	CGC Arg  AGC Ser  TTC Phe 5480  GGG Gly	CCC Ala S465	CTG Leu 5450 GGC Gly GTG Val	Ala 5435 CCC Ala CTC Leu CAC His	Ala  TOG Ser  TIC Phe	CTC Leu CTG Leu SCC Ala 5485 CTG Leu	AGC Ser CCT Pro 5470 GTG Val	GIC Val 5455 CIG Leu CCC Ala	Arg 5440 CCC Arg CTC Leu GAG Glu	Leu CCC Pro ACC Thr CCC Ala	TTT Phe TCG Ser CGT Arg 5490 TCG TTp	COG Ala GIG Val 5475 ACT Thr	3360 3408
Ser  CG Pro  CIG Leu 5460  TCC Cys  TGG Trp	Pito GCG Ala 5445 GCC Arg CTG Leu CAC His	Ala 5430 CCC Ala CCC Arg CTG Leu AGG Arg	CIC Leu CAA Glu 5495	Gly  CGC Arg  AGC Ser  TTC Phe 5480 GGly GTG	Leu  CCC Ala  CCC Ala 5465 CCC Ala Arg  CCC CCC CCC CCC CCC CCC CCC CCC CCC	His CIG Leu 5450 GGC Gly GTG Val TGG Trp	Ala 5435 GCC Ala CIC Leu CAC His CCC Arg	Ala TOG Ser TTIC Phe GIG Val 5500	CTC Leu CTG Leu CTG Leu CTG Leu CTG Leu Ala S485	AGC Ser CCT Pro 5470 GIG Val ACG	GIC Val. 5455 CIG Leu GCC Ala CIC Leu	Arg 5440 CCC Arg CIC Leu GAG Glu CGA GGA CGA CGA	ACC Thr ACC Ala S5505	Glu TTTT Phe TCG Ser CGT Arg 5490 TCG TTCC TCG TCCC	CIC	3360 3408 3456
Ser  CG Pro  CIG Leu 5460  TCC Cys  TGG Trp  CGG Arg	Pito GCG Ala 5445 GCC Arg CTG Leu CAC Trp CAG	Ala 5430 GCC Ala CCC Arg CTG Leu AGG ATG CTG Leu 55510 CTG Leu	CIC Leu CAA CIG Leu CIG	Gly  CCC Arg  ACC Ser  TTC Phe 5480  GCG Gly  GTG Val	Leu  CCC Ala  CCC Ala  S465 CCC Ala  CCC Ala  CCC CCC  CCC CCC CCC CCC CCC CCC CCC	His CTG Leu 5450 GCC GIy GTG Val TCG TTp CTG Leu GAC	Ala 5435 CCIC Ala CIC Leu CAC His Arg ACG Thr 55515 CCC Arg	Ala TOG Ala TOG Ser TTC Phe GIG Val 5500 GCG Ala	CTC Leu CTG Leu CTG Leu CTG Leu CTG Ala 5485 CTG Ala TTGG	AGC Ser CCT Pro 5470 GIG Val ACG Thr	GIC Val. 5455 CIG Leu GCC Ala CIC Leu GCC GCA Ala	Arg 5440 CCC Arg CCC Leu GAG GLu CCG LEu CCAG GLU CCG TTC Phe	ACC Thr  CCC Ala  CSC GTA  Val  CGC  CGC  CGC  CGC  CGC  CGC  CGC  C	Glu TITI Phe TOG Ser CGT Arg 5490 TOG TOG TOG TOG CCC	CIC Leu	3360 3408 3456 3504

<b>4</b> , <b>4</b> 5		21 1 1 3 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	<i>:</i>
Ala Ala Arg Gly L		CCC TTC CTG CTT TTG GTC AAG Leu Phe Leu Leu Leu Val Lys 5565 5570	3696
			3744
		CIG GGG GTC ACC TIG GGG CTG Leu Gly Val Thr Leu Gly Leu 5600	3792
		CTG GCC ATC CTG CTC GTG TCT Leu Ala Ile Leu Leu Val Ser 5615	3840
TOO TOT GTG GAC TO Ser Cys Val Asp S 5620	CC CTC TCG ACC GTG er Leu Trp Ser Val 5625	CCC CAG CCC CTG TTG GTG CTG Ala Gln Ala Leu Leu Val Leu 5630 5635	3888
Cys Pro Gly Thr G	GG CTC TCT ACC CTG ly Leu Ser Thr Leu 640	TGT CCT CCC GAG TCC TGG CAC Cys Pro Ala Glu Ser Trp His 5645 5650	3936
CTG TCA CCC CTG C Leu Ser Pro Leu L 5655	eu Cys Val Gly Leu 566		3984
GCC CTA CGG CTG GG Ala Leu Arg Leu G 5670	GG GCT GIT ATT CTC	CGC TGG CGC TAC CAC GCC TTG Arg Trp Arg Tyr His Ala Leu 5680	4032
CGT CGA GAG CTG TA Arg Gly Glu Leu T 5685	AC CCG CCC CCC TCC yr Arg Pro Ala Trp 5690	GAG CCC CAG GAC TAC GAG ATG Glu Pro Gln Asp Tyr Glu Met 5695	4080
GIG GAG TIG TIC C Val Glu Leu Phe L 5700	TG CGC AGG CTG CGC eu Arg Arg Leu Arg 5705	CTC TGG ATG GGC CTC AGC AAG Leu Trp Met Gly Leu Ser Lys 5710 5715	4128
Val Lys Glu Phe A	GC CAC AAA GTC COC rg His Lys Val Arg 720	TIT GAA GGG ATG GAG CGG CTG Phe Glu Gly Met Glu Pro Leu 5725 5730	4176
CCC TCT CGC TCC TC Pro Ser Arg Ser Se 5735	CC AGG GGC TCC AAG er Arg Gly Ser Lys 574	GTA TOC COG GAT GTG COC CCA TOTAL Val Ser Pro Asp Val Pro Pro 0 5745	4224
CCC ACC GCT GCC TV Pro Ser Ala Gly Se 5750	CC GAT GCC TCG CAC er Asp Ala Ser His 5755	CCC TCC ACC TCC TCC ACC CAG Pro Ser Thr Ser Ser Ser Gln 5760	4272
CIG GAT GGG CIG A Leu Asp Gly Leu S 5765	OC GIG ACC CIG GOC er Val Ser Leu Gly 5770	CGG CTG GGG ACA AGG TGT GAG Arg Leu Gly Thr Arg Cys Glu 5775	4320
CCT GAG CCC TCC Co Pro Glu Pro Ser A 5780	GC CTC CAA GCC GTG rg Leu Gln Ala Val 5785	TTC GAG GCC CTG CTC ACC CAG Phe Glu Ala Leu Leu Thr Gln 5790 5795	4368

<i>301,1,1</i>	
TTT GAC CGA CTC AAC CAG GCC ACA GAG GAC GTC TAC CAG CTG GAG CAG Phe Asp Arg Leu Asn Gln Ala Thr Glu Asp Val Tyr Gln Leu Glu Gln 5800 5805 5810	4416
CAG CTG CAC AGC CTG CAA GGC CGC AGG AGC AGC CGG GGC GCC GGC GG	4464
TCT TCC CGT GGC CCA TCC CCG GGC CTG CGG CCA GCA CTG CCC AGC CGC Ser Ser Arg Gly Pro Ser Pro Gly Leu Arg Pro Ala Leu Pro Ser Arg 5830 5835 5840	4512
CIT CCC CCG CCC AGT CCG CGT GTG GAC CTG CCC ACT GCC CCC ACC AGG Leu Ala Arg Ala Ser Arg Gly Val Asp Leu Ala Thr Gly Pro Ser Arg 5845 5850 5855	<b>4</b> 560
ACA CCT TOG GOC CAA GAA CAA GGT CCA CCC CAG CAG CAC TTA GTC CTC Thr Pro Ser Gly Gln Glu Gln Gly Pro Pro Gln Gln His Leu Val Leu 5860 5865 5870 5875	4608
CTT CCT CCC CCC CCG CCG CCG TCG ACT CCC ACT CCA CAC CCC TCA GTA Leu Pro Gly Gly Gly Pro Trp Ser Arg Ser Gly His Arg Ser Val 5880 5885 5890	4656
TTA CTT TCT GCC GCT GTC AAG GCC GAG GCC CAG GCA GAA TGG CTG CAC Leu Leu Ser Ala Ala Val Lys Ala Glu Gly Gln Ala Glu Trp Leu His 5895 5900 5905	4704
GTA GGT TCC CCA GAG AGC AGG CAG GGG CAT CTG TCT GTC TGT GGG CTT Val Gly Ser Pro Glu Ser Arg Gln Gly His Leu Ser Val Cys Gly Leu 5910 5915 5920	4752
CAG CAC TIT AAA GAG GCT GIG TGG CCA ACC AGG ACC CAG GGT CCC CIC Gln His Phe Lys Glu Ala Val Trp Pro Thr Arg Thr Gln Gly Pro Leu 5925 5930 5935	4800
CCC AGC TCC CTT GGG AAG GAC ACA GCA GTA TTG GAC GGT TTC Pro Ser Ser Leu Gly Lys Asp Thr Ala Val Leu Asp Gly Phe 5940 5945 5950	4842
TAGOCTICTIGA GATGCTAATT TATTTCCCCG AGTCCTCAGG TACAGCGGGC TGTGCCCCGCC	4902
COCACCCCT GGGCAGATGT CCCCCACTGC TAAGGCTGCT GGCTTCAGGG AGGGTTAGCC	4962
2TGCACCGCC CCAACCTTAC TACCTCTCCA GTTCCTACCG TACTCCCTGC	5022
ACCEPTACE TOTAL CONTROL OF THE ACCEPTANCE OF THE	5082
GTATGTCACT ATTITCACTA GGGCTGAGGG GCCTGCGCCC AGAGCTGGCC TCCCCCAACA	5142
CCTCCTCCCC TTCGTACGTG TCGTCCCGTT ATCCCACCCC CCCTCCTCCT TCGATCCCAG	5202
CITIGGCCTIG GGCCGGTGCT GGGGGCACAG CIGICTGCCA GGCACTCTCA TCACCCCAGA	5262
GCCCTTGTCA TCCTCCCTTG CCCCAGGCCA GGTAGCAAGA GAGCAGCCCC CAGGCCTGCT	5322
GGCATCAGGT CTGGGCAAGT AGCAGGACTA GGCATGTCAG AGGACCCCAG GGTGGTTAGA	5382
GCAAAAGACT OCTOCTGGGG GCTGGCTCCC AGGGTGGAGG AAGGTGACTG TGTGTGTGTG	5442
TGTGTGCCCC CCCCACCCCC GAGTGTCCTG TATCCCCCAG CCACCCTCAA CCCCCTCCCA	5502

SUBSTITUTE SHEET (RULE 26)

CCT	ECT(	TG (	XIG.	cric	rg, T	TÀC	CACT	ı Çik	उर्देख	CAT	œ	œi.	rcr /	4GAQ	αiα	SA ·	5562	2
CAC		CA Z	ACCC		AC CI	AAGC/	AGACA	A AAC	SICA	ATAA	AAG	AGCIY	ELC ,	rgác	IGCA/	AA ·	5622	2
AAA	VAAAJ	¥A		*-			=	*				• • 4.			i.u	<b>,-</b>	5633	L .
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	î.: (2	ದ)	SEQ	JENCI	E DES	CRI	ai joi	v: Si	o ii	ON C	4:	(Car	mpare	e Fig	jure	7)		
Leu 1	Asn	Glu	Glu ~.	Pro . 5	Leu	Thr	Leu	Ala	Gly 10	Glu	Glu	Ile	Val	Ala 15	Gln		- :	٠,-
Gly	Lys	Arg	Ser 20	Asp	Pro	Arg	Ser	Leu 25	Leu	Cys	Tyr	Gly	Gly 30	Ala	Pro			: f
Gly	Pro	Gly 35	Cys	His	Phe	Ser	Ile 40	Pro	Glu	Ala	Phe	Ser 45	Gly	Ala	Leu		,	:**
Ala	Asn 50	Leu	Ser	Asp	Val	Val 55	Gln	Leu	Ile	Phe	Leu 60	Val	Asp	Ser ·	Asn			
Pro 65	Phe	Pro	Phe	Gly	Tyr 70	Ile	Ser	Asn	Tyr	Thr 75	Val	Ser	Thr	Lys :	Val 80		. 4	
Ala	Ser	Met	Ala	Phe 85	Gln	Thr	Gl'n	Ala	Gly 90	Ala	Gln	Ile	Pro	Ile 95	Glu	. IF & D 98		i Ali
Arg	Leu	Ala	Ser 100	Glu	Arg	Ala	Ile	Thr 105	Val	Lys	Val	Pro	Asn 110		Ser	-	•	. ,=
Asp	Trp	Ala 115	Ala	Arg	Gly	His	Arg 120	Ser	Ser	Ala	Asn	Ser 125	Ala	Asn	Ser	: •	. <i>I</i>	•••
Val	Val 130	Val	Gln	Pro	Gln	Ala 135	Ser	Val	Gly	Ala	Val 140	Val	Thr 	Leu	Asp			-
Ser 145	Ser	Asn	Pro	Ala	Ala 150	Gly	Ļeų	His	Ļeu	Gln 155	Leu	Asn	Tyr	Thr	L <u>e</u> u 160		•	
Leu	Asp	Gly	His	Tyr 165	Leu	Ser	Glu	Glū	Pro 170	Glu	Pro	Tyr	Leu	Ala 175	Val			
Tyr	Leu	His	Ser 180	Glu	Pro	Arg	Pro	Asn 185	Glu	His	Asn	Cys	Ser 190	Ala	Ser		· • . "	· 💉
Arg 	Arg	Ile 195	Ärg	Pro	Glu	Ser 	Leu 200	Gln	Gly	Ala	Asp	His 205	Arg :	Pro	Tyr	•	J &	•
	Phe 210	Phe	_Ile	Ser	Pro	Gly 215	Ser	Arg	Asp	Pro	Ala 220	Gly	,Ser	Tyr	His	٠.	•	<i>:</i> .
Leu 225	Asn	Leu	Ser	Ser	His 230	Phe	Arg	Trp	Ser	Ala 235	Leu	Gln 	Val	Ser	Val 240		<b>^</b> .	•
Gly	Ļeu	Tyr	Thr	Ser 245		Cys	Gln	Tyr	Phe 250	Ser	Ģlu	Glu	Asp	Met 255	Val	: .	л".	

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Trp Arg Thr Glu Gly Leu Leu Pro Leu Glu Glu Thr Ser Pro Arg Gln 260 265 270 Ala Val Cys Leu Thr Arg His Leu Thr Ala Phe Gly Ala Ser Leu Phe 275 280 285 Val Pro Pro Ser His Val Arg Phe Val Phe Pro Glu Pro Thr Ala Asp 290 295 300 Val Asn Tyr Ile Val Met Leu Thr Cys Ala Val Cys-Leu Val Thr Tyr. Met Val Met Ala Ala Ile Leu His Lys Leu Asp Gln Leu Asp Ala Ser 325 330 335 Arg Gly Arg Ala Ile Pro Phe Cys Gly Gln Arg Gly Arg Phe Lys Tyr 340 345 350 Glu Ile Leu Val Lys Thr Gly Trp Gly Arg Gly Ser Gly Thr Thr Ala His Val Gly Ile Met Leu Tyr Gly Val Asp Ser Arg Ser Gly His Arg 370 375 380 His Leu Asp Gly Asp Arg Ala Phe His Arg Asn Ser Leu Asp Ile Phe 385 390 395 400 Arg Ile Ala Thr Pro His Ser Leu Gly Ser Val Trp Lys Ile Arg Val 405 410 415 Trp His Asp Asn Lys Gly Leu Ser Pro Ala Trp Phe Leu Gln His Val 420 425 430 Ile Val Arg Asp Leu Gln Thr Ala Arg Ser Ala Phe Phe Leu Val Asn Asp Trp Leu Ser Val Glu Thr Glu Ala Asn Gly Gly Leu Val Glu Lys 450 455 460 Glu Val Leu Ala Ala Ser Asp Ala Ala Leu Leu Arg Phe Arg Arg Leu Leu Val Ala Glu Leu Gln Arg Gly Phe Phe Asp Lys His Ile Trp Leu 485 490 495 Ser Ile Trp Asp Arg Pro Pro Arg Ser Arg Phe Thr Arg Ile Gln Arg 500 505 Ala Thr Cys Cys Val Leu Leu Ile Cys Leu Phe Leu Gly Ala Asn Ala 515 520 525 Val Trp Tyr Gly Ala Val Gly Asp Ser Ala Tyr Ser Thr Gly His Val 530 535 540 Ser Arg Leu Ser Pro Leu Ser Val Asp Thr Val Ala Val Gly Leu Val 545 550 560 Ser Ser Val Val Val Tyr Pro Val Tyr Leu Ala Ile Leu Phe Leu Phe

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Arg	Met	Ser	Arg. 580	Ser	Lys	Val	Ala	Gly 585	Ser	Pro	Ser	Pro	Thr 590	Pro	Aļa
Gly	Gln	Gln 595		Leu S.	Asp.	Ile	Asp 600	Ser	Cys	Leu	Asp	Ser 60		Val	Leu
Asp	Ser 610	Ser	Phe	Leu	Thr	Phe 615	Ser	Gly	Leu	His	Ala 620	Glu	Ala	Phe	Val
Gly 625		Met	Lys	Ser	Asp 630	Leu	Phe	Leu.	ýsp	Asp 635	Ser	Lys	Ser	Leu	Val 640
Cys	Trp	Pro	Ser	Gly 645	Glu	Gly	Thr	Leu,	Ser 650	Trp.	Pro	Asp :	Leu	Leu 655	Ser
Asp	Pro	Ser.	Ile 660	Val	Gly	Ser	Asn	Leu 665	Arg	Gjv	Leu	Ala	Arg 670	Gly	Gln
Ala	Gly	His 675	Gly	Leu	Gly	Pro	Glu 680	Glu	Asp	Gly	Phe	Ser 685	Leu	Ala	Ser
Pro	Tyr 690	Ser	Pro	Ala	Lys	Ser 695	Phe	Ser,	Ala	Ser	Asp 700	Glu	<b>Asp</b>	Leu	Ile
Gln 705	Gln	Val	Leu	Ala	Glu 710	Gly.	yal.	Ser	Ser	Pro 715	Ala	Pro	Thr	ĞŢυ	Asp 720
Thr	His	Met	Glu	Thr 725	Asp	Leu <sub>i</sub>	Leu	Ser	Ser 730	Leu	Ser	Ser	Thr	Pro. 735	Gly
Glu	Lys	Live	Glu 740	Thx	Leu	Ala		Gln 745	Yid	Leu	Gly	<b>Gl</b> u	Leu 750	Gly,	Pro
Pro	Ser	Pro 755	Gly	Leu	Asn	طئل	Glu 760	Gln	Pro	Gln	Ala	Ala 765	Arg	Leu	Ser
Arg	Thr 770	Gly	Leu	Val	Glu	Gly 775	<u>Ļeu</u>	Arg	Lys	Arg	Leu 780	Leu	Pro	Ala	Trp .;
Cys 785	Ala	Ser	Leu	Ala	His 790	Gly (	Ļeu	Ser	Leu	Leu 795	Leu	Val.	Ala	Val	Ala 800
Val	Ala	Val	Ser	Gly 805	Trp	Val,	Gly	Ala.	Ser 810	Phe,	Pro	Pro,	Gly	Val 815	Ser
Val	Ala	ŢŢ	Leu 820	Leu	Ser	Ser	Ser	Ala 825	Ser	Phe	Leu	Ala	Ser 830	Phe	Leu
Gly	QIT	Glu 835	Pro	Leu	Lys	Val	Leu 840	Leu	Ģlu	Ala	Leu	Tyr 845	-	Ser	Leu
Val	Ala 850	Lys	Arg	<u>Leu</u>	Hīrē	Pro 855	Asp	Glu	Asp	Asp	Thr 860	Leu	Val	Glu	Ser
Pro 865	Ala <sub>.</sub>	Val	Thṛ	Pro	Val 870	Ser <sub>.</sub>	Ala	Arg	Val	Pro 875	ýrg	.Val	Arģ	Pro	Pro 880
His	Gly	Phe	Ala	Leu 885	Phe	Leu.	Ala	Ļys	Glu 890	Glu	Ala	Arg		Val	Lys.

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				•				_	12.7						
Arg	Leu	His	Gly 900	Met	Ĺeu	Arg	Ser	Leu 905	Leu	Val	ŢŸŢ	Met	Leu 910	Phe	Leu .
Leu		Thr 915	Leu	Leu	Ala	Ser	Tyr 920	Ģly	Asp	Ala `	Ser	Cys 925	His	Gly	His
Ala	Tyr 930	Arg	Leu	Gln	Ser	Ala 935	Ile	Lys	Gln	Glu -	Leu 940	His.	Ser	Arg	Ala
Phe 945	Leu	Ala	Ile	Thr	Arg 950	Ser	Glu	Glu	Leu	Trp 955	Pro	Trp	Met	Ala	His 960
Val	Leu	Leu	Pro	Tyr 965	Val	His	Gly.	ýżu	Gln 970	Ser	Ser	Pro	Glu.	Leu 975	Gly <sub>.</sub>
Pro	Pro	Arg	Leu 980	Arg	Glņ	Val	Arg	Leu 985		Glų	Ala	Leu	Ty <u>r</u> 990	Pro	Asp
Pro	Pro	Gly 995	Pro	Arg	Val	His	Thr 1000		Ser	Ala	Ala <sub>.</sub>	Gly 1005		Phe	Ser
Thr	Ser 1010		Tyr	Asp		Gly 101		Glu	Ser	Pro	His 1020	Asn )	Gly	Ser	Gly,
Thr 1025		Ala	Туг	Ser	Ala 1030		Asp	Ļeu	Leu.	Gly 1035		Trp	Ser	Trp	Gly 1040
Ser	Cys	Ala	Val	Tyr 104		Ser	Gly	Gly	Tyr 1050		Ģln	Glu	Leu	Gly. 1055	Leu
2 Ser	Leu	Glu	Glu 1060	Ser )	Arg	Asp	Arg	Leu 1065	Arg	Phe	Leu	Glņ	Leu 1070	His ).	Asn
Trp	Leu	Asp 107		٧ٺظ	Ser	Arg	Ala 1080		Phe	Leu	Gļu	Leu 1089	Thr 5	Arg	Tyr
Ser	Pro 1090		Val	Gly	Leu	His 109		Ala	Val	Thr	Leu 1100		Leu	Glu	Phe
Pro 1105		Ala	Gly	Arg	Ala 1110		Ala	Ala	Leu	Ser 111		Arg	Pro	Phe	Ala 1120
Leu	Arg	Arg	Leu	Ser 112		Gly	Leu	Ser	Leu 1130		Ļeu	Leu	Thr	Ser. 113	Val 5
Cys	Leu	Leu	Leu 114		Ala	Val	His	Phe 114		Val	Ala	Glu	Ala 1150		Thr
Trp	His	Arg 115		Gly	Arg	Trp	Arg 116		Leu	Arg	Leu	Gly 116	Ala 5	Lib	Ala
	Trp 117		Leu	Val	Ala	Leu 117		Ala	Ala	Thr	Ala 118		Val	Arg	Leu
8 Ala 118		Leu	Gly	Ala	Ala 119		Arg	Gln	Trp	Thr 119		Phe	Val	Arg	Gly 1200
Arg 2	Pro	Arg	Arg		Thr :05	Ser	Phe	Asp		Val 10	Ala	His	Val	Ser 12	Ser 15

Ala Ala Arg Gly Leu Ala Ala Ser Leu Leu Phe Leu Leu Val Lys 1220 1225 1230

Ala Ala Gln His Val Arg Phe Val Arg Gln Trp Ser Val Phe Gly Lys 1235 1240 1245

Thr Leu Cys Arg Ala Leu Pro Glu Leu Gly Val Thr Leu Gly Leu 1250 1260

Val Val Leu Gly Val Ala Tyr Ala Gln Leu Ala Ile Leu Leu Val Ser 1265 1270 1275 1280

Ser Cys Val Asp Ser Leu Trp Ser Val Ala Gin Ala Leu Leu Val Leu 1285 1290 1295

Cys Pro Gly Thr Gly Leu Ser Thr Leu Cys Pro Ala Glu Ser Trp His 1300 1305 1310

Leu Ser Pro Leu Leu Cys. Val Gly Leu Trp' Ala Leu Arg Leu Trp Gly 1315 1320 1325

Ala Leu Arg Leu Gly Ala Val Ile Leu Arg Trp Arg Tyr His Ala Leu 1330 1340

Arg Gly Glu Leu Tyr Arg Pro Ala Trp Glu Pro Gln Asp Tyr Glu Met
1345 1350 1355 1360

Val Glu Leu Phe Leu Arg Arg Leu Arg Leu Trp Met Gly Leu Ser Lys 1365 1370 1375

Val Lys Glu Phe Arg His Lys Val Arg Phe Glu Gly Met Glu Pro Leu 1380 1385 1390

Pro Ser Arg Ser Ser Arg Gly Ser Lys Val Ser Pro Asp Val Pro Pro 1395 1400 1405

Pro Ser Ala Gly Ser Asp Ala Ser His Pro Ser Thr Ser Ser Ser Gln. 1410 1420

Leu Asp Gly Leu Ser Val Ser Leu Gly Arg Leu Gly Thr Arg Cys Glu 1425 1430 1435 1440

Pro Glu Pro Ser Arg Leu Gln Ala Val Phe Glu Ala Leu Leu Thr Gln 1445 1450 1455

Phe Asp Arg Leu Asn Gln Ala Thr Glu Asp Val Tyr Gln Leu Glu Gln 1460 1465 1470

Gln Leu His Ser Leu Gln Gly Arg Arg Ser Ser Arg Ala Pro Ala Gly 1475 1480 1485

Ser Ser Arg Gly Pro Ser Pro Gly Leu Arg Pro Ala Leu Pro Ser Arg 1490 1495 1500

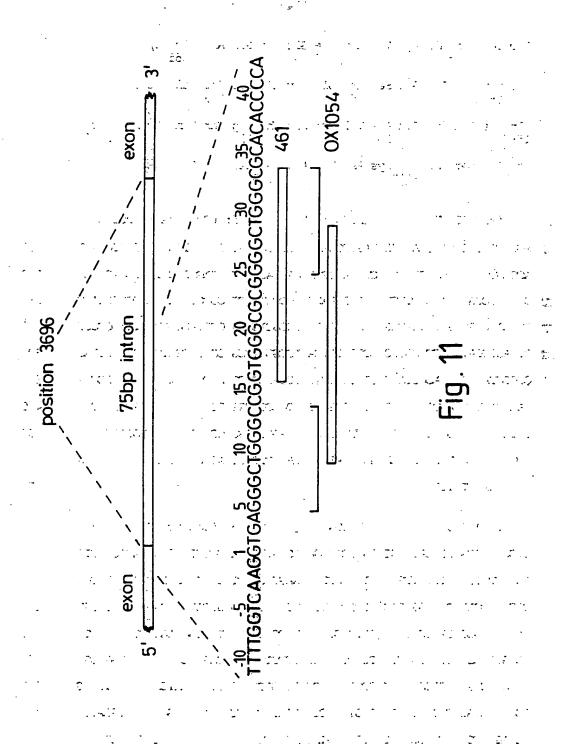
Leu Ala Arg Ala Ser Arg Gly Val Asp Leu Ala Thr Gly Pro Ser Arg 1505 1510 1520

Thr Pro Ser Gly Gln Glu Gln Gly Pro Pro Gln Gln His Leu Val Leu 1525 1530 1535

Leu Pro Gly Gly Gly Pro Trp Ser Arg Ser Gly His Arg Ser Val

1540 Leu Leu Ser Ala Ala Val Lys Ala Glu Gly Gln Ala Glu Trp Leu His 1560 Val Gly Ser Pro Glu Ser Arg Gln Gly His Leu Ser Val Cys Gly Leu 1580 1575 Gln His Phe Lys Glu Ala Val Trp Pro Thr Arg Thr Gln Gly Pro Leu 1590 1595 1585 Pro Ser Ser Leu Gly Lys Asp Thr Ala-Val Leu Asp Gly Phe 1605 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5: (Compare Figure 8) ACCITICACAC CATCAAGGC CAGITICAACT TIGIOCACGI GATCGICACC COGCIGGACT 60 ACCACTOCAA CCTOCTOTOC CTGCACTOCA GGAAAGACAT GGAGGGCCTT GTGGACACCA 120 GOGTGGCCAA GATOGTGTCT GACCGCAACC TGCCCTTCGT GGCCCGCAG ATGGCCCTGC 180 ACGCAAATAT GGOCTCACAG GTGCATCATA GCCGCTCCAA CCCCACCGAT ATCTACCCCT 240 CCAAGIGGAT TGCCCGCCIC CGCCACATCA AGCCGCTCCG CCAGCGGATC TGCGAGGAAG 300 COSCUTACTIC CAACCOCAGE CTACCTCTGG TGCACCCTCC GTCCCATAGC, AAAGCCCCTG :360 CACAGACTOC AGOOGAGOOC ACACCTGGCT ATGAGGTGGG OCAGGGGAAG OGCCTCATCT 420 480 OCTOGGTGGA GGACTICACC GAGTITGTGT GAGGCCGGGG COCTCCCTCC TGCACTGGCC 540 TIGGACOGTA TIGOCIGICA GIGAAATAAA TAAAGIOEIG ACCCCAGIGC ACAGACATAG 553 AGGCACAGAT TGC

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6: (Compare Figure 9) 60 CTGGTGTGTG TGAGACGTGC GGGCCTGGGA AGTGTTGGCA GAGCCGGGGG TACCGTCCTC ACTOCITITIG TICTITIGAC GIAAGCIGGC GAGIGGCACT GOCIGAGTIC CGCTCAGIGC 120 COSCOURGAT GROOGACCE COCROCATIC TROCKGRIAG GROGROGOGG TGROOGCIGT 180 OCCIOGIOGG CACOGAGAGI CITITOGGAGC TITIGGGGAGG TITGIGOCAAG OCTGAGOCIC 240 GACGICCCCC TICCCGGCTT TCTGTTGGCT CTTCTGAGGC CAGGGCATCT CTATGAGGCC 300 CTCCTGCTGG AGCCGTCTCT GTGGATCTCC TCTGCCATCC TGGCCCATGA GTGGGTGATG 360 CECTGECCAC CATCTGGTGA CAGTGGCCGG GCACCGCTGC CAAATGTGGG TCCCGCATCT 420 GCAAGCCCCT COCTGGGTCC CCTAGGGTAT GGGGTGGTTC TGCCACTGCC CTCGCTCCCC 480 CACCITGGGG TGCCTCTCCC CCTGCTCGTG GGGGAGA 517



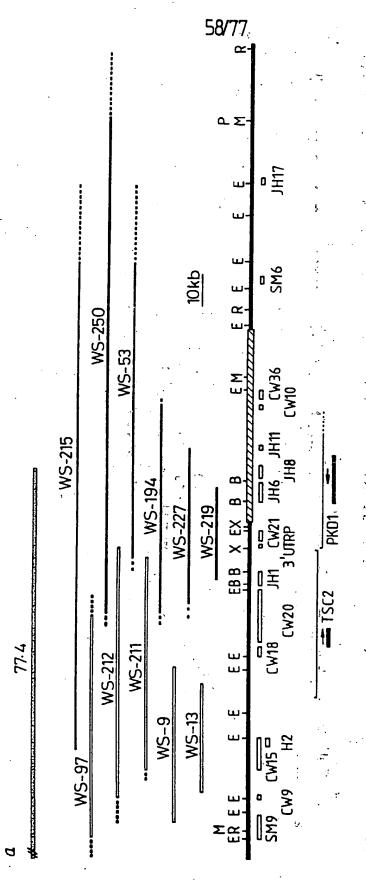
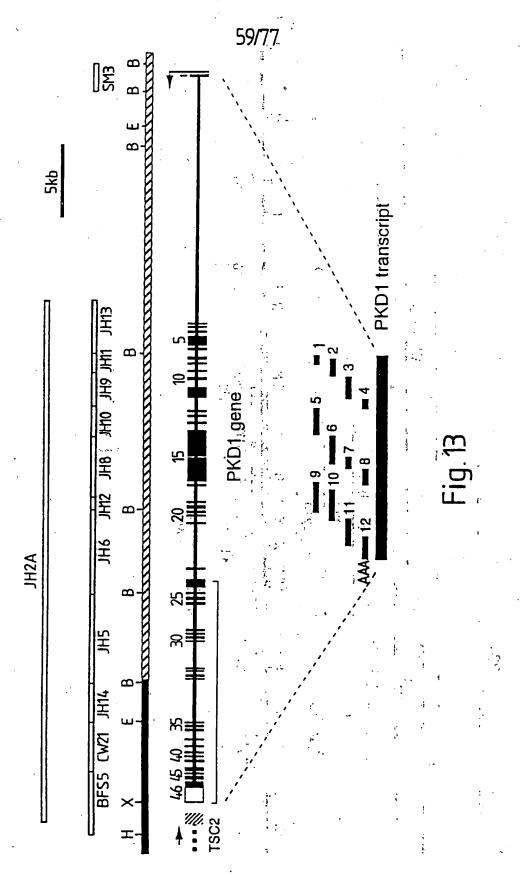
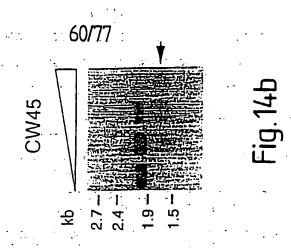


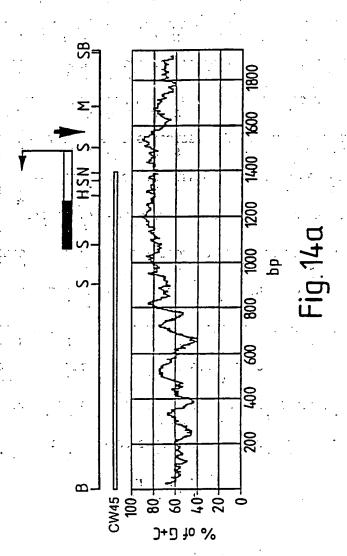
Fig. 12

SUBSTITUTE SHEET (RULE 26)



SUBSTITUTE SHEET (RULE 26)





**SUBSTITUTE SHEET (RULE 26)** 

1	GCACTGCAGCGCCAGCGTCCGAGCGGGCGGCCGAGCTCCCGGAGCGGCCTGGCCCCGAGC	60
61	CCCGAGCGGGCGTCGCTCAGCAGCAGGTCGCGGCGCGCAGCCCCATCCAGCCCCGCGCC	120
121	CGCCATGCCGTCCGCGGGCCCCGCCTGAGCTGCGGTCTCCGCGCGGGCGG	180
181 1	ACGGCGGGCCATGCGCTGCCCTAAGGATGCCGCCGCCGCCGCCCGC	240 10
241 11	GCTGGCCCTGGGCCTGTGGCTCGGGGGGGCGCGGGCGCGGCTG L A L G L G L W-L G A L A G G P G R G C	300 30
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301 31	CGGGCCCTGCGAGCCCCCTGCCTCAA G P C E P P C L C G P A P G A A C R V N *	360 50
361 51	CTGCTCGGGCCGCGGGCGGACGCCAC C S G R G L R T L G P A L R I P A D A T	420 70
		_
421 71	AGCGCTAGACGTCTCCCACAACCTGCTCCGGGCGCTGGACGTTGGGCTCCTGGCGAACCT A L D V S H N L L R A L D V G L L A N L *	480 90
481	CTCGGCGCTGGCAGAGCTGGATATAAGCAACAACAAGATTTCTACGTTAGAAGAAGGAAT	540
91	SALAELDISNNKISTLEEGI	110
541	ATTTGCTAATTTAATTTAAGTGAAATAAACCTGAGTGGGAACCCGTTTGAGTGTGA	600
. 111	FANLFNLSEINLSGNPFECD	130
601	CTGTGGCCTGGCGGCGCGATGGGCGGAGGAGCAGCAGGTGCGGGTGCAGCC	660
131	CGLAWLPRWAEEQQVRVVQP	150
661	CGAGGCAGCCACGTGTGCTGGGCTGCCTGGCTGGCCAGCCTCTGCTTGGCATCCC	720
.151	E A A T C A G P G S L A G Q P L L G I P	170
721	CTTGCTGGACAGTGGCTGTGGTGAGGAGTATGTCGCCTGCCT	780
171	LLDSGCGEEYVACLPDNSSG	190
781	CACCGTGGCAGCAGTGTCCTTTTCAGCTGCCCACGAAGGCCTGCTTCAGCCAGAGGCCTG	840
191	TVAAVSFSAAHEGLLQPEAC	210
841	CAGCGCCTTCTGCTTCTCCACCGGCCAGGGCCTCGCAGCCCTCTCGGAGCAGGGCTGGTG	900
211	SAFCFSTGQGLAALSEQGWC	230
901	CCTGTGTGGGGCGGCCCAGCCCTCCAGTGCCTCCTTTGCCTGCC	960
231	L C G A A Q .P. S S A S F A C L S L C S G	250
961	CCCCCGCCACCTCCTGCCCCCACCTGTAGGGGCCCCACCCTCCTCCAGCACGTCTTCCC	1020
251	PPPPPAPTCRGPTLLQHVFP	270
1021	TGCCTCCCAGGGGCCACCCTGGTGGGGCCCCACGGACCTCTGGCCTCTGGCCAGCTAGC	1080
271	ASPGATLVGPHGPLASGQLA	290
1081	AGCCTTCCACATCGCTGCCCCGCTCCCTGTCACTGCCACACGCTGGGACTTCGGAGACGG	1140
291	A F H I A A P L P V T A T R W D F G D G	310
1141	CTCCGCCGAGGTGGATGCCGCTGGGCCGGCTGCGTCGCATCGCTATGTGCTGCCTGGGCG	1200
311	S A E V D A A G P A A S H R Y V L P G R	330

Fig. 15

1201 331	CTATCA(																1260 350
1261 -351	GCAGGTO Q V																1320 370
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1681 491	GAGCTGC	CAGA	\CTG0	CTGC	CCGG	GGA	GCCA	CACCO	AGC	CACA	ÄĞCO	GAG	CAC	TG	CGT	CCG	1740
1741	GCTCGGG	CCCAC	CGGG	TGGT	GTAA	.CAC	CGAC	CTGT	CTC	AGCC	ccc	CAC	AGC	TÀC	CGTO	CTG	1800
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i801	CGAGCTG	CAGCO	CGGA	.GGCC	CAGT	GCA	GGAT		GAA	CCTC	CTC	GTG	IGGA	.GC	SCC	CAG	530 1860
531	CGAGCTG E L	CAGCO Q P	CCGGA G	G P	CAGT	GCA( Q	GGAT D	GCCGA A E	GAA( N	CCTC	CTC	GTC V	GG2 G	A	CCC P	AG S	1860 550
531 1861 551	CGAGCTO E L TGGGGAC G D	CAGCO Q P CCTGCA L Q	CGGA G AGGGA G	G P	CAGT V TGAC	GCA( Q GCC' P	GGAT D ICTG L	GCCGA A E GCACA A Q	GAA N .GCA Q	CCTC L GGAC D	CCTC L CGGC	GTG V CTG	IGGA G TCA S	GC( A GC( A	P P CCCC	CAG S GCA H	1860 550 1920 570
531	CGAGCTO E L TGGGGAC G D CGAGCCO	CAGCO Q P CCTGCA L Q	CCGGA G AGGGA G	AGGCC G P ACCCCC P L	CAGT V TGAC	GCA( Q GCC' P	GGAT D ICTG L GGGC	GCCGA A E GCACA A Q	GAA N .GCA Q TCT	CCTC L GGAC D	CCTC L CGGC G	GTO V CTO L	IGGA G TCA S	A A A A CTT(	CCTO	CAG S SCA H	1860 550 1920
531 1861 551 1921	CGAGCTO E L TGGGGAC G D CGAGCCO	GCAGCO Q P CCTGCA L Q CGTGGA V E	CCGGA G AGGGA AGGTC V	AGGCC G P ACCCC P L CATGG M V	CAGT TGAC TATT F	GCA( Q GCC) P CCC( P	GGAT D ICTG L GGGC G	GCCGA A E GCACA A Q CTGCG L R	GAA N GCA Q TCT L	CCTC L GGAC D GAGC S	CCTC L CGGC G CCGT R	GTC V CTC L CGAA	GGA G TCA S GCC A	GCCA	CCCC P CCTC L	CAG S SCA H CAC T	1860 550 1920 570
531 1861 551 1921 571 1981	CGAGCTO E L TGGGGAC G D CGAGCCO E P	CAGCC Q P CTGCA L Q CGTGGA V E CGAATT E F	CCGGA G AGGGA V TGGG G	AGGCCC PLATGG MV TAGACCC	TGAC TATT FAGGA	GCAC P CCCC P GCTC	GGAT  ICTG  L  GGGC  G  CCGG  R  GGAG	GCCGA A E GCACA A Q CTGCG L R CGGCC R P	GAA N GCA Q TCT L CGC A	GGAC  CCAC  CCAC	CCTC L CGGC G CCGT R CCTC	CGTC L CGAA E CGG	GGA GCC A GCC A GCTC L	AGCO ACTTO F	CCTC P CCTC L GGTC V	CAC H CAC T TSTA Y	1860 550 1920 570 1980 590
531 1861 551 1921 571 1981 591 2041	CGAGCTO E L  TGGGGAC G D  CGAGCCO E P  CACGGCO T A	GCAGCO Q P CCTGCA L Q CGAATI E F CCTCAC L S CAGGAC R T	CCCAC	AGGCCC PL CATGG MV CACCCC T Q AGCAG A G	TGACT T F AGGACT T	GCAC  P  GCTC  L  CCCCC  P	GGAT D TCTG L GGGC G CCGG R GGAG E	GCCGA A E GCACA A Q CTGCG L R CGGCC R P AACGG	GAAN  GCA  Q  TTCTC  L  CCGCC  A  CAGGG  S  A  CAGGG  G	GGAG CGAG EGGAG	CCCTC L CCGGC G CCCGT R CCCGT R L L ACGC	CGTG  CGTG  CGAA  CGAA	GGGA G TCA S AGCC A GCTC L AGCC S	AGCCAA TTTC F GCAC R AGCCR	P CCTC L CGGTC V STCC	CAC TCAC TCAC TCAC TCAC TAC TAC TAC TAC	1860 550 1920 570 1980 590 2040 610
531 1861 551 1921 571 1981 591 2041 611 2101 631	CGAGCTO E L  TGGGGAC G D  CGAGCCO E P  CACGGCTO R L  GGACAAC D **  CAACATO N I	GCAGCO Q P CCTGCA L Q CGAATI E F CCTCAC L S CAGGAC R T	CCCAC	AGGCCO P L CATGG M V CACCC T Q AGCAG A G GCTGG L A GCTGG L D	TGAC TATT F AGGAC TCCCC ACCCC	GCAC P CCCC P GCTC A CCCCC S	GGAT D TCTG L GGGGC G CCGG R GGAG E GGAG C C C C C C C C C C C C C C C C C	GCCGA A E GCACA A Q CTGCG L R CGGCC R P AACGG M P ATGCC H P	GAAG N GCAG Q TCTC L CGCC A CAGG S CAGG G	CCTC L GGAC D CCAC S CCAC Q CGAC E GGGGA G GGCC A	CCCTC L CCGGC R R GCCTC L ACGCC R	CGTC  CGAA  CGAA	GGGA GCCC A GCCC A GCCC C S GCCC C C C	AGCO A TTTO F GCAO R GCCO R GCCO F GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	P CCCTC P CCCTC V S CCCTC S CCCTC CC	CAC T CCC P AGC A	1860 550 1920 570 1980 590 2040 610 2100 630 2160 650
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3001 931 3061 951 3121 971 3181 991 3241 1011	GCC P AGT V CGA D GGT V CGT V	CATO I GAGO R CAAO K CTTO F AACO T	CTG' C GTAC Y CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	TGGG G CAGG S GTCG S CTG L	CCTCL P CCCTCL L CTCL S GCGCR	CCGC R CCGTC V CGACCC T CACCCC M	CGCCAA GGTTC V CTTC F GACC T GAAC	CACC T EGGA E ECCAC Q GGC A EAGC R EGGT	GCCC P GGCC A N CTCC S GATC	CAG S CGG C CGT V CAA N CGA Q	* CCCCAC  BGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	EGAC EGGAC D CTTC F CGTC V	GGCI A CATY M CAA' N SAGGCAG S	CCGT R GGT( V PGT( V ** GGT( V	V CTTC F CATT I CCGTC V CTCC S	ACTO L CCGGG R TTAT Y CACC T CACC T GGAC	GCA( Q GTG( W CCGT( Q AGTG V	GGGA G T GAGG S S S S S S S S S N C C C C C C C C C C	AGTO V CATO I CGCO A CTAC Y CGCO A	CCTOL LCAA N SGC A CAA N CGT V		3060 950 3120 970 3180 990 3240 1010 3300 1030	لنند
3001 931 3061 951 3121 971 3181 991 3241 1011	GCC P AGT V CGA D GGT V CGT V	CATO I GAGO R CAAO K CTTO F AACO T	CTG' C GTAC Y GGCAC Q C C C C C C C C C C C C C C C C C	TGGG G S S GTCC S GCTC L L	CCTCL P CCCTCL L CTCL S GCGCR	CCGC R CCGTC V CGACCC T CACCCC M	CGCCAA GGTCC V CTTC F. GACCCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	CACC T GGA( E CCAC Q GGCC A CAGG R GGTI V	GCCC P GGCC A N CTCC S GATC M	CAG S CGG G CGT V CAA N GCA Q	* CCCC P CTCC S GGTC V CCCAC H GGGG G	EGAC EGGAC DCTTC F CCGTC V	GGCI A CATY M CAA' N SAGGCAG S	CCGT R GGT( V PGT( V ** GGT( V	V CTTC F CATT I CCGTC V CTCC S	ACTO L CCGGG R TTAT Y CACC T CACC T GGAC	GCA( Q GTG( W CCGT( Q AGTG V	GGGA G T GAGG S S S S S S S S S N C C C C C C C C C C	AGTO V CATO I CGCO A CTAC Y CGCO A	CCTOL LCAA N SGC A CAA N CGT V		3060 950 3120 970 3180 990 3240 1010 3300 1030	لنند
3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031	GCC P AGT V CGA D GGT V CGT. V	CATO	CTG' C GTAC Y C GCAC Q C C C C C C C C C C C C C C C C C	TGGG G S S GGTC S GGAC E CAA:	CCTC L CCCCTC L CCTC S CCCCC R R R R CCCCC A	CCGC R CGTC V GGACC T ACTC M M CACC	CGCCAA GGTCV CTTC F GAACC N ACTC	CACO T GGAO E CCAO Q GGGCC A CAGO R	GCCC P GGCC A S GGAA N CTCC S M M	CAG S CGG G CCAA N CCAA Q CCAC Q	# CCCC P CTCC S GGTC V CCAC H CGGGG G	EGAC EGGAC DCTTC F CCGTC V ICTC C	GGCCAAN M CAAN N SAGCAC V V	CCGC R GGTC V CAAC N *GGTC V	V CTTC F CATT I CGTC V CTCC S	L CCCGG R R TATA Y TACK T T T T T T T T T T T T T T T T T T T	GCAC Q W CCGTCAC V AGTC V	GGGGA G G G G G G G G G G G G G G G G G	AGTO V I CGCO A CTAC Y	CCT L CAA N CGC A	A.S.	3060 950 3120 970 3180 990 3240 1010 3300 1030 3360 1050	
3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361	GCC P AGT V CGA D GGT V CGT L GGT	CATO	CTG' C GTAC Y GGCAC Q CAAC K CCGTC V CCCCC P	TGGG G CAGG S SCTC L CAAC N *	CCTC L CCCCC P CCCTC L CTCCC R CCCCC R CCCCC A	CCGC R CGTC V GACTC T ACTC M CACTC	CGCCAA  GGTC V CTTC F GACCC N ACTC	CACO T GGAO E CCAO Q GGCCO A CAGO R V	GCCC P GGCC A GGAAC N CTCC S M M ACTC	CAG S CGG G CCAA CCAA Q CCAA T T	* CCCC P CTCC S GGTC V CCCAC H CGGGG G	EGA(  E  CGA(  D  CTT(  F  CGT(  V  TCT(  G  G  GCA(  GCA(  CGA(  CGA(	GGCCAA'  M CAA' N SAGCAC V V GGCCC	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	V CCAC	CCACO	GCAC Q W CCGTCAC V AGTC V	GGGA G G G G G G G G G G G G G G G G G	AGTO V I CCGCO A CTAC Y	CCT L CAA N CGC A CAA N CGT V	A.S.	3060 950 3120 970 3180 990 3240 1010 3300 1030 3360 1050	
3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361	GCC P AGT V CGA D GGT V CGT. V	CATO	CTG' C GTAC Y GGCAC Q CAAC K CCGTC V CCCCC P	TGGG G CAGG S SCTC L CAAC N *	CCTC L CCCCC P CCCTC L CTCCC R CCCCC R CCCCC A	CCGC R CGTC V GACTC T ACTC M CACTC	CGCCAACTCCCCCCAACTCCCCCCCCCCCCCCCCCCCCC	CACO T GGAO E CCAO Q GGCO R CAG V V TGGO G	GGCC P GGCC A GGAA M M ACTC L GGAA D	CAG S CGG G CGT V CAA O CGCA O GAC G	* CCCC P CTCC S GGTC V CCAC H GGGG G GGGAC E	EGA(  E  CGA(  D  CTT(  F  CGT(  V  TCT(  L  G  G  G  G  G  G  G  G  G  G  G  G	GGCCAA'  M CAA' N SAGCAC V V GGCCC	CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	V CCAC	CCACO	GCAC Q W CCGTCAC V AGTC V	GGGA G G G G G G G G G G G G G G G G G	AGTO V I CCGCO A CTAC Y	CCT L CAA N CGC A CAA N CGT V	A.S.	3060 950 3120 970 3180 990 3240 1010 3300 1030 3360 1050	
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3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361 1051 3421	GCC P AGT V CGA D GGT V GGT V GGT L GGT V GTA	CATU	CTG' CTG' Y GCAC K CCGTC CCCC P CCCCC F CCGAC E	TGGG G S S S S S C T C C S S S C T C C S S S C T C C S S C T C C S C T C C S C T C C S C T C C S C T C C C C T C C C T C	CCTC L CCCCC P CCTCL S GCGCGC R GGCGC W CCTTC	CCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CGCCAA  GGTC V  CTTC F.  GAAC N  ACTC F.  CTTC ACTC GGTC GGTC GGTC GGCT GGCT GGC	CACO T GGA E CCA Q GGC A CAG C GGC C	GCCC P GGCC A GAA N CTCC S GAA M ACTC L GGA D	CAG G G CGT V CAA N G G G G G G G G G G G G G G G G G G	*CCCC P CTCC S GGTC CCAC H CGGG G GGGC CTCC CTCC CTCC CTCC CTCC	CGA(CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	GGCCAACCAACCAACCAACCAACCAACCAACCAACCAAC	CCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	V CTTC F CATT I CGTC V CTCC S GGTC V CCAC H	LCCCGC R TTAT Y LACO T CGGAC CCAC Q GCTC	GCAC  Q  CCAC  Q  CGTC  V  CTCF  SGTC  F	GGGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	AGTO V CATO I CGCO A TAC Y GGCO A CGCO A CGCO A CGCO A CGCO A CGCO CGCO	CCTD. LCAA N CGC A CGA CGA CGA CGA CGA CGA CGA CGC CCC CC		3060 950 3120 970 3180 990 3240 1010 3300 1030 3420 1070	
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3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361 1051 3421 1071	GCC P AGT V CGA D GGT V GGT L GGT Y TGT	CATU  GAGG  R  CAAG  K  CTTO  F  GTCO  S  GGCO  N  * CAAG	CTG' C GTA( Y GGCA( K CCGTC V CCCCC F CCGA( E GGA( E G E GGA( E GGA( E G E G E G E G E G E G E G E G E G E	TGGG G S S S S S S S S S S S S S S S S S	CCTC P CCCTC P CCTC S GCGG R GGGG W CTTC F CTTAC	CCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CGCCAACTCCACCACCACCACCACCACCACCACCACCACCAC	CACCACACACACACACACACACACACACACACACACAC	GCCC GGCCC A A BAAC M CTCCC S GGATC D AGAC D	CAGG CGGC CGT V CAAG CGCG GGCCCC P	* CCCC P CTCCC S GGTC CCAC H GGGG CCAC CCCAC CCCCAC CCCAC CCCAC CCCAC CCCAC CCCAC CCCAC CCCAC CCCAC CCCAC CCCCAC CCCCAC CCCCAC CCCCAC CCCCCAC CCCCAC CCCCCC	CGA( CTTC CGTC V TCTC CGTC CGTC CGTC CGTC CG	CATOM CAAO CAAO CAAO CAAO CAAO CAAO CAAO CA	CCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CONTROL OF THE CONTRO	CCCCC R PTATI Y CACCA T CCACC CCACC CCACC CCACCC CCACC CCACCC CCACC CCACCC CCACC CCACCC CCACCC CCACCC CCACCC CCACC CCACC CCACC CCACCC CCACC CCACC CCACC CCACC CCACC CCACC	GCAC Q GTGC W CCAC Q CGTC V TCF S GTTC V GGC	GGGGAGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	AGTO V CATO I CGCO A CG	CCTOL  CAA  N  CGC  CGA  CGA  CGA  CGA  CGA		3060 950 3120 970 3180 990 3240 1010 3300 1050 3420 1070 3480 1090	
3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361 1051 3421 1071 3481 1091	GCC P AGT V CGA D GGT V CGT L GGTA Y TGT V	CATO I GAGO R CAAO K CTTO F AACO T GTCO S GGCO A CAAO M * CATO	CTG'C GTAG Q CAAG K CCGTC V CCCCC P CCCCC P CCGAG E CCGAG H	TGGG G CAGG S GTCG S GCTC CGGAG  ** CCTC L GTCG CTC CAGG CTC CTC CTC CTC CTC CTC CTC CTC CTC C	CCTCL CCCCC P CCTCL CTCL S GCGC R GCGC A GTGC Y	CCGGCCAA	CGCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAGGCCCAAAAGGCCCAAAAGGCCCAAAAGGCCCAAAAGGCCCAAAAGGCCCAAAAGGCCCAAAAGGCCCAAAAAGGCCCAAAAAA	CACC T GGAC Q CCAC Q GGCC A CAGC CAGC CCC P	GCCC P GGCC A GAAA N CTCC S GATC D ACTC D ACTC D	CAGG SECON CAAN CAAN GCAN GCAN GCCO GCCO P	* CCCC P CTCC S GGTC V CCCAC H GGGG G GGGC CTCCC S GGAC E C C C C C C C C C C C C C C C C C C	EGA( E EGA( D ETT( F EGT( V EGT( L EGG( G G G V EGT( V EGG( C C E E E E E E E E E E E E E E E E E	CATC M CAAC N CAAC N S S GGCAC V V GGCC A	CCGCCR  GGTC  V  CAACC  N  *GGTC  CCCTC  CCCACC  Q  GGACC  T	CCAC H GGTC V CCAC CCAC V CCAC V CCAC V CCGTC V	CCGCC R TTAT Y CACA T CACA CCAC CCAC CCAC CCAC	GCAC Q GTGC Q CGTC V AGTC S GTGC V GGC A GGC A	GCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGTO V CATO I CGCO A TAO CGCO V CCCT P CCAO H	CCTOL LCAA N CGC A CGA CGA CGA CGA CGA CGA CGA CGC CGA CGA		3060 950 3120 970 3180 990 3240 1010 3300 1030 3420 1050 3420 1070 3480 1090	
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3001 931 3061 951 3121 971 3181 991 3241 1011 3301 1031 3361 1051 3421 1071 3481 1091	GCC P AGT V CGA D GGT V GCTA V GTA Y CTTC	CATU GAGGE R CAAC K CTTC F AACC T GTCC S GGCC A CAAC M ** CCATC	CTG'C GTAC Y GCAC CCCC CCCCC P CCCCC F CCGAC E GCAC H GGAAC	TGGG CAGG S STCO S SCTO L CAGG CAGG CAGG CAGG CAGG CAGG CAGG CA	CCTCL CCCCC P CCTCL CTCL S GCGC R GCGC A GTGC Y CTTCL Y GACC	CCGCCCCCCCCCCA	CGCCAACTCCCCAACTCCCCCAACCCCCCCCCCCCCCCC	CACCATOR TO THE PROPERTY OF TH	GCCC P GGCC A SAA N CTCC S SATC M ACTC D AGA D	CAG SECON CAAN CAAN CAAN CCCO CCCO PCCCO PCCCO PCCCO	* CCCC P CTCC S GGT V CCAC H GGGG G GGGC CTCC S GGAC E CTCC S GGAC E GGAC GGAC	CGA( E  GGA(  D  CTTC  F  CGTC  V  CGTC	GGCCCAACCCCCAACCCCCCAACCCCCCCCCCCCCCCC	CCGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CACO CACO CACO CACO CACO CACO CACO CACO	CCACO	GCAC  CCAC  CCAC	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	AGTO V CATO I CGCO A CTAO Y CGCO A CGCO H CGCO H CGCO CA CGCO CGCO CGCO CGCO CGCO CGCO C	CCT) CAA N CGC A CAA CCA CCC P CAA N CGC A CGC CGC CGC CGC CGC CGC CGC CGC C		3060 950 3120 970 3180 990 3240 1010 3300 1030 3420 1050 3420 1070 3480 1090	

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3601 1131	TGTGGGTGTGACTGACGGCCTCCTGGTGGCCGGCCCGCCC	3660 1150
3661 1151	GCTGCCCTCGCCTGGGGGTGTTCTTTACACGTGGGACTTCGGGGACGGCTCCCCTGTCCT L P S P G G V L Y T W D F G D G S P V L	.3720 1170
3721 1171	GACCCAGAGCCAGCCGGCTGCCAACCACCTATGCCTCGAGGGGCACCTACCACGTGCG T Q S Q P A A N H T Y A S R G T Y H V R	3780 1190
3781 1191	CCTGGAGGTCAACAACACGGTGAGCGGTGCGGGGGGCCCAGGCGGATGTGCGCGTCTTTGA L E V N T V S G A A A Q A D V R V F E	.3840 1210
3841 1211	GGAGCTCCGCGGACTCAGCGTGGACATGAGCCTGGCCGTGGAGCAGGGCGCCCCGTGGT E L R G L S V D M S L A V E Q G A P V V	3900 1230
3901 1231	GGTCAGCGCCGCGGTGCAGACGGCGACGGCGACGGCCTCGACATGGGGGACGGCCCCCGCGCACGGCCGACGGCCGACGGCCGACGGCGACGGCCGACGG	3960 1250
3961 1251	CACCGTGCTGTCGGGCCCGGAGGCAACAGTGGAGCATGTGTGTG	4020 1270
4021 1271	CACAGTGACCGTGGGCGGCCAGCCCGGCCACCTGGCCCGGAGCCTGCACGTGCT T V T V G A A S P A G H L A R S L H V L	4080 1290
4081 1291	GGTCTTCGTCCTGGAGGTGCTGCGCGTTGAACCCGCCGCCTGCATCCCCACGCAGCCTGA V F V L E V L R V E P A A C I P T Q P D	4140 1310
4141 1311	CGCGCGGCTCACGGCCTACGTCACCGGGAACCCGGCCCACTACCTCTTCGACTGGACCTT A R L T A Y V T G N P A H Y L F D W T F	4200 1330
4201 1331	CGGGGATGGCTCCTCCAACACGACCGTGCGGGGGTGCCCGACGGTGACACAACTTCAC G D G S S N T T V R G C P T V T H N F T	4260 1350
4261 1351	GCGGAGCGCACGTTCCCCCTGGCGCTGGTGCTGTCCAGCCGCGTGAACAGGGCGCATTA R S G T F P L A L V L S S R V N R A H Y	4320 1370
4321 1371	CTTCACCAGCATCTGCGTGGAGCCAGAGGGCA F T S I C V E P E V G N V T L Q P E R Q	4380 1390
4381 1391	GTTTGTGCAGCTCGGGGACGAGGCCTGGCTGGCATGTGCCTGGCCCCCGTTCCCCTA F V Q L G D E A W L V A C A W P P F P Y	4440 1410
444 <u>1</u> 1411	CCGCTACACCTGGGACTTTGGCACCGAGGAAGCCGCCCCACCCGTGCCAGGGGCCCTGA R Y T W D F G T E E A A P T R A R G P E	4500 1430
4501 1431	GGTGACGTTCATCTACCGAGACCCAGGCTCCTATCTTGTGACAGTCACCGCGTCCAACAA V T F I Y R D P G S Y L V T V T A S N N	4560 1450
4561 1451	CATCTCTGCTGCCAATGACTCAGCCCTGGTGGAGGTGCAGGAGCCCGTGCTGGTCACCAG I S A A N D S A L V E V Q E P V L V T S	4620 1470
4621 1471	CATCAAGGTCAATGGCTCCCTTGGGCTGGAGCTGCAGCAGCCGTACCTGTTCTCTGCTGT I K V N G S L G L E L Q Q P Y L F S A V	4680 1490
4681 1491	GGGCCGTGGGCGCCCGCCAGCTACCTGTGGGATCTGGGGGACGGTGGGTG	4740 1510
4741 1511	TCCGGAGGTCACCCACGCTTACAACAGCACAGGTGACTTCACCGTTAGGGTGGCCGGCTG PEVTHAYNSTGDFTVRVAGW	4800 1530

4801 GAATGAGGTGAGCCGCAGCGAGGCCTGGCTCAATGTGACGGTGAAGCGGCGCGTGCGGGG 4860 NEVSRSEAWLNVTVKRRVRG 1550 - .\* 4861 GCTCGTCAATGCAÁGCCGCACGTGGTGCCCCTGAATGGGAGCGTGAGCTTCAGCAC 4920 1551 L V V N A S R T V V P L N G S V S F S T 1570 4921 GTCGCTGGAGGCCGCAGTGATGTGCGCTÄTTCCTGGGTGCTCTGTGACCGCTGCACGCC 4980 SLEAGSDVRYSWVLCDRCTP 4981 CATCCCTGGGGGTCCTACCATCTCTTACACCTTCCGCTCCGTGGGCACCTTCAATATCAT 5040 I P G G P T I S Y T F R S V G T F N I I 1610 5041 CGTCACGGCTGAGAACGAGGTGGGCTCCGCCCAGGACAGCATCTTCGTCTATGTCCTGCA 5100 V T A E N E V G S A Q D S I F V Y V L Q 1630 5160 LIEGLQVVGGGRYFPTNHTV 1650 5161 ACAGCTGCAGGCCGTGGTTAGGGATGGCACCAACGTCTCCTACAGCTGGACTGCCTGGAG 5220 Q L Q A V V R D G T N V S Y S W T A W R 1670 5221 GGACAGGGGCCCGGCCTGGCCGGCAGCGGCAAAGGCTTCTCGCTCACCGTGCTCGAGGC 5280 DRGPALAGSGKGFSLTVLEA 1690 5281 CGGCACCTACCATGTGCAGCTGCGGCCACCAACATGCTGGGCAGCGCCTGGGCCGACTG 5340 G T Y H V Q L R A T N M L G S A W A D C 1710 5341 CACCATGGACTTCGTGGAGCCTGTGGGGTGGCTGATGGTGACCGCCTCCCCGAACCCAGC 5400 T M D F V E P V G W L M V T A S P N P A 1730 5460 A V N T S V T L S A E L A G G S G V V Y 1731 1750 CACTTGGTCCTTGGAGGAGGGGCTGAGCTGGGAGACCTCCGAGCCATTTAGCACCCATAG 5520 T W S L E E G L S W E T S E P F T T H S. CTTCCCACACCCGGCCTGCACTTGGTCACCATGACGGCAGGGAACCCGCTGGGCTCAGC 5580 F P T P G L H L V T M T A G N P L G S A 1790 5581 CAACGCCACCGTGGAAGTGGATGTGCAGGTGCCTGTGAGTGGCCTCAGCATCAGGGCCAG 5640 1791 1810 CGAGCCCGGAGGCAGCTTCGTGGCGGCCGGGTCCTCTGTGCCCTTTTGGGGGGAGCTGGC 5700 E P G G S F V A A G S S V P F W G Q L A 1830 5701 CACGGGCACCAATGTGAGCTGGTGCTGGGCTGTGCCCGGCGGCAGCAGCAAGCGTGGCCC 5760 T G T N V S W C W A V P G G S S K R G P 5761 TCATGTCACCATGGTCTTCCCGGATGCTGGCACCTTCTCCATCCGGCTCAATGCCTGCAA 5820 H V T M V F P D A G T F S I R L N A S N 1870 CGCAGTCAGCTGGGTCTCAGCCACGTACAACCTCACGGCGGAGGAGCCCATCGTGGGCCT 5880 AVSWVSATYNLTAEEPIVGL 1890 GGTGCTGTGGGCCAGCAGCAAGGTGGTGGCGCCCGGGCAGCTGGTCCATTTTCAGATCCT 5940 V L W A S S K V V A P G Q L V H F Q I L 1910 5941 GCTGGCTGCCGGCTCAGCTGCACCTTCCGCCTGCAGGTCGGCGGGGCCAACCCCGAGGT 6000 1911 LAAGSAVTFRLQVGGANPEV

6001 1931	GCTCCCCGGGCCCCGTTTCTCCCACAGCTTCCCCCGCGTCGGAGACCACGTGGTGAGCGT L P G P R F S H S F P R V G D H V V S V	6060 1950
6061 1951	GCGGGGCAAAAACCACGTGAGCTGGGCCCAGGCGCAGGTGCGCATCGTGGTGCTGGAGGC R G K N H V S W A Q A Q V R I V V L E A	5120 1970
6121 1971	CGTGAGTGGGCTGCAGATGCCCAACTGCTGCGAGCCTGGCATCGCCACGGGCACTGAGAG V S G L Q M P N C C E P G I A T G T E R	6180 1990
6181 1991	GAACTTCACAGCCCGCGTGCAGCGCGGGCTCTCGGGTCGCCTACGCCTGGTACTTCTCGCT N F T A R V Q R G S R V A Y A W Y F S L *	6240 2010
6241 2011	GCAGAAGGTCCAGGGCGACTCGCTGGTCATCCTGTCGGGCCGCGACGTCACCTACACGCC Q K V Q G D S L V I L S G R D V T Y T P	6300 2030
6301 2031	CGTGGCCCCGGGGCTGTTGGAGATCCAGGTGCGCCCTTCAACGCCCTGGGCAGTGAGAA V A A G L L E I Q V R A F N A L G S E N *	6360 2050
6361 2051	CCGCACGCTGGTGCTGGAGGTTCAGGACGCCGTCCAGTATGTGGCCCTGCAGAGCGGCCCRTLVLVLEVQDAVQVVALQSGP	6420 2070
6421 2071	CTGCTTCACCAACCGCTCGGCGCAGTTTGAGGCCGCCACCAGCCCCAGCCCCCGGCGTGT C F T N R S A Q F E A A T S P S P R R V	6480 2090
6481 2091	GGCCTACCACTGGGACTTTGGGGATGGGTCGCCAGGGCAGGACACAGATGAGCCCAGGGCAGGCCAGGGCAGGACACAGATGAGCCCAGGGCAGGCA	6540 2110
6541 2111	CGAGCACTCCTACCTGAGGCCTGGGGACTACCGCGTGCAGGTGAACGCCTCCAACCTGGT E H S Y L R P G D Y R V Q V N A S N L V	6600 2130
6601 2131	GAGCTTCTTCGTGGCGCAGGCCACGGTGACCGTCCAGGTGCTGGCCTGCCGGGAGCCGGA S F F V A Q A T V T V Q V L A C R E P E	6660 2150
6661 2151	GGTGGACGTGGTCCTGCCCCTGCAGGTGCTGATGCGGCGATCACAGCGCAACTACTTGGA V D V V L P L Q V L M R R S Q R N Y L E	6720 2170
6721 2171	GGCCCACGTTGACCTGCGCGACTGCGTCACCTACCAGACTGAGTACCGCTGGGAGGTGTA A H V D L R D C V T Y Q T E Y R W E V Y	6780 2190
6781 2191	TCGCACCGCCAGCGCCCAGCGCGCGTGTGGCCCTGCCCGGCGTGGART ASCQR PGRPARVALPGVD	6840 .2210
6841 2211	CGTGAGCCGGCCTCGGCTGCTGCCGCGCTGCCTGTGGGGCACTACTGCTTVSRPRCTCTGCTGCGCGCTGCCTGTGGGGCACTACTGCTT	6900 2230
6901 2231	TGTGTTTGTCGTGTCATTTGGGGACACGCCACTGACACAGAGCATCCAGGCCAATGTGAC V F V V S F G D T P L T Q S I Q A N V T	6960 2250
	GGTGGCCCCGAGCGCCTGGTGCCCATCATTGAGGGTGGCTCATACCGCGTGTGGTCAGA V A P E R L V P I I E G G S Y R V W S D	7020 2270
7021 2271	CACACGGGACCTGGTGCTGGATGGGAGCGAGTCCTACGACCCCAACCTGGAGGACGGCGA T R D L V L D G S E S Y D P N L E D G D	7080 2290
7081 2291	CCAGACGCCGCTCAGTTTCCACTGGGCCTGTGTGGCTTCGACACAGAGGGAGG	7140 2310
	GTGTGCGCTGAACTTTGGGCCCCGCGGGAGCAGCACGGTCACCATTCCACGGGAGCGGCT C A L N F G P R G S S T V T I P R E R L	7200 2330